

International Journal of ChemTech Research

CODEN (USA): IJCRGG, ISSN: 0974-4290, ISSN(Online):2455-9555 Vol.10 No.3, pp 90-99, 2017

ChemTech

Bioactive Compounds From Majapahit Fruit (*Crescentia cujete*) As a Potential Natural Antibacterial

Sri Rahmaningsih¹*, Arief Prajitno², Aulanni'am Aulanni'am ³, Maftuch²

¹Graduate School of Fishery and Marine Sciences ²Department of Aquaculture, Faculty of Fishery and Marine Sciences ³Department of Chemistry, Faculty of Sciences Brawijaya University, Malang, Indonesia

Abstract : The source of active ingredients that are commonly used in treating or preventing diseases, and are thought to play a beneficial role in health, for example are plants. Majapahit (Crescentia cujete) plant is one of such plants, whose leaves, barks, fruits, or roots have medicinal and preservative properties. These studies are needed to ascertain the compound of Majapahit fruit as a Natural Antibacterial and Identification of active ingredients using various characterisation techniques. The methanol, ethyl acetate, and, n-hexane extract of these fruit were prepared, and agar diffusion method with paper disc to investigate antimicrobial activity. Identification of active ingredients with; Phytochemicals Test, UV-VIS Spectrophotometry, FTIR spectrophotometer and Gas Chromatography-Mass Spectrometry (GC-MS). Results of the phytochemical test revealed the presence of flavonoids, saponins, , and triterpenoid in the Majapahit fruit. The methanol extract was the most effective (17.29 mm) in antibacterial activity. Ultraviolet spectrum is with maximum peaks at 407.0,- 396.9 nm and 313.0, - 219 nm, result of FTIR spectroscopy are wave numbers 3662 cm⁻¹, - 518 cm⁻¹ that indicates of; OH⁻ bending, -CH- (SP3), OH⁻, -C=C-, -CH₂-, -CH₃-, and -C=CH-. The GC-MS chromatogram indicating the presence of 12 phytochemical constituents from 5 peaks highest. Majapahit (Crescentia cujete) fruit which implies that the extracts have a potential natural antibacterial. The phytochemical test revealed the presence of flavonoids, saponins, and triterpenoids, and The bioactive constituents of the plant extract were analysed by FTIR, UV-VIS spectrophotometry and GC-MS spectrometry, and they revealed varied and wide compounds.

Keywords: Majapahit (*Crescentia cujete*) fruit, antibacterial, flavonoids, saponins,triterpenoids.

Introduction

Medicinal plants abound around the world, and in Indonesia, it is reported that out of about 30,000 plant species, about 9,600 species have medicinal properties¹. Despite this abundance, only about 300 Indonesian medicinal plant species have been (industrially) explored for their beneficial properties¹. These beneficial properties are conferred on these plants by their bioactive ingredients, and plants are a green technology that can be used in treating or preventing diseases, preserving foods and controlling food spoilage. They are, therefore, valuable in food and healthcare systems.

91

Majapahit (*Crescentia cujete*) plant is one of such plants, whose leaves, barks, fruits, or roots have medicinal and preservative properties. Various parts of the plant have been used as a purgative, diuretic, analgesic, or anti-inflammatory drug possibly because they contain alkaloids, flavonoids, saponins, phenolics tannins, steroids, vitamins, carbohydrates, organic acids, amino acids and mineral ^{2,3,4,5} Specific bioactive compounds in the plant include β -sitosterol, estigmastrol, α - and β -amirina, apigenins, and carotenoids ^{2,6} These constituents are also responsible for the antibacterial properties of the plant, having been proven to effectively inhibit the growth of *Staphylococcus aureus, Enterococcus faecalis, Streptococcus pneumoniae, Streptococcus pyogenes, Escherichia coli, Candida albicans*, and *Ralstonia solanacearum*^{7,8} Although these are foodborne organisms or organisms with healthcare concerns, the effectiveness of Majapahit plant for specific organisms that are responsible for diseases, and spoilage and health concerns in seafood, such as *Vibrio* spp., have not been extensively studied.

