



Antioxidant activity of dried and rehydrated *Kappaphycus alvarezii* from Langkawi, Kedah and Semporna, Sabah

**Norhidayu Mohamed¹, Aminah Abdullah², Sahilah Abd. Mutalib^{1*},
Aishah Elias¹, Rul Aisyah Mat Repin¹**

¹School of Chemical Sciences and Food Technology, Faculty of Science and Technology Universiti Kebangsaan Malaysia (UKM), 43600 UKM Bangi, Selangor, Malaysia.

²Natural Medicine Research Centre, Universiti Islam Malaysia, Blok I, Bangunan MKN Embassy Techzone, Jalan Teknokrat 2, 63000 Cyberjaya, Selangor Darul Ehsan, Malaysia.

Abstract : The aim of this study was to determine the antioxidant activity of dried and rehydrated *Kappaphycus alvarezii* from Langkawi, Kedah and Semporna, Sabah. The antioxidant activity in *K. alvarezii* was determined by using total phenolic content (TPC), 2,2-diphenyl-1-picrylhydrazyl (DPPH) scavenging assay, Trolox equivalent antioxidant capacity (TEAC), Ferric reducing antioxidant power (FRAP) and oxygen radical absorbance capacity (ORAC). Comparison between dried and rehydrated of *K. alvarezii* indicated that the rehydrated was significantly ($p < 0.05$) high in antioxidant activity compared to dried. While, the Langkawi's rehydrated seaweed (119.3 mg GAE/100 g of dried weight (DW) sample) has significantly ($p < 0.05$) higher TPC value compared to Semporna (89.2 mg GAE/100 g of DW sample). The DPPH value of rehydrated seaweed from Semporna was 34.6% and significantly ($p < 0.05$) higher compared to seaweed from Langkawi which was 27.5% in value. Antioxidant activity for FRAP and ABTS assay of rehydrated seaweed from both locations showed no significant different ($p > 0.05$) value. While, the Langkawi's rehydrated seaweed (73 $\mu\text{mol TE}/100$ g of DW sample) showed significantly ($p < 0.05$) higher ORAC value compared to Semporna (53.8 $\mu\text{mol TE}/100$ g of DW sample). Thus, the finding in this study, demonstrated that rehydrated *K. alvarezii* possesses higher antioxidant activity compared to dried seaweed.

Keywords : Total phenolic content (TPC), antioxidant activity, seaweed, *Kappaphycus alvarezii*.

Introduction

In Malaysia, *Kappaphycus alvarezii* is the most common species that were cultivated for food as well as for carrageenan production, where almost all of them were cultivated in fishing community around Semporna, Sabah¹⁻². Due to high market value of *K. alvarezii*, the variety of seaweed from Semporna, Sabah was

transferred and planted to Langkawi, Kedah. Langkawi was chosen as the seaweed farming place due to environmental conditions such as water temperature and salinity level that are conducive for farming. Therefore, Langkawi was expected to transform the seaweed industry into a larger and more profitable scale.

Seaweed has low calorie content, high fibre and mineral content, and significant amount of protein, vitamins and trace elements³. Furthermore, seaweeds are capable to generate essential defence mechanisms against oxidation. Therefore seaweeds are important source of antioxidant that may able to protect human body against reactive oxygen species⁴. While, a lot of studies have reported about significant amount of antioxidant activity in the *K. alvarezii*⁵⁻⁶.

Normally, the dried seaweeds are rehydrated in water to restore their original structure and the excess water is removed before they can be used⁷⁻⁸. During the rehydration process, the dry porous material submerged in water undergoes several changes towards its moisture, porosity and volume⁹. However, the dehydration and rehydration process may affect the nutritional value of *K. Alvarezii*, by causing changes in chemical composition as well as bioactivities such as antioxidant activity. Therefore, in this study we determined the effects of rehydration on antioxidant activity using *K. alvarezii* from Langkawi, Kedah and Semporna, Sabah.

Materials and Methods

Sample preparation

Dried seaweed of *K. alvarezii* from Langkawi was supplied by Maya System Enterprise, Malaysia. While sample from Semporna was purchased from Sabah local supplier in November 2014. The samples were prepared as method described by¹⁰ with slight modification. The seaweeds were rinsed using distilled water. Then, they were rehydrated three times using distilled water in the ratio 1: 5. A total of 200 g seaweed was soaked into 1 L of distilled water in biker for 1 min at 28°C. After rehydrated, the water was discarded and seaweeds were dried for 15 mins to remove-excess water. The dried and rehydrated seaweeds were dried in the oven at 50°C for 72 hr. Finally, they were ground into powder form using universal cutting mill (Fritsch Industries, Germany) and was stored at 4°C until further analysis.

