



International Journal of PharmTech Research CODEN (USA): IJPRIF ISSN : 0974-4304 Vol. 3, No.1, pp 50-57, Jan-Mar 2011

Proteomics in Computer-assisted Molecular Design

Dubey R D^{1*}, Paroha S¹, Wani V K¹, Pandey A K¹, Verma S¹, Daharwal S J¹,

Dewangan D², Prasad Reddy S L N³

¹Institute of Pharmacy, Chhatauna, Mandir Hasaud, RITEE, Raipur, Chhattisgarh, India.

²Rungta College of Pharmaceutical Science and Research, Bhilai, Chhattisgarh, India.

³Samskruti College of Pharmacy, Kondapur, Ghatkesar, RR Dist, Hyderabad, Andhra Pradesh, India.

*Corres. author: dubey_pharma2000@yahoo.com, Mob. No. 09039150435

Abstract: Computer-assisted molecular design (CAMD) also called Computer-assisted drug design (CADD) represents more recent applications of computers as tools in the drug design process. The proteome is the entire protein complement expressed by a genome, and proteomics is the study of the proteome. Proteomics is a research field that involves large scale identification, characterization and quantitation of proteins expressed in a cell, tissue, or organism under given conditions. The ultimate goal of proteomic analysis is a comprehensive and quantitative description of protein expression and quantitative description of protein expression and alterations associated with biological perturbations under a given condition. In most current applications of CADD, attempts are made to find a ligand that will interact favorably with a receptor that represents the target site. Binding of ligand to the receptor may include hydrophobic, electrostatic, and hydrogen-bonding interactions. Computational assessment of the binding affinity of enzyme inhibitors prior to synthesis is an important component of CADD paradigms. **Key words:** CADD, Proteome, Ligand, Genome.

1. INTRUDUCTION

Although no single drug has been designed solely by computer techniques, the contribution of these methods to drug discovery is no longer a matter of dispute. All the world's major pharmaceutical and biotechnology companies use computational design tools. At their lowest level the contributions represent the replacement of crude mechanical models by displays of structure which are a much more accurate reflection molecular reality, capable of of demonstrating motion and solvent effects. Beyond this, theoretical calculations permit the computation of binding free energies and other relevant molecular properties.

This process extensively uses mathematical models and simulation tools based on the evaluation of potential risks from drug safety and the experimental design of new trials [1-3]. The ability to rapidly and accurately dock large numbers of candidate molecules into the binding site of a target macromolecule is a key component of lead generation in structure-based drug design [4, 5]. The most widely used computational docking method is the program DOCK [6] which has been and continues to be developed by Kuntz and his colleagues at the University of California and other scientists worldwide [6-9]. The success application of DOCK includes the in silico virtual high throughput screen for high affinity cytochrome p450cam substrates [10] and the computer-assisted design of selective imidazole inhibitors for cytochrome p450 enzymes [11]. Besides DOCK, numerous other programs have been created for virtual screening. Programs such as ADAM [12, 13], AutoDOCK [14-16], FlexX [10, 17-19], and SLIDE [20-22], and other dock databases of compounds can score candidate molecules according to their interactions with the selected site of target protein. De novo generation of ligands can be performed with computer programs including 3D-OSAR [23-25], DISCO [26], GRID [27-30], LUDI [31-33], MCSS [27, 34], and PASSA [35]. With the rapid accumulation of biological and chemical information, CADD has been dramatically reshaping research and development pathways in drug candidate identification. On the other hand, the escalating number of therapeutic candidates is increasing demand on new technologies and strategies to streamline the process of screening for safe and effective therapies. As an emerging technology, CADD accelerates drug development by making use of the accumulated information of existing drugs and diseases, combined with inter-disciplinary inputs from other fields. In this review article, we aim to briefly summarize the recent progresses in pharmacoproteomics and their potential application in CADD.

2. PROTEOMICS

The proteome is defined as the entirely expressed protein complement of a cell, organ or organism and it includes all isoforms and post-translational variants. The proteome is the entire protein complement expressed by a genome, and proteomics is the study of the proteome [36, 37]. Proteomics is a research field that involves largescale identification, characterization, and quantitation of proteins expressed in a cell, tissue, or organism under given conditions such as drug treatment [38-40]. Proteomic technology attempts to separate, identify and characterize a global set of proteins in an effort to provide information about protein abundance, location, modification and proteinprotein interaction in a proteome of a given biological system [41, 42].