Various studies on antimicrobial properties of plants have been reported.⁹ studied patikan kerbau (*Euphorbia hirta*), and found it to be effective against bacteria *Vibrio alginolitycus* and *Aeromonas hydropila*. Extracts of plant *Acumata Alstonia* were shown to have antibacterial properties against *Vibrio harveyi*¹⁰, which was also inhibited ¹¹ by the leaf extracts of kopasanda (*Chromolaena odorata* L.).¹² showed the extracts of seaweed *Caulerpa* spp. to be most active against *Pseudomonas bacteria*, and bacteria *Vibrio* spp. was inhibited ¹³ by other seaweed species (*Gelidium* spp., *Sargasum polycistum* and *Eucheuma cottoni*).⁵ investigated antimicrobial activities of certain species of the family Biogenaceae (*Kigelia aficana, Jacaranda mimosifolia, Millingtonia hortensis, Tabebuia argentia, Dolichandron* spp. and *Haplophragm* spp.), and possibly because of their potentials, these authors advised more studies on the species of the family Bignoniaceae. Majapahit plant, is a specie of the family Bignoniaceae, and ¹⁴, showed that the leaf extracts had a greater inhibition zone than the fruit and bark extracts against bacteria *Vibrio alginolyticus*, possibly because of differences in the bioactive constituents of these parts. However, ¹⁴ did not comprehensively investigate the differences in the constituents of these plant parts. The solubility of these constituents in polar and non-polar solvents will influence the concentrations of the plant constituents in the extracts, and consequently, the extents of their medicinal and preservative properties ^{2,15,16}.

The common solvents used in extracting plants constituents include ethanol, chloroform, benzene, water, n-hexane, methanol, and ethyl acetate ^{17, 18,19,20}. ¹⁸ extracted the leaf and bark of Majapahit plant with ethanol and chloroform, and found the ethanol extracts to be more effective against *Staphylococcus aureus* and *Escherichia coli*.¹⁹ investigated the antimicrobial properties of *Sonneratia lanceolate* against *Vibrio harveyi* by using water and methanol extracts of the plant. These authors found the water extracts to be more effective. Antifungal activities of *Aegle marmelos* leaf extracts in hexane, benzene, chloroform, ethyl acetate, methanol, water, and ethanol were investigated ²⁰ and chloroform, methanol, water, and ethanol extracts had the least minimum inhibition concentration and were more effective⁻⁵ extracted the stem, bark and leaves of *Cresentia cujete* using water and ethanol. These authors concluded that the aqueous extracts of the plant parts. Despite its potential beneficial properties for health and food, studies on the constituents of Majapahit plant are limited, and the differences between the various parts have not been comprehensively examined, nor possible differences in the effectiveness of extracting solvents. These studies are needed to ascertain or classify Majapahit fruits as a medicinal plant with potential natural antibacterial and food preservative properties. The present study was aimed along these lines using novel characterisation techniques.

Material and Methods

Materials

Majapahit fruit was obtained from Tuban, Indonesia. The fruit was correctly identified as *Cresentia cujete* L. by the Indonesian Centre for Research in Biology, Bogor, Indonesia. TLC plates were a product of Merck (Darmstadt, Germany). All chemicals, reagents and solvents used were pure and/or of analytical grade.

Preparation of raw materials and extraction

The fruit was cleaned, cut into small pieces and air-dried, pulverized in a blender (HR2102, Phillips, Indonesia) to a fine (passed through a 250-µm sieve) powder. About 50 g of the powder was soaked in 250 mL of methanol for about 72 hr in a glass jar, before the supernatant was separated from the pulp. The supernatant

was then evaporated in a rotary evaporator under vacuum at about 45° C to obtain a concentrated extract with no trace of the solvent. Other solvents investigated following the same procedure (with or without vacuum) were water, ethanol, acetone, ethyl acetate, diethyl ether, n-hexane, and chloroform. These solvents were chosen in order to examine the best that maximises the beneficial properties of the Majapahit plant.

Phytochemicals test

The phytochemical test procedure of ²¹ was used, with about 20 mg of the powder dissolved in 25 mL of 96% ethanol to detect alkaloids, flavonoids, saponins, tannins, triterpenoids and steroids. The TLC followed standard procedure, and the plates were activated in the oven at a temperature of 100 - 105°C for 30 min.