Antioxidant extraction

The extraction of antioxidant was done as described by¹¹ with slight modification. About 0.5 g of seaweed powder was mixed with 10 mL of 50% (v/v) acetone (HmbG Chemicals, German). Then, samples were homogenized using high speed homogenizer (T250, IKA, Germany) at 24,000 rpm for 1 min. After that, the samples were mixed using magnetic stirrer (Heidolph, MR3001, K, Germany) for 24 hrs at 1,000 rpm. All extracted samples were centrifuged for 10 mins at 1580 rcf. The supernatant was collected and stored at -20°C.

Determination of total phenolic content (TPC)

The determination of total phenolic content analysis was conducted according to the method by¹¹ with slight modification. 200 µL of seaweed extract was mixed with 0.5 mL diluted FC (Merck, Germany) reagent (1 mL FC reagent: 10 mL distilled water) and the mixture was left for 5 mins. Then, 0.4 mL of 7.5% (w/v) sodium carbonate (Sigma, Germany) was added to the mixture and the solutions were incubated in room temperature for 2 hrs. After that, the absorbance was taken at 765 nm wavelength using spectrophotometer (BMG Labtech, Germany). Standard calibration curve of gallic acid (Sigma, Germany) was set up to estimate the activity capacity of samples. TPC was expressed in terms of mg gallic acid equivalents per 100 g dried weight (DW) sample (mg GAE/100 g of DW sample).

Determination of the free radical scavenging activity

Scavenging activity determination of the seaweed extracts on the stable free radical DPPH were done as described by¹¹ with slight modification. The DPPH stock solution was prepared by dissolving 40 mg DPPH (Sigma, Germany) in 100 mL methanol (Merck, Germany). The DPPH working solution was freshly prepared by diluting the stock solution with methanol until the absorbance reached 0.7 ± 0.01 . Then, 300 uL extract or blank were mixed with 1 mL DPPH working solution and the mixture were incubated for 24 hrs at room

temperature. Absorbance of the mixture was measured using spectrophotometer (BMG Labtech, Germany) at 517 nm wavelength. The scavenging activity was determined using the following equation:

$$\text{DPPH scavenging activity} = \left[\frac{\text{Absorbance}_{\text{blank}} - \text{Absorbance}_{\text{sample}}}{\text{Absorbance}_{\text{blank}}} \right] \times 100$$

Determination of ferric reducing antioxidant power (FRAP)

The antioxidant capacity of each extract was measured according to procedure by¹¹ with some modification. FRAP reagent consists of 300 mM acetate buffer pH 3.6, 10 mM TPTZ (2,4,6-tri(2-pyridyl)-s-triazine) (Sigma Aldrich, USA) dissolved in 40 mM HCl (Merck, Germany), and 20 mM ferric chloride hexahydrate (FeCl₃.6H₂O) (Merck, Germany). All chemical were mixed in the ratio of 10: 1: 1 (300 mM acetate buffer: 10 mM TPTZ: 20 mM FeCl₃.6H₂O). After that, 1 mL FRAP reagent was mixed with 200 µL seaweed extract or standards solutions. The mixture was incubated for 30 mins at room temperature. Then, the absorbance of the mixture was measured using spectrophotometer (BMG Labtech, Germany) at 595 nm. Calibration curve of trolox (Sigma, Germany) was set up to estimate the antioxidant activity of samples. The result was interpreted in terms of micromol trolox equivalents per 100 g of dried weight (DW) sample (µmol TE/100 g of DW sample).

Determination of trolox equivalent antioxidant capacity (TEAC)

Antioxidant activities of seaweed extract was determined using TEAC method as described by⁶ with slight modification. The stock solution was prepared by mixing the 7 mM 2,2'-Azino-bis(3-ethylbenzothiazoline-6-sulfonic acid) diammonium salt (Sigma, Germany) with 2.45 mM potassium persulphate (J. Kollin, UK) in the ratio of 1:1. Next, the stock solution was incubated for 16 hrs at room temperature. The working solution was prepared by diluting stock solution with methanol (Merck, Germany) until the absorbance reached approximately 0.7 ± 0.01 at 734 nm. An aliquot (200 µL) of extracts or standard solutions were mixed with 1 mL of working solution and the mixture were incubated for 1 hr at room temperature. The absorbance of the solution was measured using spectrophotometer (BMG Labtech, Germany). The calibration curve of trolox (Sigma, Germany) was plotted. The result was interpreted in terms of micromol trolox equivalents per 100 g of dried weight (DW) sample (µmol TE/100 g of DW sample).

