

International Journal of ChemTech Research

CODEN (USA): IJCRGG, ISSN: 0974-4290, ISSN(Online):2455-9555 Vol.10 No.2, pp 961-967, 2017

ChemTech

AReview on Vicia faba as a plant test system in toxicity evaluation of various metals: Vicia-micronucleus test (Vicia-MCN)

Sathya Prabhu, Venkat Kumar, Jai Bharath, Mohamed Thaha and Devi Rajeswari*

Department of Biomedical Sciences, School of Biosciences and Technology, VIT University, Vellore – 14, Tamilnadu, India.

Abstract : Toxicity evaluation is a vital practice for evaluating various toxic levels excreted by pollutants. The pollutants were harmful to the environment and it may exhibit serious hazards to the mankind. We need tools to rule out the toxicity levels of pollutants in short period of time. In this regard, Vicia-micronucleus test was a plant test system widely used for evaluating toxicity in various applications. *Vicia faba* plant system is a reliable, easy, sensitive and efficient biological system to carry on genotoxicity and other related studies. Majority of the countries worldwide grow *Vicia faba*, a legume, which has ability to grow in any environmental conditions. However, the damage caused by pollutants to this plant can be treated using breeding analysis. This review article focuses on *Vicia faba's* role in toxicological studies as a test plant system and breeding analysis. **Keywords :** *Vicia faba*; Legume; Vicia-micronucleus test; Toxicity; Pollutants.

Introduction

Since many years, due to industrialization, our environment is polluted by various pollutants like metals ^{1, 2, 3}. Currently, various chemical analysis are available to determine toxic levels, however, reliable biological tests are required to assess the exact damage caused by pollutants⁴. A well know *Vicia*-micronucleus assays (Vicia-MCN) have been used to detect the genotoxic effects of various pollutants, metals, radioactive materials. This simpler method was cheap, easy and widely used in many studies to identify oxidative damage, genotoxic effects and mitotic index (MI) ⁵. The *Vicia faba* (*V. faba*) is common plant grown in major parts of the world. It is a legume, with high nutrition belongs genes *Vicia* and family Fabaceae^{6, 7}. It is also known as Broad bean, Horse bean, Windsor bean, Tick bean, Fava bean. Despite of its origin in the east, the plant was grown all over the world as it is favorable in all climatic conditions ⁸⁻¹¹. Apart from genotoxicity evaluation the *V. faba* is used to evaluate the efficacy of herbicides like glyphosate and Imazapic against *Orobanche crenate* ¹². The foliar evaluation was done using *V. faba* plantas a model¹³⁻¹⁵. Similarly, Gibberellic acid, Uniconazole, Cobalt, α -tochopherol, Citric acid, Oxalic acid effect of growth was tested using *V. faba* plantas a model¹⁶⁻¹⁹.

Genotoxicity can be defined as toxicity to the genome 20 . The legume, *V.faba* plant was widely employed for micronucleus assay by which genotoxicity in various applications can be determined. The test was performed by exposing the *V.faba* plant to pollutants in various concentrations. Further, the plant or plant parts were analyzed for genotoxicity evaluation²¹. This test is standardized by French Member Organization as a prominent test to determine the toxicity ⁴. The toxicity executed by these pollutants can be alleviated by new

breeding techniques. Some of the important genotoxicity evaluation using *V.faba* asplant system and breeding analysis is briefly described in this review.

Cesium genotoxicity evaluation

Vicia-MCN plant bioassay is used to study the genotoxicity of radioactive materials. ¹³⁷Cs is harmful to human beings. DNA damage by ¹³⁷Cs (Radioactive) is assessed by Vicia-MCN and other test methods like tradescantia-micronucleus test (Trad-MCN). In addition, Tradescantia-Stemen-hair mutation test also carried out to show toxic effects of ¹³⁷Cs. In vicia micronucles test (Vicia-MCN) the *V. faba* seeds were soaked in ionized water and it is allowed to germinate at a temperature of 22 degrees centigrade for three days and it is allowed to grow under nourishment of hoagland's solution for four days. This test results showed genotoxicity of ¹³⁷Cs, concluding this method is a sensitive and cost effective bio assay in detecting genotoxicity of ¹³⁷Cs. Likewise, this bioassay can be used to evaluate genetoxicity of various radioactive materials which is contaminated in soil and water ²².