Antibacterial test

The extracts were tested against *Vibrio harveyi* at a concentration of 10 mg/mL using the agar diffusion method with paper disc ^{22,11} Sterile paper discs (Oxoid, Basingstoke RG24 8PW, UK) were dipped in the extracts for 15 - 30 min. before placing the paper disc in the Mueller-Hinton Agar (MHA) media that had been inoculated with the bacteria. Measurements were made after an incubation period of 24 hr at 37°C by observing the presence or absence of clear zones formed around the paper disc ^{23,11} *V. harveyi* is a well-known food poisoning bacteria found in marine animals ^{24,25} and it was chosen because, one of the aims of the long-term research under which the present study falls, was to find a green technology for preservation of seafood.

Identification of active ingredients

UV-VIS Spectrophotometry

A known volume (0.1 mL) of the extracts was diluted to 10 mL with appropriate solvents, and the solutions were measured at wavelengths of 200 - 400 nm using a UV-VIS spectrophotometer (Varian Type Cary 50, Agilent, Santa Clara, CA 95051, USA).

Fourier Transform Infra-Red Spectroscopy (FTIR)

Infrared spectroscopy was applied to confirm functional groups in the extracts. A drop of each extract was mixed with KBr to make pellets (2g), which were subsequently analyzed in an FTIR spectrophotometer model (FTIR 1000 SCIMITAR, California, USA) at wavenumbers 4000 - 800 cm⁻¹.

Gas Chromatography-Mass Spectrometry (GC-MS)

The extracts were analysed by GC-MS (GC-MS QP2010, Shimadzu, Kyoto 604-8511, Japan), using helium as the carrier gas. The injection temperature was 320° C, and thermal gradients were applied from 700° C - 300° C within 4 hr with a gradient rate of 100° C/hr.

Result and discussion

Phytochemical profile

The phytochemical test revealed the presence of flavonoids, , saponins, and triterpenoid, in the Majapahit plant (Table 1). Flavonoids are reported to be the most natural, widely spread components in plants ^{22,9,11,10} and their presence in the fruit is not surprising.

Tabel 1. Result for phytochemical screening in Majapahit fruit (Crescentia cujete)

| Compound | Observation | Inference |
|---------------|---------------|-----------|
| Flavonoids | Orange | Present |
| Saponins | Little foam | Present |
| Tanins | Brown | Absent |
| Steroids | Brown | Absent |
| Triterpenoids | Brown | Present |
| Alkaloids | Orange Cloudy | Absent |

Antibacterial activity

Solvents vary in polarities, and consequently, the abilities to dissolve bioactive constituents of the Majapahit plant fruit. With differences in the dissolved constituents, the extracts might vary in their overall beneficial properties, and Table 2 shows that the efficacy of the extracts was solvent dependent as expected. As highlighted above ¹⁸ found the ethanol extracts of the leaf and bark of Majapahit plant to be more effective than the chloroform extracts on *Staphylococcus aureus* and *Escherichia coli*. Similar solvent differences were reported by ^{19,20} amongst others. In the present study, it appears that methanol was the most effective, while n-hexane was ineffective.

| Samples | Zone of Inhibition (mm) | | | | |
|-----------------|-------------------------|--------------------|---------------------|--|--|
| | n-Hexane | Ethyl acetate | Methanol | | |
| Majapahit fruit | 2.4 ^c | 14.66 ^b | 17.29 ^a | | |

Table 2. Extract of Majapahit fruit (Crescentia cujete) inhibition zone against V. harveyi

*Values with the same letters are not significantly (p > 0.05) different. The zone of inhibition of erythromycin (positive control) was 26.4 mm

Table 1 shows the presence of flavonoids, saponin and triterpenoid in the fruits, and these have antibacterial properties, and the presence of these compounds could explain the results in Table 2. However, different solvents differently solubilize various compounds of the flavonoid group. Flavonoids are phenolic structures containing one carbonyl group complexes with extra cellular and soluble protein and bacterial cell wall, and they exhibit antibacterial activities through these complexes ²⁶. Flavonoids inhibit the growth of bacteria or microorganisms by interfering with their cell wall permeability, and the consequent disruptions cause other compounds such as saponins, triterpenoids, phenolics, alkaloids, and tannins to penetrate and damage the cell wall ²⁷ It was thought that because flavonoids are polar compounds, they easily penetrate the cell walls of microorganisms.