By studying the interrelationships of protein expression and modification in health and disease, or drug treatment, proteomics can be applied to biomarker discovery and drug target validation [42-44]. The ultimate goal of proteomic analysis is a comprehensive and quantitative description of protein expression and alterations associated with biological perturbations under a given condition. By studying interrelationships of protein expression and modification in health and disease or drug treatment, proteomics contributes important insights into determining the pathophysiological basis of disease [45], validating drug targets [46], and illustrating drug action [47], toxicity and side effects [48].

3. PROTEOMICS IN THE MULTI-STEP PROCESS OF DRUG DISCOVERY

Drugs exert their actions mainly by targeting functional proteins. Therefore, it appears straightforward to focus on proteins in order to investigate drug effects. Unfortunately, it is not easy to screen for protein alterations because of their high complexity. Traditional methods such as NMR analysis [49, 50] or yeast two hybrid systems [51, 52] for mapping protein-protein interactions are laborious and cannot meet the need for large scale analysis. Recently developed proteomic approaches have dramatically increased the efficiency and applicability of mapping drug-protein and protein-protein interactions. Proteomics can provide valuable information for drug discovery including target identification and validation [53, 54], lead selection [55], small-molecular screening and optimization [56, 57], and toxicity testing [58, 59]. The opportunities offered by proteomics are not limited to a list of proteins. Instead, the scope of proteomics covers the analysis of protein cellular activities and functions, including the characterization of the flow of information within the cell.

4. MAJOR TECHNOLOGICAL PLATFORMS FOR PROTEOMICS

A number of complementary technologies have been developed to analyze proteomes in a global scale. Currently, the most commonly used proteomic platforms include two-dimensional gel electrophoresis (2DE), protein chip arrays and liquid chromatography, incorporated with matrix-assisted laser desorption/ionization time of flight (MALDI-TOF), surface enhanced laser desorption ionization time of flight (SELDI-TOF) and/or tandem mass spectrometry (MS/MS). Liquid chromatography/tandem mass spectrometry (LC-MS/MS) is an analytical method for identifying multiple components of a protein mixture [60]. The peptide mixtures in very complex protein samples are physically resolved by chromatographic separation prior to injection into the mass spectrometer to generate a more informative map, consisting of both the unique elution characteristics (column retention times) as well as m/z ratios of individual peptides [61]. LC-MS/MS is well-suited to examine complex protein samples, since peptides with the same nominal m/z are less likely to be introduced to MS/MS at the same time, and fewer artifacts arise due to ion suppression or ion-ion interference. LC-MS/MS can also overcome the difficulties of 2DE in the identification of very large and basic proteins by pre-fractionation using 1DE.

5. SUB-DISCIPLINES OF PROTEOMICS IN COMPUTER-AIDED DRUG DESIGN

There are a series of sub-disciplines of proteomic technologies, including chemical proteomics, computational proteomics, structural proteomics, and topological proteomics, are taking part drug design research fields.

5.1 Chemical Proteomics

It describes chemical nature of protein. Chemical proteomics makes use of synthetic organic chemistry, cell biology, biochemistry, and mass spectrometry to design specific protein-modifying reagents that can be used for functional studies of distinct proteins within a certain proteome [62]. The most important tool of this field is carefully designed chemical probes that can specifically target diverse sets of enzyme families. A chemical probe contains three parts, a reactive ligand that can covalently bind to the target protein/enzyme, a linker region modulating the reactivity and specificity of the reactive ligand, and a tag for identification and purification of the target protein/enzyme [62, 63].

5.2 Computational Proteomics

It is very wider and most important part. Computational proteomics refers to the large-scale generation and analysis of 3D protein structural information [64]. Accurate prediction of protein contact maps is the beginning and essential step for computational proteomics. (Table-01) provide a broad range of structural and functional annotations for proteins from sequenced genomes and protein 3D structures, which make a solid foundation for computational proteomics.

5.3 Structural Proteomics

Describe structure of a protein. Structural proteomics is the determination of the relationship of all the

proteins or protein complexes in a specific cellular organelle and the establishment of the relationship of these proteins in a proteome-wide scale. Combining structural biology with computational and medicinal chemistry, structural proteomics can help design drugs effectively. The major goal of structural proteomics is to determine the 3D structures of as many as possible proteins, so that other proteins in an organelle can be computationally modeled on the basis of similarity of their amino acid sequences [65, 66].