Determination of oxygen radical absorbance capacity

The determination of antioxidant capacity in seaweed was determined by using ORAC method as described by¹² with slight adjustment. 150 µL of fluorescein solution (Sigma Aldrich, USA).was pipetted in the black plate. Then, 25 µL trolox (Sigma, Germany), sample or blank was added, and the plate was incubated for 30 min at 37 °C in the microplate reader (POLARstar Omega, BMG Labtech, Germany). After incubation, fluorescence measurements were taken every 90 sec at the emission 520 nm and excitation 485 nm. At the fourth cycles, 25 µL of 240 mM 2,2'-azo-bis(2-amidinopropane) dihydrochloride (AAPH) (Sigma, Germany) was injected using the onboard injector. The test was resumed and fluorescence intensity was recorded up to 120 min. The calibration curve of trolox were plotted and the result was interpreted in terms of micromol trolox equivalents per 100 g of dried weight (DW) sample (µmol TE/100 g of DW sample).

Statistical analysis

All data were analysed statistically using Statistical Package for Social Science (SPSS) version 20 (IBM, California, USA) using independent T test and Pearson's correlation coefficient test. A significant difference was assumed at the level of p<0.05.

Results and Discussion

The effect of rehydrated procedure to the total phenolic content (TPC) of *K. alvarezii* from Langkawi, Kedah and Semporna, Sabah was shown in Table 1. As indicated in Table 1, comparing between two locations of Semporna and Langkawi, the TPC in rehydrated *K. alvarezii* was significantly (p<0.05) higher compared to dried *K. alvarezii* for both location. Our result is similar with¹³ who reported significant increase in total phenolic contents when dry common beans and pinto beans were soaked in cold water.

Table 1 Antioxidant activities of dried and rehydrated *Kappaphycus alvarezii* that were obtained from Langkawi, Kedah and Semporna, Sabah

Antioxidative activities	Isolation Location of <i>Kappaphycus alvarezii</i>	
	Semporna, Sabah	Langkawi, Kedah
TPC (mg GAE/100 g of DW sample)		
Dried seaweed	41.4 ± 0.7 ^{bA}	33.8 ± 2.3 ^{bB}
Rehydrated seaweed	89.2 ± 4.8 ^{aB}	119.3 ± 10.4 ^{aA}
DPPH (%)		
Dried seaweed	31.7 ± 0.3 ^{bA}	17.2 ± 0.2 ^{bB}
Rehydrated seaweed	34.6 ± 1.0 ^{aA}	27.5 ± 3.1 ^{aB}
FRAP (µmol TE/100 g of DW sample)		
Dried seaweed	13.4 ± 0.7 ^{bA}	9.4 ± 1.9 ^{bB}
Rehydrated seaweed	22.5 ± 0.1 ^{aA}	17.7 ± 2.0 ^{aA}
TEAC (µmol TE/100 g of DW of sample)		
Dried seaweed	90.4 ± 12.0 ^{bA}	112.2 ± 15.0 ^{bA}
Rehydrated seaweed	138.4 ± 4.1 ^{aA}	130.9 ± 7.0 ^{aA}
ORAC (µmol TE/100 g of DW sample)		
Dried seaweed	31.4 ± 1.1 ^{bB}	36.4 ± 6.9 ^{bA}
Rehydrated seaweed	53.8 ± 7.3 ^{aB}	73.0 ± 18.2 ^{aA}

DW: Dried weight; ± Standard deviation

^{A-B} Different capital letters at the same row shows significant differences ($p < 0.05$) between location of the seaweeds were taken.

^{a-b} Different small letters at the same column shows significant differences ($p < 0.05$) between dried seaweed and rehydrated seaweed.

In the present study, the TPC of rehydrated seaweed from Langkawi (119.3 mg GAE/100 g of DW sample) was significantly ($p < 0.05$) higher compared to rehydrated seaweed from Semporna (89.2 mg GAE/100 g of DW sample). The result obtained maybe was related to the environmental factor where *K. alvarezii* was harvested. The total phenolic compound can be affected by many factors such as season and agroclimatic conditions¹⁴. This finding was also supported by¹⁵ who reported *Sargassum muticum* collected from different countries had a significant effect on the TPC value. No report has been obtained for *K. alvarezii* however the similar reason reported by¹⁵ may explain in our result.