Triterpenoids are a diverse group of secondary metabolites that are associated with a variety of biological activities ²⁸ The main groups of triterpenoids are represented by tetracyclic and pentacyclic derivatives²⁹ Important elements that play a role in the antibacterial activity of triterpenoids relate to chemical compositions that include functional groups and hydroxyl groups of phenolic triterpenoids as well as the number of single components. However, when triterpenoids are used as single compound, they are less effective as an antibacterial. The concentrations are important too ²⁶ because at low concentrations, triterpenoids only affect enzymes involved in energy production while at high concentrations, triterpenoids can lyse membranes. Bacterial species of *Mycobacterium* are supposed to be sensitive to triterpenoids³⁰. Triterpenoids showed bacteriostatic effects against *Staphylococcus aureus* ²⁹. Triterpenoids reportedly exhibit antimicrobial activity against gram-positive bacteria, block cell divisions by inhibiting DNA synthesis and macromolecular synthesis in *Bacillus subtilis*. The inhibition of macromolecular synthesis could be due to the damage to cell membranes³¹ influence of cell morphology, enhancement of detergent-induced lysis, and autolysis of isolated cell walls.

Saponins are glycosides occurring widely in plants, and are complex compounds of condensation products of a sugar with an organic hydroxyl compound, which when hydrolyzed would produce sugars (glikon), non-sugar (aglycone) and foam ³² Saponins compose of two groups, triterpenoid saponins and steroidal saponins, which are respectively fat- and water-solubles. When concentrated on the cell membrane, they lower the surface tension and works as an antimicrobial to destabilize bacterial cell membranes causing bacterial cells to die through lysis ^{33,15,32} As surface active agents, they interfere with or alter the permeability of cell walls, thereby facilitating the entry of toxic materials to or leakages of vital constituents from the cells ^{34,32} showed the crude saponin extracts prepared from *Sorghum bicolor* posses antimicrobial property, and were active against *Staphylococcus aureus*, a pathogen, which has been implicated in several human and animal infections. This is similar to reports by³³ that saponins from *Hibiscus sabdariffa* were also active against *Escherichia coli, Klebseilla pneumoniae, Bacillus cereus, and Micrococcus luteus*. In order to fully understand the measured antibacterial activities of the Majapahit fruit, specific information on the extracts was obtained using the following techniques.

Analysis compounds of the extracts majapahit fruit by UV-VIS Spectrophotometry

There were 47 wavelength spectra from 200 to 800 nm (Fig. 1), with maximum peaks at 407.0 - 396.9 nm and 313.0 - 219 nm. The occurrence of a strong absorption in the ultraviolet spectrum showed the presence of a number of C = C double bonds in the compounds from the Majapahit fruit. This strong uptake showed a phenol compound and its derivatives.



Figure 1. A typical spectrum of the compound fractions of Majapahit fruit

From published results, the possible compounds that correspond to the prominent wavelengths of maximum absorption from the extracts of the Majapahit fruit, are summarized in Table 3.

. ~

| No | wavelength result of UV-VIS | Wavelength Range based on the literature | Compounds | PubChem CID |
|----|-----------------------------------|--|---|--------------|
| | 269.0 | 269.0 | Ezetimibe | 150311 |
| | 223,0 | 223,0 | Fosinopril | 55891 |
| | 288.0 | 288.0 | Gemcitabine | 60750 |
| | 257.1 | 257.1 | ketoconazole | 47576 |
| | 284.1 | 284.1 | Rabeprazole | 5029 |
| | 253.0 | 253.0 | repaglinide | 65981 |
| | 308.0, | 308.0, | rofecoxib | 5090 |
| | 293 | 293 | Vincmine | 15376 |
| | 271.0 | 271.0 | Aceclofenac; Paracetamol | 71771;1983 |
| | 280.0 | 280.0 | Ibuprofen; Paracetamol | 3672;1983 |
| | 294 | 294 | Levamisole; oxyclozanide | 26879; 16779 |
| | 313.0 | 313.0 | Aceclofenac; tizanidine | 71771; 5487 |
| | 219 | 219 | Amitriptyline | 2160 |
| | 247.8 | 247.8 | Diflucortolone valerate | 33140 |
| | | | Isoconazole nitrate | |
| | 271.0 | 271.0 | Hydrochlorothiazide Olmesartan Medoxomil | 9940864 |
| | 295.0 | 295.0 | Ofloxacin; Tinidazole | 4583; 5479 |
| 1 | 274, 271, 267, | 240 - 290 | Tramadol hydrochloride | 63013 |
| | 261, 250.9, | | | |
| | 247.9, 245 | | | |
| | | 240 - 285 | Ibuprofen (IB);Pseudoephedine | 3672; 7028 |