5.4 Topological Proteomics

Topological proteomics aims at localizing and characterizing entire protein networks within a single cell, providing quantitative insights into their basic organization, which are valuable information in identifying new drug targets and selecting potential lead compounds [67, 68]. The proprietary technology, Multi-Epitope-Ligan Kartographie (MELK), is an ultra-sensitive topological proteomics technology for analyzing proteins on a single cell level. MELK can trace out large scale subcellular protein patterns simultaneously within a cell, hence unravelling hierarchies of proteins related to a particular cell function or dysfunction [69]. Another topological proteomic program, TopNet, is an automated web tool designed to facilitate the analysis of interaction networks, which is available from TopNet [70] (Table-01).

 Table-01: Useful Websites in Proteomics and Computer-Aided Drug Design

Websites	Database Description
http://www.ebi.ac.uk/dali/	Network service for comparing protein structures in 3D
http://www.rcsb.org/pdb/	Protein Data Bank
http://www.tops.leeds.ac.uk/	Topology of Protein Structure
http://www.biochem.ucl.ac.uk/bsm/PROCAT/PROCAT.html	PROCAT 3D enzyme active site templates
http://www.uhnres.utoronto.ca/proteomics/	Ontario Center for Structural Proteomics
http://cl.sdsc.edu/ce.html	Databases and Tools for 3-D Protein Structure
	Comparison and Alignment
http://www.protein.bio.msu.su/issd/	Integrated Sequence—Structure Database
http://scop.mrc-lmb.cam.ac.uk/scop/	Structural Classification of Proteins
http://networks.gersteinlab.org/genome	TopNet for Topological Proteomics
http://www.blueprint.org/bind/bind.php	Biolmolecular interaction network database
http://www.embl-	Protein sequence analysis and structure prediction
heidelberg.de/predictprotein/predictprotein.html	
http://www.ecoli-york.org/	Database for Escherichia coli.
http://www.genome.ad.jp/kegg/metabolism.html	Molecular interaction networks, includingmetabolic and
	regulatory pathways, andmolecular complexes
http://geneontology.org/	Controlled vocabulary describing molecular function,
	biological process, and cellular component.
http://www.broad.mit.edu/cgi-bin/cancer/datasets.cgi/	Repository of microarray data from cancer genomics
	publications

6. CHALLENGES IN PROTEOMIC APPROACHES

Proteomics provides a large number of validated targets for drug design and thus optimal methods have to be created to handle this challenge. This high dimensionality of data generated from these studies requires the development of advanced bioinformatics tools for efficient and accurate data analyses. For proteome profiling of a particular system or organism, a number of specialized software tools and advanced informatics are needed to support the analysis and management of these massive amounts of data. The rapidly emerging field of bioinformatics has the capacity to greatly enhance treatment efforts by serving as a bridge between proteomic raw data and applicable output [71, 72, 73]. By correlating genetic variation and potential changes in protein structure with clinical risk factors, disease presentation and differential response to treatment and drug candidates, it may be possible to obtain valuable new insights to support and guide rational decision-making, both at the clinical and public health levels. Application of this emerging integrated technology in drug development can be divided into three categories: target discovery and validation, illustration of efficacy and toxicity of compounds and identification or prediction of drug response.

7. CURRENT ACHIEVEMENTS AND APPLICATION OF PROTEOMICS IN COMPUTER AIDED DRUG DISIGN

Biomarker (Proteomic Signature) Discovery

Biomarkers are usually proteins that have their expression altered in response to a disease condition. Biomarkers can be used as signatures to determine drug efficacy and clinical effects. Since the introduction of proteomics technology, 2DE, protein chip arrays together with mass spectrometry have been extensively used in biomarker discovery. Biomarkers can provide a basis for the selection of lead candidates for clinical trials and for the understanding of candidate's pharmacology. They can also help in the characterization of the subtypes of diseases for which a therapeutic intervention is most appropriate.

7.1 Action Mechanisms of Drugs

Drugs exert their functions mainly by affecting on proteins. Therefore, it seems straightforward to focus on proteins in order to investigate the effects of drugs. Unfortunately, proteins are of very high complexity, making it much more difficult to screen for protein alterations than gene regulation. However, the efficiency and applicability of proteome analysis have been dramatically increased recently. Investigation of altered protein expression in response to drug treatment in established model systems is becoming a commonly used strategy to examine drug action mechanisms.