In Table 1 the DPPH scavenging percentages of rehydrated seaweeds (Semporna's seaweed; 34.6% and Langkawi's seaweed; 27.5%) are significantly ($P < 0.05$) higher compared to dried seaweed for both locations (Semporna's seaweed; 31.7% and Langkawi's seaweed; 17.2%). Our finding was consistent with¹⁶. Though this study was not for *K. alvarezii* but¹⁶ found that the radical scavenging activity of rehydrated *Himanthalia elongata* was higher than dried sample. Comparing between rehydrated seaweed from location, DPPH scavenging percentages of seaweed from Semporna (34.6%) was significantly ($p < 0.05$) higher compared to seaweed from Langkawi (27.5%).¹⁵ reported that the different of value DPPH radical scavenging activity of *Sargassum muticum* may due to this algae was collected from distantly geographical countries. This may explain why the different value DPPH radical scavenging activity between rehydrated *K. alvarezii* from Semporna, Sabah and Langkawi, Kedah.

For FRAP assay, rehydrated seaweed (Semporna; 22.5 µmol TE/100 g of DW sample and Langkawi; 17.7 µmol TE/100 g of DW sample) were significantly ($p < 0.05$) higher than dried seaweed (Semporna; 13.4 µmol TE/100 g of DW sample and Langkawi; 9.4 µmol TE/100 g of DW sample) for both location. This result was similar with¹⁷ who reported, the FRAP value of soaked beans were remained at 80% or higher compared to the raw beans. Similar assumption may explain the FRAP value in this study. There is no significant ($p > 0.05$) difference between rehydrated sample from Semporna and Langkawi for FRAP.

In TEAC analysis, as presented in Table 1, the rehydrated seaweed (Semporna's seaweed: 138.4 µmol TE/100 g of DW sample and Langkawi's seaweed: 130.9 µmol TE/100 of g DW sample) significantly ($p < 0.05$) has higher TEAC value compared to dried seaweed (Semporna's seaweed: 90.4 µmol TE/100 g of DW sample and Langkawi's seaweed: 112.3 TE/100 g of DW sample). There is no significant ($p > 0.05$) difference between rehydrated sample from Semporna and Langkawi for TEAC assay.

ORAC analysis value for rehydrated seaweeds (Semporna; 53.8 $\mu\text{mol TE}/100\text{ g}$ of DW sample and Langkawi; 73 $\mu\text{mol TE}/100\text{ g}$ of DW sample) were significantly ($p < 0.05$) higher compared to dried seaweed (Semporna; 31.4 $\mu\text{mol TE}/100\text{ g}$ of DW sample and Langkawi; 36.4 $\mu\text{mol TE}/100\text{ g}$ of DW sample) for both location. Similar observation was reported by ¹⁸. In their study they reported the ORAC value was significantly higher in rehydrated sea cucumber than in the fresh one with the internal organs included. Though the sample was not similar, this reason may also explain the value of ORAC in *K. alvarezii*. The ORAC analysis involves a hydrogen atom transfer (HAT) to peroxy radical which both, ¹⁸ and result in this finding may have similar mechanism.

Pearson correlation coefficient between antioxidant assays of dried and rehydrated seaweed were shown in Table 2. As indicated in Table 2, there was strong significant correlation between TPC with FRAP ($r = 0.728$, $p < 0.01$), TEAC ($r = 0.755$, $p < 0.01$), and ORAC ($r = 0.930$, $p < 0.01$) value. Previous studies also showed that there was positive correlation between TPC with FRAP¹⁹⁻²¹, TPC with TEAC²² and TPC with ORAC²³ value of different plant extracts. This result indicated there is a relation between phenolic concentrations with FRAP, ABTS radical scavenging (TEAC), and peroxy radical scavenging (ORAC) in seaweed extracts.

Table 2 Pearson's correlation coefficients between antioxidant assays of dried and rehydrated *Kappaphycus alvarezii* from Langkawi, Kedah and Semporna, Sabah.