| Table 3.Possible | compounds from | n the prominent | wavelength from | the extracts o | f Maiapahit fruit |
|------------------|----------------|------------------|------------------------|----------------|--------------------|
| | compounds not | n vine prominent | , way chongen it offic | | i initujupume muit |

| | | 210 - 280 | Hydrochlorothiade; Quinaphril | 3639; 54892 |
|---|------------|-----------|-------------------------------|-------------|
| | | | (QA) | |
| 2 | 237,5, 245 | 235 - 245 | Letrozole | 3902 |
| | | 210 - 280 | Hydrochlorothiade; Quinaphril | 3639; 54892 |
| | | | (QA) | |
| 3 | 232 | 227 - 232 | Cabergoline | 54746 |
| | | 210 - 280 | Hydrochlorothiade; Quinaphril | 3639; |
| | | | (QA) | 54892 |
| 4 | 274, 271 | 271 - 275 | Acerclofenac;Paracetamol | 71771; 1983 |
| ~ | 35.36 | | | |

Source: ^{35,36}

Analysis compounds of the extracts majapahit fruit by FTIR

FTIR spectroscopy showed (Fig. 2) the presence of OH⁻ bending with a wide ribbon shape at the absorption wavenumbers 3662 cm⁻¹, absorption at 2923 cm⁻¹, with a weak ribbon shape that indicates -CH-(SP3) aliphatic stretching. Absorptions at 1645 cm⁻¹ and 1556 cm⁻¹ indicated a -C=C- aromatic with a moderate ribbon shape. Wavenumber 1058 cm⁻¹ showed a primary OH⁻, while wavenumbers 1416, 688, 590, and 518 respectively indicated -CH₂-, -CH₃- scissoring, -C=CH- out of plane, and mono-branched alkanes (-CH₃-) that formed a weak ribbon.



Based on Fig. 2, the suspected compounds from the extracts Majapahit fruit are summarised in Table 4

| Table 4 | Absorption ban | d, wave number | , the type of vi | ibration and FI | FIR results ribbor | ı shape extracts |
|----------|----------------|----------------|------------------|-----------------|--------------------|------------------|
| fruit ma | ijapahit | | | | | |

| Absorption band | Wavenumber (cm ⁻¹) | Reference (Harborne, 1994) | Type of vibration | Shape ribbon |
|--------------------|-----------------------------------|-------------------------------|--|--------------|
| 1 | 3851 | | | |
| 2 | 3734 | | | |
| 3 | 3662 | 3500 - 3200 | -OH bending | splay |
| 4 | 2923 | 3000 - 2700 | -C-H (SP ³) stretching | Weak |
| | | | aliphatic | |
| 6 | 1556 | | | Moderate |
| 7 | 1416 | 1470 - 1450 | -CH ₂ , -CH ₃ scissoring | Weak |
| 8 | 1058 | ~ 1050 | -OH primer | Moderate |
| 9 | 688 | 690 - 590 | -C=CH out of plane | Weak |
| 10 | 590 | | | Weak |
| 11 | 518 | ~ 570 - 540 | Monobranched alkanes | Weak |
| | | | (-CH ₃) | |

Analysis compounds of the extracts majapahit fruit by GC-MS

Figure 3 shows a typical spectrum from the GC-MS test, while Table 5 summarises the suspected compounds and their structures. There were 45 diverse compounds in the Majapahit plant, with four prominent compounds having peaks at 2.64, 5.98, 6.98, and 8.81.



