



International Journal of ChemTech Research CODEN (USA): IJCRGG, ISSN: 0974-4290, ISSN(Online):2455-9555 Vol.10 No.5, pp 681-687, 2017

# Validated Stability-Indicating Isocratic RP-HPLC Method of estimation of Montelukast Sodium and Fexofenadine hydrochloride in Bulk and in Solid dosage by Vieordt's method

Rajeev kumar P\*<sup>1</sup> and Rekha Rajeevkumar<sup>2</sup>

 <sup>1</sup>Department of Pharmaceutics, School of Pharmacy, PRIST University, Manamai-Nallur, ECR, Near Mahabalipuram, Chennai, Tamilnadu -603102, India.
 <sup>2</sup>Department of Pharmaceutical Analysis, School of Pharmacy, PRIST University, Manamai-Nallur, ECR, Near Mahabalipuram, Chennai, Tamilnadu -603102, India.

Abstract : A simple, speedy, precise, and accurate, stability-indicating reversed phase high performance liquid chromatographic method was developed and validated for simultaneous (Vieordt's method) determination of montelukast sodium(MTK) and fexofenadine hydrochloride (FXD). The sample injection volume was 20 µl and the quantification was attained by UV-VISIBLE detector at 248nm. The chromatographic separation was achieved on X bridge C<sub>18</sub>, 250 x 4.6 mm, 5 µm particle column, using an isocratic mobile phase comprising of acetonitrile: buffer (10 mM potassium dihydrogen phosphate solutions): methanol of pH 4.5 and in the ratio of 50:30:20 v/v/v at a flow rate of 1.5 ml/ min.The retention times for montelukast sodium and fexofenadine hydrochloride were found to be 3.62 min and 7.43 min, respectively. The drugs were exposed to thermal, photolytic, hydrolytic, and oxidative stress conditions, and the stressed samples were analyzed by the suggested method. Validation of the method was done as per International Conference on Harmonization (ICH) guidelines. Linearity was established for montelukast sodium and fexofenadine hydrochloride in the range of 0.020-0.100 mg/ml and 0.016-0.064 mg/ml, respectively. The limits of detection were 0.04  $\mu$ g/ml and 0.07  $\mu$ g/ml, respectively and the LOQ value 0.11 µg/ml and 0.023 µg/ml, for montelukast sodium and fexofenadine hydrochloride respectively. The method was found to be specific and stability-indicating as no interfering peaks of degradants and excipients were perceived. The proposed method is hence suitable for use in quality-control laboratories for quantitative analysis of both the drugs bulk and in combination, since it is simple and fast with good accuracy and precision.

**Key Words:** Montelukast sodium (MTK), Fexofenadine hydrochloride (FXD), reversed-phase HPLC, stability-indicating assay, forced degradation studies, method validation.

# Introduction

Montelukast sodium (MTK),  $1-[(\{(R)-m-[(E)-2-(7-chloro-2-quinolyl) vinyl]-\alpha-[o-(1-hydroxyl-1-methylethyl) phenethyl] benzyl thio) methyl] cyclopropaneacetate sodium[1,2] (Figure 1a). Montelukast (trade name$ **Singulair**) is a leukotriene receptor antagonist (LTRA) used for the maintenance treatment of asthma and to relieve symptoms of seasonal allergies. It is usually administered orally.Leukotriene receptor antagonist (LTRA) used for the maintenance treatment of asthma (LTRA) used for the maintenance treatment of asthma and to relieve symptoms of seasonal allergies [3].

Fexofenadine hydrochloride (FXD) [4-6] (Allegra, Telfast, Fastofen, Tilfur, Vifas, Telfexo, Allerfexo) is an antihistamine drug used in the cure of hay fever and similar allergy symptoms. It was developed as a successor of and substitute to terfenadine (brand names include Triludan and Seldane), an antihistamine with potentially serious contraindications.  $\alpha$ ,  $\alpha$ -dimethyl-4-[1-hydroxy-4-[4-(hydroxydiphenylmethyl)-1-piperidinyl] butyl] benzeneneacetic acid, is the most important terfenadine metabolite shown in Figure 2.H<sub>1</sub>-receptor antagonist recommended in patients with allergic rhinitis[7]. Fexofenadine hydrochloride, like other second and third-generation antihistamine-receptor antagonists. It works by being an antagonist to the H<sub>1</sub> receptor. It has been termed as both second-generation and third-generation [3, 8].