Sulcotrione genotoxicity

Sulcotrione (2-(2-chloro-4-(methylsulfonyl) benzoyl)-1, 3 cyclohexanedione), is an herbicide used to control grasses weed. The genotoxicity of sulcotrione was determined using Vicia-MCN as treatment system. In brief, the *V.faba* seedlings is treated for 45 h with sulcotrione, after treatment for 42h, potent mutagenic effect in hydroponic conditions was observed²³. The study results prove Sulcotrione cause genotoxicity in plants.

Antimutagenic activity of origanum majorana

Origanum majorana (O. majorana) is awidely used medicinal plant in folk medicine, Saudi Arabia. Using Vicia-MCN as a control test system the antimutagenic activity O. majorana was evaluated. In this study, the root tips of V.faba and O. majorana are treated with Sodium azide in concentrations of 250 and 350μ g/ml and 50, 100, 200μ g/ml for 6 h and 20 h, respectively. The study results show no aberrations in sodium azide treated root tips of O. majorana with the control, where no aberrations found. This study is an evidence that O.majorana has anti-mutagenic activity using V.faba plant system as control²⁴.

Arsenic (As) genotoxicity evaluation

Arsenic toxicity through water has been reported in many parts of the world. Arsenic is an element, which is highly toxic to humans. Arsenic toxicity can be evaluated using simple, efficient and reproducible bioassay, Vicia-MCN.It causes skin, bladder, lung, liver cancers, and also serious effects like neurotoxicity, birth defects, and metabolic disorders. When arsenic is given to plant models i.e., Vicia-MCN,modifications in the root color, stiffened roots, and decrease in root length, mitotic index and chromosomic aberrations was observed. In addition, arsenic can cause oxidative stress, cytotoxicity, and genotoxicity in plants^{5, 25}.

Cadmium (Cd) genotoxicity evaluation

Cadmium is a metal, which is present in air, water and soil. Cadmium can induce genotoxity and toxic effects by inducing micronuclei in the root tips of *Vicia* species. Vicia-MCN was carried out in soil spiked with $CdCl_2$ at different concentrations. Experiment was carried out in two terms, short term and long term. In the short term *V.faba* seeds were exposed to soil spiked with $CdCl_2$ at different concentration for a period of five days. Similar treatment was given in long term experiment for a month. Genotoxicity was observed in both short and long term experiments. This study is an indication that micronucleus assay can be used as a marker for determining the genotoxicity of plants. Similar results were found, when *V. faba* is treated with low concentrations of cadmium, which caused genotoxicity and micronuclei was found after 48th hour^{21,26}.

Copper (Cu) genotoxicity evaluation

Copper (Cu) is an essential element for nutrition of humans as well as for plant growth. The toxicity of Cu was determined using Vicia-MCN. Interestingly, *V. faba* was found to be resistance to the toxic effects of Cu. In a study, *V. faba* is collected from heavy metal contaminated soil to evaluate its activity under Cu stress. The improvement of biomass and faba bean resistance to the toxic effects of Cu was noted. Whereas, low levels of Cu shown beneficial effects in *V.faba* plant by promoting growth of the plant. Additionally, antioxidant enzymes such as superoxide dismutase (SOD), catalase (CAT), ascorbate peroxidase (APX) and peroxidase

(POX) was found in high concentrations. Toxicity was observed when the plant was exposed to copper level more than >2mM concentration²⁷. It is important to note the levels of Cu present in soil as it is beneficial to plants in upto a certain concentration (>2mM).