The majapahit fruit extract showed 5 peaks highest in the GC-MS chromatogram indicating the presence of 12 phytochemical constituents. Through comparison with mass spectra 12 phytochemical constituents were characterized and identified.

| 1 able 5. The prominent compounds in the majapanit iru | Fable 5. The | prominent | compounds i | n the | majapahit | fruit |
|--|---------------------|-----------|-------------|-------|-----------|-------|
|--|---------------------|-----------|-------------|-------|-----------|-------|

| No | RT | Area | Compound name | Chemical formula | Compound structure | PubChem CID |
|----|------|------|---|--|--------------------|----------------|
| 1 | 2.21 | 2.64 | Furfural | C ₅ H ₄ O ₂ | Furfural | 7362 |
| | | | Pyrazole,1,4-dimethyl, 3,5- dimethyl-1H-pyrazole | C ₅ H ₈ N ₂ | | 6210 |
| | | 5 24 | | | | 1174 |
| 2 | 5.12 | J.24 | 2,4(1h,3h)- pyrimidinedione | $C_4H_4N_2O_2$ | HN | |
| | | | 1.2.3 Benzenetriol | $C_6H_6O^3$ | ОН | 10787 |
| 3 | 5.81 | 5.98 | 4H-Pyran-4-one, | C ₅ H ₄ O ₂ | ОН | 7968 |
| | | | | | l. | |
| | | | 2,5 difluorophenylhydrazine | $C_6H_6F_2N_2$ | F NH2 | 10920925 |

| | | | 1,2,4,5-tetrazine-3,6- diamine | C ₂ H ₄ N ₆ | H ₂ N NH ₂ | 49863143 |
|---|------|------|-----------------------------------|--|----------------------------------|----------|
| 4 | 7.12 | 6.98 | Furancarboxaldehyde | C ₅ H ₄ O ₂ | | 7968 |
| | | | 4-mercaptophenol | C ₆ H ₆ OS | HSOH | 240147 |
| 5 | 8.89 | 8.81 | 2-propenoic acid, 3-phenyl | C ₉ H ₈ O ₂ | ОН | 444539 |
| | | | 1,3,5-Triazine-2,4,6- triamine | C ₂ H ₄ N ₆ | N NH2 H2N N NH2 | 61176 |
| | | | Trans-Cinnamic acid | C ₉ H ₈ O ₂ | ОН | 444539 |

Conclusion

Majapahit (*Crescentia cujete*) fruit were extracted using methanol more effective against *V. harveyi*, which implies that the extracts have a potential natural antibacterial. The phytochemical test revealed the presence of flavonoids, , saponins, and triterpenoid, and The bioactive constituents of the plant extract were analysed by FTIR, UV-VIS spectrophotometry and GC-MS spectrometry, and they revealed varied and wide compounds.

References

- 1. Akbar, H.,R. Isolation and Identification Group Dandang Gendis Leaf Flavonoid (Clinacanthus nutans) Potential As an antioxidant. *Thesis*. Unpublished, Department of Chemistry, Faculty of Mathematics and Natural Sciences Institut Agriculture Bogor. 2010.
- 2. Ogbuagu M.N. The Nutritive and anti-nutritive Compositions of Calabash (*Crescentia cujete*) Fruit Pulp. Journal of Food Technology. 2008, 6, 267 270.
- 3. Ejelonu, B.C., Lasisi, A.A., Olaremu, A.G., Ejelonu, O.C. The chemical constituents of calabash (*Crescentia cujete*). African Journal of Biotechnology. 2011, 10 (84), 19631-19636
- 4. Das, N., Islam, Md.E., Nusrat Jahan, Islam, M.S., Khan, A., Islam, Md.R., Parvin, S. Antioxidant activities of ethanol extracts and fractions of *Crescentia cujete* leaves and stem bark and the involvement of phenolic compounds BMC Complementary and Alternative Medicine. 2014.
- 5. Agarwal, M., Chauhan, S. Anti-Mycobaterial Potential of *Crescentia cujete* (Bignoniaceae). International Journal of Advanced Research in Botany (IJARB). 2015, 1 (1), 1-9
- 6. Baliga., M. S., Harshith, P., Bhat, Joseph, N., Fazal, F. Phytochemistry and medicinal uses of the bael fruit (*Aegle marmelos* Correa): A concise review, Food Research International. 2011, 44, 1768–1775.
- Rojas, G., Levaro, J., Tortoriello, J., Navarro, V. Antimicrobial evaluation of certain plants used in Mexican traditional medicine for the treatment of respiratory diseases. Journal of Ethnopharmacology. 2001, 74, 97-101
- 8. Dewi. M.K, Evie, R., Guntur, T. Antibacterial Activity Majapahit Leaf Extract (*Crescentia cujete*) on the Growth Bacterial Wilt Disease Causes Ralstonia solanacearum, Lentera Bio. 2014, 3, 51–57
- 9. Saloso, Y., Jasmindar, Y. Potential of Patikan Kebau (*Euphorbia hirta*) as Antibacterial on Aeromonas hydropilla and Vibrio alginolitycus in Fish Culture. Aquaic Science and Technology. 2014, 2, 1