To the best of our knowledge no method is reported in literature for simultaneous determination of Montelukast sodium and Fexofenadine by high performance liquid chromatography (HPLC) using the specified chromatographic condition. The survey of literature showed few UV Spectrophotometric [9], HPLC [10-13], LC-ESI–MS/MS method [14] are available for the estimation of Montelukast sodium in pharmaceutical preparation and in biological fluids. Literature survey also showed that available for the estimation of UV spectrophotometric [15-17] for the estimation of Fexofenadine in pharmaceutical preparation.

The objective of the present work was to develop and validate a simple, economic, rapid, precise, gradient and accurate stability-indicating method with good sensitivity for simultaneous determination of montelukast sodium and fexofenadine hydrochloride in accordance with ICH guidelines [18]. The proposed method was successfully applied to a solid dosage form of montelukast sodium and fexofenadine hydrochloride and analyzed in presence of commonly used tablet excipients. This method can also be employed for quality control during manufacture of drug product.

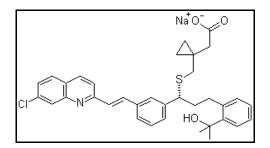


Figure 1: (a) Structure of MTK.

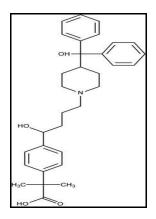


Figure 1: (b) Structure of FXD.

### **Materials and Methods**

#### Chemicals, Reagents, and Solutions

Montelukast sodium and fexofenadine were gift sample from Ami Life science, Baroda, Gujarat. HPLC grade acetonitrile, methanol and analytical grade potassium dihydrogen ortho phosphate,ortho phosphoric acid, triethylamine was obtained from S.D. Fine chemicals Ltd. (Mumbai, India). Hydrochloric acid, sodium hydroxide pellets and 3% v/v hydrogen peroxide solution were obtained from Ranbaxy Fine Chemicals, New Delhi (India). High purity water was prepared by Millipore Milli-Q plus purification system (Millipore, Bedford, USA).

#### **HPLC Instrumentation and Chromatographic Conditions**

The chromatographic system used to perform development and validation of this assay method was comprised of a Isocratic (LC 20AT) pump, UV-VISIBLE [SPD 20A] DETECTOR and a rheodyne manual injector with 20 $\mu$ l loop, Column X bridge C<sub>18</sub>, 250 x 4.6 mm, 5  $\mu$ m particle connected to a multi-instrument data acquisition and data processing system. The flow rate of the mobile phase was 1.5mL/ min. The sample injection volume was 20  $\mu$ L and the quantification was achieved by UV-VISIBLE (SPD 20A) detector at 248nm. The isocratic program applied was carried out.

#### Mobile phase preparation

Mobile phase consisted of A, B and C. Mobile phase A and C was HPLC grade acetonitrile and methanol. Mobile phase B composed of buffer solution of 0.2 M potassium dihydrogen orthophosphate in 0.1M phosphoric acid adjusted with triethylamine (0.5%) to pH 4.5. The above solution was filtered through 0.45  $\mu$ m nylon filter. Finally the mobile phase A, B and C was mixed in the ratio of acetonitrile: buffer: methanol solutions of pH 4.5 and in the ratio of 50:30:20 v/v/v.

#### **Standard Preparation**

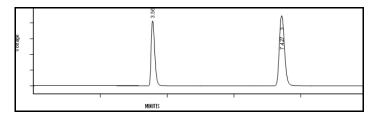
Accurately weighed and transferred 10 mg of montelukast and 120 mg of fexofenadine working standards into two separate 100 ml volumetric flask.add 70ml of mobile phase and sonicated for 10 minutes and diluted to volume with mobile phase. Further 1ml of montelukast sodium and 1 ml of Fexofenadine standard stock solution was transferred into 100 ml volumetric flask and made up to the mark with mobile phase. The concentration obtained was 10  $\mu$ g/ml of montelukast sodium and 12 $\mu$ g/ml of fexofenadine. The above solution was filtered through a 0.45 $\mu$  nylon membrane filtered.