7.2 Molecular Drug Target

Target identification and validation are the first key steps in the drug discovery pipeline. Reliable technologies for addressing target identification and validation are the foundation of successful drug development. Proteomics has been well utilized in protein expression profiling and tissue/cell-scale target validation.

7.3 Drug Toxicity and Side Effects

Given the low success rate in drug development, detection of potential toxicity and side effects in early stages of drug candidate identification can save money and time by focusing resources on those safe drug leads and candidates. By establishing a database that defines the response of a tissue proteome to specific drugs, comparative proteomics can be used to determine the propensity for a new compound. Proteomic signatures can also be constructed based on the toxicity responses previously observed with known agents. This can provide information to screen similar compounds for modification and improvement in drug design.

7.4 Cellular Signaling Network Reconstruction

It is becoming increasingly clear that proteins perform their functions concurrently in complex networks. The rapid accumulation in genomics and proteomics information and the development of large-scale experimental techniques motivate us to develop computational approaches to dissecting different and complex signaling pathways and interactions within them.

8. CONCLUSIONS AND FUTURE PROSPECTS

Proteomic technology has progressed substantially from the simple concept of 2DE into a series of technologies capable of investigating the total protein content of a biological system and its response to changing conditions. This technology has revolutionized the way in which researchers analyze the presence and relative abundance of proteins and expedite the screening and validation process for drug discovery. Proteomics applications in drug discovery in recent years have demonstrated the potential value of proteomics in drug development. Proteomic approaches can provide valuable information for target identification and validation, lead selection, smallmolecular screening and optimization.

REFERENCES

- Carlson H. A. and McCammon J. A., Accommodating Protein Flexibility in Computational Drug Design, Mol. Pharmacol., 2000, 57, 213-218.
- 2. Stret A. G. and Mayo S. L., Consolidating critical binding determinants by noncyclic rearrangement of protein secondary structure, Structure Fold Des., 1999, 7, 105-109.
- Veselovsky A. V. and Ivanov A. S., Strategy of computer-aided drug design, Curr. Drug Targets Infect. Disord., 2003, 3, 33-40.
- Kuntz I. D., Structure-Based Strategies for Drug Design and Discovery, Science, 1992, 257, 1078-1082.
- Good A. C., Ewing T. J., Gschwend D. A., and Kuntz I. D., New molecular shape descriptors: application in database screening, J. Comput. Aided Mol. Des., 1995, 9, 1-12.
- Kuntz I. D., Blaney J. M., Oatley S. J., Langridge R. and Ferrin T. E., A geometric approach to macromolecule-ligand interactions, J. Mol. Biol., 1982, 161, 269-288.
- Erickson J. A., Jalaie M., Robertson D. H., Lewis R. A. and Vieth M., Lessons in molecular recognition: the effects of ligand and protein flexibility on molecular docking accuracy, J. Med. Chem., 2004, 47, 45-55.
- 8. Schafferhans A. and Klebe G., Docking ligands onto binding site representations derived from proteins built by homology modeling, J. Mol. Biol., 2001, 307, 407-427.
- Yamamoto Y., Ishihara Y., and Kuntz I. D., Docking analysis of a series of benzylamino acetylcholinesterase inhibitors with a phthalimide, benzoyl, or indanone moiety, J. Med. Chem., 1994, 37, 3141-3153.
- Keseru G. M. A., virtual high throughput screen for high affinity cytochrome P450 molecule databases, J. Comput. Aided Mol. Des., 2001, 15, 649-657.
- Verras A., Kuntz I. D., and Ortiz de Montellano P. R., Characterizing Metabolic Inhibition Using Electrochemical Enzyme/DNA Biosensors, J. Med. Chem., 2004, 47, 3572-3579.
- Adam B. L., Vlahou A., Semmes O. J., and Wright G. L., Biomarker selection, employing an iterative peak selection method, and prostate spectra characterization for identifying biomarkers related to prostate cancer, Jr. Proteomics, 2001, 1, 1264-1270.