Correlation coefficient (r)	TPC	DPPH	FRAP	TEAC	ORAC
TPC	1	0.433	0.728*	0.755*	0.930*
DPPH		1	0.751*	0.205	0.154
FRAP			1	0.650	0.515
TEAC				1	0.683
ORAC					1

* Correlation is significant at $p < 0.01$ level

The example of phenolic compound that are abundant in algae is phlorotannins²⁴, phenolic acid and flavonoid compound²⁵ which also were contribute to antioxidant activity of seaweed extract²⁶ (Jimenez et al. 2001). While, the phenolic content and radical scavenging activities in DPPH analysis did not show any significant correlation (Table 2). This result was consistent with²⁷ who reported that there was no significant correlation between TPC and DPPH of brown seaweed extract. This indicated that the seaweed extract contained other compound which could involve in antioxidant activity.

A significant ($p < 0.01$) strong positive correlation was also found between DPPH with FRAP assays ($r = 0.751$). The studied²⁸ reported similar significant correlation between DPPH with FRAP assay, in methanol extract of guava fruit. The correlation between DPPH and FRAP assay might be due to similar reaction of mechanism which is single electron transfer mechanism²⁹.

Conclusion

In conclusion, rehydrated seaweed possesses higher antioxidant activity compared to dried seaweed for both locations (Semporna Sabah and Langkawi, Kedah). The antioxidant activity in *K. alvarezii* was different according to location, except for FRAP and TEAC assay. Strong positive correlations between phenolic content and FRAP, TEAC, and ORAC value showed that, phenolic compounds were the contributors of antioxidant activity in *K. alvarezii*.

Acknowledgements

The authors would like to express their gratitude for grants and financial support from STGL-016-2012, DPP-2015-044 and IP-2014-011. Acknowledgement also goes to L2M Cycle 2 for providing the facilities for the antioxidant analysis.

References

1. Ahemad, S., Ismail, A. and Mohammad, R. M. A. The seaweed industry in Sabah, East Malaysia, *Jati.*, 2006, 11, 97–107.
2. Phang, S. M. Seaweed resources in Malaysia: Current status and future prospects, *Aquatic Ecosystem Health and Management.*, 2006, 9(2), 185–202.
3. Chang, V. S., Okechukwu, P. N. and Teo, S. S. The properties of red seaweed (*Kappaphycus alvarezii*) and its effect on mammary carcinogenesis, *Biomedicine and Pharmacotherapy.*, 2017, 87, 296–301.
4. Plaza, M., Cifuentes, A. and Ibanez, E. In the search of new functional food ingredients from algae, *Trends in Food Science and Technology.*, 2008, 19(1), 31-39.
5. Farah Diyana, A., Abdullah, A., Shahrul Hisham, Z. A. and Chan, K.M. Antioxidant activity of red algae *Kappaphycus alvarezii* and *Kappaphycus striatum*, *International Food Research Journal.*, 2015, 22(5), 1977–1984.
6. Matanjun, P., Mohamed, S., Mustapha, N. M., Muhammad, K. and Cheng, H. M. Antioxidant activities and phenolic content of eight species of seaweeds from north Borneo, *Journal of Applied Phycology.*, 2008, 20, 367–373.
7. Ledesma, B. H. and Herrero, M. Bioactive compounds from marine foods: Plant and animal sources, John Wiley and Sons Limited and the Institute of Food Technologists, Chicago, 2014.
8. Mouritsen, O. G. Seaweeds: Edible, Available and Sustainable, The University of Chicago Press, Chicago, 2013, 121.
9. Krokida, M. K. and Philippopoulos, C. Drying technology: An rehydration of dehydrated foods, *An International Journal.*, 2005, 23(4), 799 – 830.
10. Xiren, G. K. and Aminah, A. Elimination of seaweed odour and its effect on antioxidant activity, *AIP Conference Proceedings.*, 2014, 1614, 399.
11. Musa, K. H., Aminah, A., Khairiah, J. and Subramaniam, V. Antioxidant activity of pink flesh guava (*Psidium guajava* L.): Effect of extraction techniques and solvents, *Food Analytical Methods.*, 2010, 4(1), 100–170.
12. Cao, G., Alessio, H. M. and Cutler, R. G. Oxygen radical absorbance capacity assay for antioxidants, *Free Radical Biology and Medicine.*, 1993, 14, 303–311.
13. Akillioglu, H. G. and Karakaya, S. Changes in total phenols, total flavonoids, and antioxidant activities of common beans and pinto beans after soaking, cooking, and in vitro digestion process, *Food Science and Biotechnology.*, 2010, 19(3), 633–639.
14. Iqbal, S. and Bhangar, M. I. Effect of season and production location on antioxidant activity of *Moringa oleifera* leaves grown in Pakistan, *Journal of Food Composition and Analysis.*, 2006, 19(6–7), 544–551.
15. Tanniou, A., Vandanjon, L., Incera, M., Serrano Leon, E., Husa, V., Le Grand, J., Nicolas, J. L., Kervarec, N., Engelen, A., Walsh, R., Guerard, F., Bourgougnon, N. and Stiger-Pouvreau, V. Assessment of the spatial variability of phenolic contents and associated bioactivities in the invasive alga *Sargassum muticum* sampled along its European range from Norway to Portugal, *Journal of Applied Phycology.*, 2014, 26(2), 1215–1230.
16. Cox, S., Gupta, S. and Abu-Ghannam, N. Effect of different rehydration temperatures on the moisture, content of phenolic compounds, antioxidant capacity and textural properties of edible Irish brown seaweed, *LWT - Food Science and Technology.*, 2012, 47(2), 300–307.
17. Siah, S., Wood, J. A., Agboola, S., Konczak, I. and Blanchard, C. L. Effects of soaking, boiling and autoclaving on the phenolic contents and antioxidant activities of faba beans (*Vicia faba* L.) differing in seed coat colours, *Food Chemistry.*, 2014, 142, 461–468
18. Zhong, Y., Khan, M. A. and Shahidi, F. Compositional characteristics and antioxidant properties of fresh and processed sea cucumber (*Cucumaria frondosa*), *Journal of Agricultural and Food Chemistry.*, 2007, 55, 1188–1192.
19. Ee Shian, T., Aminah, A., Musa, K. H., Maskat, Y. M. and Maaruf, A. G. Antioxidant properties of three banana cultivars (*Musa acuminata* sp. berangan, mas dan raja) extracts, *Sains Malaysiana.*, 2012, 41(3), 219-324.
20. Alyaqoubi, S., Aminah, A., Samudi, M., Norrakiah, A., Addai, Z. R. and Al-Ghazali, M. Effect of different factors on goat milk antioxidant activity. *International Journal of ChemTech Research.*, 2014, 6(5), 3091-3196.