### **Test Preparation**

Twenty tablets were weighed and the average weight of tablet was determined. The tablets were powdered and accurately weighed quantity of montelukast sodium equivalent to 10 mg of montelukast and 120mg of fexofenadine in a 100 ml volumetric flask in mobile phase. This solution is then sonicated for 10 minutes and diluted to volume with mobile phase. Further 1ml of this stock solution is taken in a 100 ml volumetric flask and made up to the mark with mobile phase (this  $10\mu g/ml$  of montelukast sodium and  $12 \mu g/ml$  of fexofenadine sample solution. The sample was filtered through 0.45 $\mu$ m nylon syringe filter.

#### **Results and Discussion**

The conditions tested for the method development indicates that all the system suitability parameters according to ICH guidelines was achieved by using X bridge  $C_{18}$ , (250 x 4.6 mm, 5 µm particle) column using mobile phase composition consisting of a mixture of acetonitrile : buffer (0.2 M potassium dihydrogen orthophosphate in 0.1M phosphoric acid adjusted with triethylamine (0.5%) to pH 4.5) : methanol (50:30:20, v/v) using a isocratic program with a flow rate of 1.5 ml/min throughout isocratic program with a detection wavelength of 248nmfor both the compounds with a injection volume of 20 µl. The peak at  $R_t$ 3.62 min and 7.43 min for Montelukast sodium and Fexofenadine was observed in the chromatogram of the drug samples extracted from the tablet shown in figure. 3.



#### **Figure 3: Chromatogram of test preparation**

To validate RP-HPLC method [19], a series of tests were made using the most promising conditions. A calibration curve was made and concentration examined within the detection range from 0.020-0.100 mg/ml for Montelukast sodium and 0.016-0.064 mg/ml for fexofenadine. The correlation coefficient was found to be 0.9991 and 0.9998 for Montelukast sodium and fexofenadine respectively. The assay values shown in (Table 1).

Table 1: Assay of	Montelukast sod	ium and Fexo	fenadine hv	vdrochoride

Brand name	Compound	Amount found(mg)	%Assay
MONTAIR FX	Montelukast sodium	10.13	101.3
	Fexofenadine	119.89	99.9

The percent recovery of Montelukast sodium and fexofenadine were determined at determined at three different concentration levels like 80, 100 and 120%. The mean recovery for Montelukast sodium was 98.36-100.15 % and 99.68-100.8 % for fexofenadine. Data obtain from precision experiments are carried out for intraday and interday precision study for both Montelukast sodium and fexofenadine. The RSD values for intra day precision study and interday precision study was < 2.0 % for Montelukast sodium and fexofenadine. The result indicating that the method was accurate and confirms good precision. The results are presented under (Table 2) and (Table 3).

Set	Montelukast (%Assay)	sodium	Fexofenadin	Fexofenadine (%Assay)	
	Intraday (n = 6)	Interday (n = 6)	Intraday (n = 6)	Interday (n = 6)	
1	101.4	100.4	100.5	98.9	
2	100.9	100.8	100.1	101.5	
3	98.5	101.3	100.9	101.5	
4	99.7	99.9	98.9	99.8	
5	100.8	101.7	99.8	99.9	
6	100.1	100.5	99.6	99.7	
Mean	100.6	100.7	99.96	100.2	
Standard deviation	0.513	0.219	1.10	0.423	
% RSD	0.827	0.119	1.12	0.386	

## Table 2: Results of precision study

## Table 3: Results of accuracy study

	Level (%)	Theoretical Concentration <sup>a</sup> (µg/ml)	Observed Concentration <sup>a</sup> (µg/ml)	% Recovery	% RSD
Montelukast	80	41.52	41.35	99.59	0.49
sodium	100	51.96	51.11	98.36	1.38
	120	62.28	62.90	100.15	0.32
fexofenadine	80	482.6	481.1	100.08	1.23
	100	602.7	601.83	99.68	1.37
	120	723.2	729.2	100.8	1.68

<sup>a</sup> Each value corresponds to the mean of three determinations

The method was demonstrated to be robust over an acceptable working range of its HPLC operational conditions. The system suitability results within the acceptable limits and selectivity of individual substances were also not affected when subjected deliberately for varied chromatographic conditions. The result of the study confirms the robustness of the method. The result of robustness study of the developed assay method was established in (Table 4) and (Table 5).