- Mizutani M. Y., Tomioka N. and Itai A., Protein kinase C modulators bearing dicarbacloso-dodecaborane as a hydrophobic pharmacophore, J. Mol. Biol., 1994, 243, 310-326.
- Buzko O. V., Bishop A. C. and Shokat K. M., Modified AutoDock for accurate docking of protein kinase inhibitors, J. Comput. Aided Mol. Des., 2002, 16, 113-127.
- 15. Goodsell D. S., Morris G. M. and Olson A. J. Automated docking of flexible ligands: applications of AutoDock, J. Mol. Recognit., 1996, 9, 1-5.
- Osterberg F., Morris G. M., Sanner M. F., Olson A. J. and Goodsell D. S., Automated docking to multiple target structures: incorporation of protein mobility and structural water heterogeneity in AutoDock, Proteins, 2002, 46, 34-40.
- Hindle S. A., Rarey M., Buning C. and Lengaue T. J., Flexible Docking under Pharmacophore Type Constraints, Comput. Aided Mol. Des., 2002, 16, 129-149.
- Knegtel R. M., Bayada D. M., Engh R. A., von der Saal W., van Geerestein V. J. and Grootenhuis P. D., Comparison of two implementations of the incremental construction algorithm in flexible docking of thrombin inhibitors, J. Comput. Aided Mol. Des., 1999, 13, 167-183.
- 19. Vigers G. P., and Rizzi J. P., Multiple active site corrections for docking and virtual screening, J. Med. Chem., 2004, 47, 80-89.
- Hawkins C. A., Watson C., Yan Y., Gong B., Wemmer D. E., Modes of a Rationally Designed Photoactivated DNA Nuclease Determined by NMR, Nucleic Acids Res., 2001, 29, 936-942.
- 21. Marzilli L. G., Saad J. S., Kuklenyik Z., Keating K. A. and Xu Y., NMR and X-ray Structural Characterization of a Cisplatin Analogue Able To Slow Down the Pt–N7 Rotation of a Coordinated Guanine Base by a Billion-Fold Times: 2,2'-Bipiperidine(dimethylmalonato)platinum(II) Complex, J. Am. Chem. Soc., 2001, 123, 2764-2770.
- 22. Sinay P., Synthetic chemistry: Sugars slide into heparin activity, Nature, 1999, 398, 377-378.
- 23. Jozwiak K., Ravich S., Collins J. R. and Wainer I. W., Proteomics in Computer-Aided Drug Design, J. Med. Chem., 2004, 47, 4008-4021.

- Purushottamachar P., Kulkarni V. M., CoMFA on the melanogenesis inhibitory activity of alkyl-3,4-dihydroxybenzoate, N-alkyl-3,4dihydroxybenzamide analogues, and prediction of higher active compounds, Bioorg. Med. Chem., 2003, 11, 3487-3497.
- Santos-Filho O. A., Mishra R. K. and Hopfinger A. J., Interpretable correlation descriptors for quantitative structure-activity relationships, J. Comput. Aided Mol. Des., 2001, 15, 787-810.
- 26. Myers A. M., Charifson P. S., Owens C. E., Kula N. S., McPhail A. T., Baldessarini R. J., Booth R. G. and Wyrick S. D., MDF – A New QSPR/QSAR Molecular Descriptors Family, J. Med. Chem., 1994, 37, 4109-4117.
- 27. Bitetti-Putzer R., Joseph-McCarthy D., and Hogle J. M, Karplus M Functional group placement in protein binding sites: a comparison of GRID and MCSS, J. Comput. Aided Mol. Des., 2001, 15, 935-960.
- Cruciani G., and Watson K. A., An Innovative Application of the "Flexible" GRID/PCA Computational Method, J. Med. Chem., 1994, 37, 2589-2601.
- 29. Polanski J., Gieleciak R., Magdziarz T. and Bak A., Comparative molecular field analysis using GRID force field and GOLPE variable selection method in a study of inhibitors of glycogenphosphorylase, J. Chem. Inf. Comput. Sci., 2004, 44, 1423-1435.
- Tomioka N., Itai A. and Iitaka Y. A., method for fast energy estimation and visualization of protein-ligand interaction, J. Comput. Aided Mol. Des., 1987, 1, 197-210.
- 31. Bogacewicz R., Trylska J. and Geller M., Proteomics in Computer-Aided Drug Design, Acta. Pol. Pharm., 2000, 57, 25-28.
- Bohm H. J., Proteomics in Computer-Aided Drug Design, J. Commut. Aided Mol. Des., 1992, 6, 593-606.
- 33. Wolf K. and Dormeyer M., Information-based methods in the development of an- tiparasitic drugs., Parasitol. Res., 2003, 90, 91-96.
- Joseph-McCarthy D., Hogle J. M. and Karplus M., Use of multiple copy simultaneous search (MCSS) mehthod to design a newclass of picorna viruscapsid binding drugs, Proteins, 1997, 29, 32-58.
- 35. Tondel K., Anderssen E. and Drablos F. A., framework for significance analysis of gene expression data using dimension reduction methods, J. Comput. Aided Mol. Des., 2002, 16, 831-840.