- 10. Dangeubun, J.L. Phytochemistry And Antibacterial Activity Test From Methanol Extract of *Alstonia Acuminata* Tree Bark Against *Vibrio Harveyi* Bacterium in In-Vitro Manner Journal of Biology and Life Science. 2015, 6, 1
- Harlina H., Prajitno, A., Supriyanto, E., Nursyam H., Rosmiati. Potential Study of Kopasanda (*Chromolaena odorata* L.) Leaves as Antibacterial Against Vibrio harveyi, Disease Causative Agent of Tiger Shrimp (Penaeus monodon Fabricius) Post Larvae. Aquaculture Research & Development. 2015, 6, 10.
- 12. Izzati, M. Screening Potential Antibacterials in Some Species of Seaweed against Bacterial Pathogens at Tiger Shrimp. BIOMA. 2007, 9, 62-67
- 13. Prajitno A. Sensitivity Test Flavonoids Seaweed (*Eucheuma Cottoni*) As Bioactive Natural Against Bacteria Vibrio Harveyi. Protein journal. 2007, 15, 2.
- 14. Rinawati, N.D. Antibacterial Power Plant Majapahit (*Crescentia Cujete* L.) against the bacterium Vibrio alginolyticus. Essay. Not published. Surabaya: Department of Biology, Faculty of Science, Institute of Technology Sepuluh November. 2011.
- Sule W. F., Okonko I. O., Omo-Ogun S., Nwanze J. C., Ojezele M. O., Ojezele O. J., Ali J. A., Soyemi E. T. and Olaonipekun, T. O. Phytochemical properties and *in-vitro* antifungal activity of *Senna alata* Linn. crude stem bark extract. Journal of Medicinal Plants Research. 2011, 5, 176-183.
- 16. Boufadi, Y.M., Soubhye, J., Jean, N., Antwerpen, P.V., Riazi, A. Antimicrobial effects of six Algerian propolis extracts. International Journal of Food Science and Technology. 2016, 51, 2613–2620.
- 17. Mahbub, K. R., Hoq, M. M., Ahmed, M. M., Sarker, A. In vitro antibacterial activity of *Crescentia cujete* and *Moringaoleifera*. Bangladesh Research Publications Journal. 2011, 5, 337 43.
- 18. Parvin, Mst.S., Das, N., Jahan N., Akhter, M.A., Nahar, L., and Islam, Md.E. Evaluation of in vitro anti-inflammatory and antibacterial potential of *Crescentia cujete* leaves and stem bark. BMC Res Note. 2015, 8, 412
- 19. Muliani, R., Susianingsih, E. Activities anti *Vibrio harveyi* Water Extract Mangrove *Sonneratia lanceolata* and toxicity Against Post Larvae Tiger Shrimp *Penaeus monodon*. Proceedings of the Aquaculture Technology Innovation Forum. 2015.
- 20. Balakumar, S., Rajan, S., Thirunalasundari, T., Jeeva, S. Antifungal activity of Aegle marmelos (L.) Correa (Rutaceae) leaf extract on dermatophytes. Asian Pacific Journal of Tropical Biomedicine. 2011, 309-312.
- 21. Harborne, J.B. Chemical methods, Guidance modern way of analyzing the plant. ITB Bandung. 1994.
- Gurinder, J., Kaur, Daljit, S., Arora. Antibacterial and phytochemical screening of *Anethum graveolens*, *Foeniculum vulgare* and *Trachyspermum ammi*. BMC Complementary and Alternative Medicine. 2009, 9. 30
- 23. Obinna N.C., Nwodo S.C., Olayinka O.A. Evaluation of antibacterial activity of *Pisidium guajava* and *Gongronema Latifolium*. Journal of Medicinal Plants Research. 2008, 2(8), pp. 189-192
- 24. Gauger, E., Slomowitz, R., Uhlinger, K., Casey, J., Gomez-Chiarri, M. *Vibrio harveyi* and other bacterial pathogens in cultured summer flounder, paralichthys dentatus. Aquaculture 2006, 260:10-20
- 25. Maftuch, Toban, M.H., Risjani, Y. Administration of marine algae (*Gracilaria verrucosa*) immunostimulant enhances some innate immune parameters in black tiger shrimp (*Penaeus monodon* Fabricus) against *Vibrio harveyi* infection. Journal of Applied Sciences Research. 2012, 8(2): 1052-1058
- 26. Ravikumar, S.M., Gnanadesigan, P., Suganthi, Ramalakshmi, A. Antibacterial potential of chosen mangrove plants against isolated urinary tract infectious bacterial pathogens. International Journal of Medicine and Medical Sciences. 2010, 2(3) pp. 94-99,
- 27. Anyasor, G.N., Aina, D.A., Olushola, M., Aniyikaye A.F. Phytochemical constituent, proximate analysis, antioxidant, antibacterial and wound healing properties of leaf extracts of *Chromolaena Odorata*. Annals of Biological Research, 2011, 2 (2):441-451
- Ery, O., Fukushima, Seki, H., Ohyama, K., Ono, E., Umemoto, N., Mizutani, M., Saito, K., and Muranaka, Y. CYP716A Subfamily Members are Multifunctional Oxidases in Triterpenoid Biosynthesis.Plant Cell Physiol. 2010, 52(12): 2050–2061
- 29. Chung, P.Y., Navaratnam, P., Chung, L.Y. Synergistic antimicrobial activity between pentacyclic triterpenoids and antibiotics against Staphylococcus aureus strains. Annals of Clinical Microbiology and Antimicrobials. 2011, 10:25.