Robust conditions	% Assay	System suitability parameters	
		Theoretical plates	Asymmetry
Flow 0.9 ml/min	101.2	5002	1.65
Flow 1.5 ml/min	100.8	5022	1.34
Buffer pH 5	99.0	5010	1.54
Buffer pH 5.5	99.4	5100	1.38
Buffer-ACN (60:40:10% v/v)	98.7	5018	1.73
Buffer-ACN (50:30:20% v/v)	99.5	5108	1.45
Column change	100	5090	1.39

Robust conditions	% Assay	System suitability parameters	
		Theoretical plates	Asymmetry
Flow 0.9 ml/min	100.4	10156	1.65
Flow 1.5 ml/min	101.5	10256	1.44
Buffer pH 5	100.9	10100	1.50
Buffer pH 5.5	99.9	10313	1.33
Buffer-ACN (60:40:10% v/v)	99.7	10005	1.56
Buffer-ACN (50:30:20% v/v)	99.9	10181	1.45
Column change	101.1	10162	1.67

 Table 5: Evaluation data of robustness study of Fexofenadine

The stability of sample was checked by forced degradation in different stress conditions condition like acidic, alkali, thermal, oxidative degradation, photolytic and humidity conditions and % of degradation was calculated. The peak purity of the analyte was passed in all conditions (purity angle should be less than the threshold value). The following values in (Table 6) indicate that any other impurity is not merging with the main peak (figure 4-9).

 Table 6: Forced degradation studies of control sample (1mg/mL) solution.

Type of Degradation	No of unknown impurities	Montelukast sodium and fexofenadine peak area %	% Degradation
Control sample	Nil	99.63	_
Acid	Nil	99.92	-
Alkali	Nil	99.89	
Peroxide	Nil	99.74	-
Thermal	Nil	99.96	_
Photolytic	Nil	99.97	_
Humidity	Nil	99.91	_

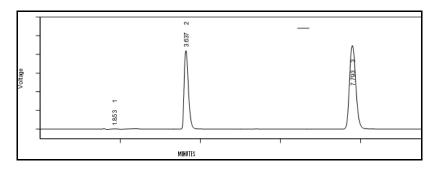


Figure 4: Chromatogram of acidic forced degradation study

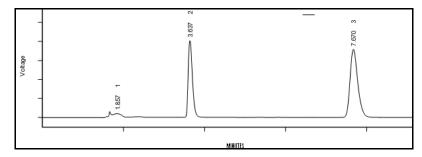


Figure 5: Chromatogram of alkali forced degradation study

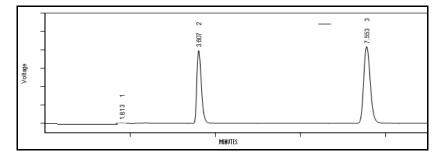


Figure 6: Chromatogram of oxidative forced degradation study

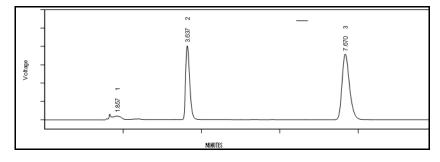


Figure 7: Chromatogram of thermal degradation study

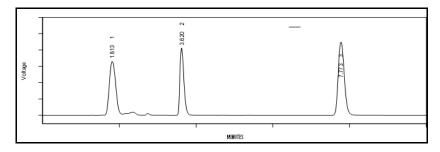


Figure 8: Chromatogram of UV-light degradation study

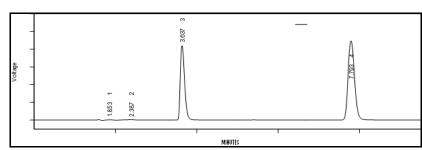


Figure 9: Chromatogram of humidity degradation study

It was determined that the test preparation solution was found stable up to 48 hr at 2 - 5 °C and room temperature, as during this time the result was not decrease below the minimum percentage.