- 36. Wasinger V. C., Cordwell S. J., Cerpa-Poljak A., Yan J. X., Gooley A. A., Wilkins M. R. Duncan M. W., Harris R., Williams K. L. and Humphery-Smith I., Progress with gene product mapping of the mollicutes:Mycoplasma genitalium, Electrophoresis, 1995, 16, 1090-1094.
- 37. Wilkins M. R., Sanchez J. C., Gooley A. A., Appel R. D., Humphery-Smith I., Hochstrasser D. F. and Williams K. L., Strategic Importance of Research Support through Pathology, Biotechnol. Genet. Eng. Rev., 1996, 13, 19-50.
- He Q. Y., Chiu J. F., Proteomics in biomarker discovery and drug development, J. Cell. Biochem., 2003, 89, 868-886.
- Pandey A. and Mann M., Proteomics to study genes and genomes, Nature, 2000, 405, 837-846.
- Tyers M. and Mann M., Proteomic analysis of post-translational modifications, Nature, 2003, 422, 193-197.
- 41. He Q. Y. and Chiu J. F., Silencing USP22 by asymmetric structure of interfering RNA inhibits proliferation and induces cell cycle arrest in bladder cancer cells, J. Cell Biochem., 2003, 89, 868-86.
- 42. Stoughton R. B., Friend S. H., How molecular profiling could revolutionize drug discovery, Nat Rev Drug Discov, 2005, 4, 345-50.
- 43. Wang Y., Chiu J. F. and He Q. Y., Proteomics in computer-aided drug design, Current Computer Aided Drug Design, 2005, 1, 43-52.
- 44. Fliser D., Wittke S. and Mischak H., Capillary electrophoresis coupled to mass spectrometry for clinical diagnostic purposes, Electrophoresis, 2005, 26, 2708-2716.
- 45. Hanash S., Capillary electrophoresis coupled to mass spectrometry for clinical diagnostic purposes, Nature, 2003, 422, 226-232.
- 46. Rosamond J. and Allsop A., Harnessing the Power of the Genome in the Search for New Antibiotics, Science, 2000, 287, 1973-1976.
- 47. Graves P. R., Kwiek J. J., Fadden P., Ray R., Hardeman K, Coley A. M., Foley M. and Haystead T. A., Discovery of Novel Targets of Quinoline Drugs in the human purine binding proteome, Mol. Pharmacol., 2002, 62, 1364-1372.
- 48. Venkatraman A., Landar A., Davis A. J., Chamlee L., Sanderson T., Kim H., Page G., Pompilius M., Ballinger S., Darley-Usmar V. and Bailey S. M., Modification of the mitochondrial proteome in response to the

stress of ethanol-dependent hepatotoxicity, J. Biol. Chem., 2004, 279, 22092-22101.