- 30. Krystyna, I., Wolska, Anna, M., Grudniak, Fiecek, B., Kraczkiewicz-Dowjat, A., Kurek, A. Antibacterial activity of oleanolic and ursolic acids and their derivatives. Central European. Journal of. Biologi. 2010, 5(5).543-553
- 31. Leon, de., L., Beltran, B., Moujir, L. Antimicrobial activity of 6-oxophenolic triterpenoids. Mode of action against Bacillus subtilis. Planta Med. 2005, 71:313-319.
- Soetan k. O., Oyekunle M. A., Aiyelaagbe O. O. and Fafunso M. A. Evaluation of the antimicrobial activity of saponins extract of *Sorghum Bicolor* L. Moench. African Journal of Biotechnology. 2006, 5 (23), pp. 2405-2407
- 33. Olaleye, M.T. Cytotoxicity and antibacterial activity of Methanolic extract of *Hibiscus sabdariffa*. Journal of Medicinal Plants Research. 2007, 1(1), pp. 009-013,
- 34. Onwuliri, F.C, Wonang D.L. Studies on the combined antibacterial action of Ginger (*Zingiber officinale* L) and Garlic (*Allium sativum* L) on some bacteria. Nig. J. Bot. 2005, 18: 224-228
- 35. Rojas, F.S., Ojeda, C.B. Recent development in derivative ultraviolet/visible absorption spectrophotometry: 2004–2008 A review. Analytica Chimica Acta. 2009, 635, 22–44
- 36. Ojeda, C.B., Rojas, F.S. Recent applications in derivative ultraviolet/visibleabsorption spectrophotometry: 2009–2011 A review. Microchemical Journal. 2013, 106, 1–16