21. Hlail, S. H., Ahmad, W. J. W. and Abdullah, A. The influence of growing environment on the total phenol content and antioxidant activity of *Ficus hispida* leaves and fruits, *International Journal of ChemTech Research.*, 2014, 6(7), 3742–3747.
22. Dudonné, S., Vitrac, X., Coutière, P., Woillez, M. and Mérillon, J. M. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays, *Journal of Agricultural and Food Chemistry.*, 2009, 57(5), 1768–1774.
23. Wang, T., Jónsdóttir, R. and Ólafsdóttir, G. Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds, *Food Chemistry.*, 2009, 116(1), 240–248.
24. Okada, Y., Ishimaru, A., Suzuki, R. and Okuyama, T. A new phloroglucinol derivative from the brown alga *Eisenia bicyclis*: potential for the effective treatment of diabetic complications, *Journal of Natural Products.*, 2004, 67, 103–105.
25. Santoso, J. Y., Yoshie, T. and Suzuki, T. The distribution and profile of nutrients and catechins of some Indonesian seaweed, *Fish Sciences.*, 2002, 68, 1647–1648.
26. Jimenez-Escrig, A., Jimenez-Jimenez, I., Pullido, R. and Saura-Calixto, F. Anti-oxidant activity of fresh and processed edible seaweeds, *Journal of the Science of Food and Agriculture.*, 2001, 81, 530-534.
27. Chakraborty, K., Praveen, N. K., Vijayan, K. K. and Rao, G. S. Evaluation of phenolic contents and antioxidant activities of brown seaweeds belonging to *Turbinaria* spp. (Phaeophyta, Sargassaceae) collected from Gulf of Mannar, *Asian Pacific Journal of Tropical Biomedicine.*, 2013, 3(1), 8–16.
28. Thaipong, K., Boonprakob, U., Crosby, K., Cisneros-Zevallos, L. and Hawkins Byrne, D. Comparison of ABTS, DPPH, FRAP, and ORAC assays for estimating antioxidant activity from guava fruit extracts, *Journal of Food Composition and Analysis.*, 2006, 19(6–7), 669–675.
29. Schaich, K. M., Tian, X. and Xie, J. Reprint of “Hurdles and pitfalls in measuring antioxidant efficacy: A critical evaluation of ABTS, DPPH, and ORAC assays”, *Journal of Functional Foods.*, 2015, 18, 782–796.