- 49. Fleming K. G. and Engelman D. M., Determination of Membrane Protein Molecular Weight Using Sedimentation Equilibrium Analytical Ultracentrifugation, Proc. Natl. Acad. Sci. USA, 2001, 98, 14340-14344.
- 50. [50] Page M. J., Amess B., Townsend R. R., Parekh R., Herath A., Brusten L., Zvelebil M. J., Stein R. C., Waterfield M. D., Davies S. C., and Hare M., Proteomic definition of normal human luminal and myoepithelial breast cells purified from reduction mammoplasties, J. Proc. Natl. Acad. Sci. USA, 1999, 96, 12589-12594.
- 51. Huang J. and Schreiber S. L., A yeast genetic system for selecting small molecule inhibitors of protein–protein interactions in□nanodroplets, Proc. Natl. Acad. Sci. USA, 1997, 94, 13396-13401.
- 52. Kang J., Kim T., Ko Y. G., Rho S. B., Park S. G., Kim M. J., Kwon H. J. and Kim S particulate aminoacyl-tRNA synthetases from rat liver, J. Biol. Chem., 2000, 275, 31682-31688.
- 53. Drummelsmith J., Brochu V., Girard I. and Messier N., Ouellette M Proteome Mapping of the Protozoan Parasite Leishmania and Application to the Study of Drug Targets and Resistance Mechanisms, Mol. Cell Proteomics, 2003, 2, 146-155.
- 54. Greenbaum D. C., Baruch A., Grainger M., Bozdech Z., Medzihradszky K. F., Engel J., DeRisi J., Holder A. A. and Bogyo M., A role for the protease falcipain 1 in host cell invasion by the human malaria parasite, Science, 2002, 298, 2002-2006.
- 55. Bleicher K. H., Bohm H. J., Muller K. and Alanine A. I., Hit and lead generation: beyond high-throughput screening, Nat. Rev. Drug Discov., 2003, 2, 369-378.
- Baker K., Bleczinski C., Lin H., Salazar-Jimenez G., Sengupta D., Krane S. and Cornish V. W., Chemical complementation: a reactionindependent genetic assay for enzyme catalysis, Proc. Natl. Acad. Sci. USA, 2002, 99, 16537-16542.
- 57. Kridel S. J., Axelrod F., Rozenkrantz N. and Smith J. W., Orlistat is a novel inhibitor of fatty acid synthase with antitumor activity, Cancer Res., 2004, 64, 2070-2075.
- 58. Imanishi S. and Harada K., Proteomics approach on microcystin binding proteins in

mouse liver for investigation of microcystin toxicity, Toxico., 2004, 43, 651-659.

- Keightley J. A., Shang L. and Kinter M., Proteomic Analysis of Oxidative Stressresistant Cells, Mol. Cell Proteomics, 2004, 3, 167-175.
- 60. Florens L., Washburn M. P., Raine J. D., A proteomic view of the Plasmodium falciparum life cycle, Nature, 2002, 419, 520-26.
- 61. Listgarten J. and Emili A., Statistical and computational methods for comparative proteomic profiling using liquid chromatography-tandem mass spectrometry, Mol Cell Proteomics 2005, 4, 419-34.
- Jeffery D. A. and Bogyo M., Chemical proteomics and its application to drug discovery, Curr. Opin. Biotech, 2003, 14, 87-95.
- 63. Adam G. C., Cravatt B. F. and Sorensen E. J., Profiling the specific reactivity of the proteome with non-directed activity-based probes, Chem. Biol., 2001, 8, 81-95.
- 64. Maggio E. T. and Ramnarayan K., Recent developments in computational proteomics, Drug Discov. Today, 2001, 6, 996-1004.
- 65. Renfrey S. and Featherstone J., Structural proteomics. Nat. Rev. Drug Discov., 2002, 1, 175-176.
- 66. Sali A., Glaeser R., Earnest T. and Baumeister W., From Words to Literature in Structural Proteomics, Nature, 2003, 422, 216-225.
- Linding R., Jensen L. J., Diella F., Bork P., Gibson T. J. and Russell R. B., Protein disorder prediction: implications for structural proteomics. Structure (Camb.), 2003, 11, 1453-1459.
- Amaral L. A., Scala A., Barthelemy M., Stanley H. E., Classes of small-world networks Proc. Natl. Acad. Sci. USA, 2000, 97, 11149-11152.
- 69. Girvan M. and Newman M. E., Community structure in social and biological networks, Proc. Natl. Acad. Sci. USA, 2002, 99, 7821-7826.
- Schubert W., Topological proteomics, toponomics, MELK-technology, Adv. Biochem. Eng. Biotechnol., 2003, 83, 189-209.
- Stanislaus R., Chen C., Franklin J., Arthur J., and Almeida J. S., AGML Central: web based gel proteomic infrastructure, Bioinformatics, 2005, 21, 1754-1757.
- 72. O'Riordan E., Orlova T. N. and Mei J. J., Proteomic-Based Detection of Urine Proteins

Associated with Acute Renal Allograft Rejection, J. Am Soc Nephrol, 2004, 15, 3240-3248.

73. Zhang X., Leung S. M., Morris C. R. and Shigenaga M. K., Evaluation of a novel, integrated approach using functionalized magnetic beads, bench-top MALDI-TOF-MS with prestructured sample supports, and pattern recognition software for profiling potential biomarkers in human plasma, J. Biomol. Tech., 2004, 15, 167-75.