### Conclusion

A simple, fast, accurate, and precise stability-indicating HPLC analytical method has been developed and validated for the routine analysis of montelukast sodium and fexofenadine hydrochloride in active pharmaceutical ingredient(API) and can be useful to pharmaceutical sold dosage forms. The results of stress testing was carried out according to the ICH guidelines reveal that the method is specific and stabilityindicating. The developed method has the capability to isolate these drugs from their degradation products; common excipients used in pharmaceutical solid dosage forms, and hence can be applied to the analysis of routine quality control samples and samples obtained from stability studies.

## Acknowledgments

The authors wish to thank School of Pharmacy, PRIST University, Chennai, Tamilnadu for providing necessary facilities and to carry over the work. The authors wish to express their gratitude to Ami Life Science, Baroda, Gujarat and Cadila pharmaceutical, Ahmedabad for providing the authentic sample (Montelukast sodium and Fexofenadine hydrochloride).

# References

- 1. Sweetman SC, editor. Martindale: The Complete Drug Reference. 33<sup>rd</sup> ed., London: Pharmaceutical Press; 2002.
- 2. Budavari S, editor. Merck & Co Inc., The Merck Index. 12<sup>th</sup> ed., Whitehouse Station, NJ; 1996.
- 3. JD Morrow; LJ Roberts. In: Goodman and Gilman's The Pharmacological Basis of Therapeutics. 10th ed. Hardman JG, Limbird LE, Gilman AG, editors. New York: McGraw-Hill; 2001.
- 4. K Katagiri; S Arakawa; Y Hatano; S Fujiwara. J of Dermatol, 2006; 9-75.
- 5. PV Dicpinigaitis and YE Gayle. Br J of Clin Pharmacol, 2003;01-04.
- 6. GA Vena; N Cassano; M Filieri ;R Filotico ;V D'Argento ;C Coviello. Int J of Immunopathol and Pharmacol, 2002; 24-217.
- 7. Guidelines for the Diagnosis and Management of Asthma. 3<sup>rd</sup> report. National Heart, Lung, and Blood Institute; 2007 (US department of health and human services).
- 8. Lipkowitz ; A Myron ; Navarra and Tova .The Encyclopedia of Allergies 2<sup>nd</sup> ed. 2001.
- 9. RM Singh; PK Saini; SC Mathur; GN Singh and Lal B. Indian J Pharm Sci, 2010,37-235.
- 10. BR Challa; BZ Awen; BR Chandu; M Khagga and CB Kotthapalli. Sci Pharm, 2010, 22-411.
- 11. SA Patel ; SK Patel ; DJ Patel and NJ Patel.Int J of Pharm Tech Res, 2010, 71-1767.
- 12. DJ Patel ; SA Patel and SK Patel. Int J of Pharma and Biomed Res, 2010, 5-71.
- 13. S Ashokkumar; M Senthil raja and P Perumal. Int J of Pharm Research, 2009, 8-12.
- 14. K Suresh Kumar; V Ravichandran; MK Mohan; R Thyagu and A Dharamsi. Indian J Pharm Sci, 2006, 42-841.
- 15. DC Oliveira; A Weigch and CM Rolim. Pharmazie, 2007, 96-100.
- 16. PV Polawar; UD Shivhare ; KP Bhusari ; VB Mathur . Research J Pharm and Tech, 2008, 1(4).
- 17. I Kozan; IM Palabiyik; E Karacan and F Onur . Turk J Pharm Sci, 2008, 89-175.
- 18. ICH Harmonised tripartite guideline: Text on validation of Analytical Procedures.Q2A and Q2B; 1995; 1-5 and 1-7.

\*\*\*\*\*