Lanthanum (La) genotoxicity evaluation

Lanthanum (La) is a rare earth metal, can react with nucleoproteins, amino acids, phospholipids, enzymes, intermediate metabolite. In china, La is used in fertilizers for high yield crop production. However, when La was studied using Vicia-MCN test system, reactive oxygen species (ROS), HSP70, Isoenzymes like endopeptidase, superoxide dismutase (SOD), catalase (CA), ascorbate peroxidase (APX), guaiacolperioxidase (GPX) are produced in order to alleviate La-induced oxidative damage. The production of ROS and other enzymes is an indication that La is toxic to plants as well to animals. The toxicity levels of La could be determined by quantitative measurement of these enzymes²⁸.

Lead (Pb) genotoxicity evaluation

Toxicity of lead (Pb) is evaluated using Vicia-MCN. Pb is widely used metal in various applications, but it exerts toxicity to animals and plants. It inhibits plant growth, also causes medical complications to human health and believed to be a possible carcinogen. Pb induced DNA fragmentation was observed at peak levels at 10 M concentration and thereafter the gradual decrease of toxicity was noted. So we can determine Pb was toxic to plants at threshold concentration²⁹.

Aluminium (Al) genotoxicity evaluation

Aluminium (Al) is a metal, which is toxic to plants by inhibiting the growth of roots. The investigation of Al toxicity using Vicia-MCNand *Zea mays* as plant system showed Al was toxic in both dicot and monocot plants. Using Laser Microprobe Mass Analysis (LAMMA) localization of Al was assessed in root tips. Al has reached stele in *Zea mays* and it is confined to outer cortex in *V.faba*. This indicates that Al has stronger affinity in dicots. However, Al was seen intracellular in 60 mins of treatment in both dicots and monocots. So, it is well understood, long term exposure Al can induce toxicity in both monocots and dicots ³⁰.

Selenium (Se) genotoxicity evaluation

Vicia-MCN and sister chromatid exchange (SCE) tests were employed to evaluate selenium toxicity. Selenium is important and essential trace element with anti-mutagenic and anti-cancer properties. Its deficiency may cause certain types of cancers and cardiovascular diseases. But it is conceived to be toxic at higher concentrations and it is toxic to humans and animals. The evaluation of sodium selenite and sodium biselenite showed an increase in MN frequency, SCE frequency and decrease in mitotic index (MI) indicating selenium exhibits genotoxicity ³¹.

Sulfur dioxide (SO₂) genotoxicity evaluation

Yi and meng in 2003 investigated genotoxicity of sulfur dioxide (SO₂), a common air pollutant. SO₂ and its hydrates, bisulfite are investigated using *Allium stavium* and Vicia-MCN bio assay, considering it as simple, and efficient bio assay. In this study, both the plants are treated with the mixture of SO₂hydrates at different concentrations. The results of this study showed SO₂exhibits genotoxicity, increase in Micronuclei (MN) frequencies was observed ³².

Parthenin genotoxicity evaluation

Allelotoxic effects of parthenin, a natural constituent of *Parthenium hysterophorous*. *Parthenium hysterophorous* is a highly spreading weed in India and other parts of the world was studied using Vicia-MCN. In brief, the seeds of *V.faba* are treated with parthenin for 8 h in concentrations of 100,200,300 and 400 IM. It has been observed that high concentration of parthenin exhibits mutagenic and cytotoxic which affects the growth, quality and quantity of the plant, when compared with control³³.

Municipal solid waste (MSW) landfills toxicity evaluation

Worldwide, municipal solid waste (MSW) landfills are widely employed to dispose the waste. In china, landfill leachate was collected from different seasons and evaluated using Vicia-MCN. The MSW samples has exhibited decrease in the mitotic index (MI) and increase in Micronucleus(MN) in *V.faba* root tips was observed. This results provide an evidence MSW could be harmful to plants and humans³⁴.

Silver nanoparticles genotoxicity evaluation

Nowadays silver nanoparticles (AgNPs) is being widely used in various applications such as in health care items, pipe lines, and washing machines as an antibacterial agent ^{35, 36}. In wound dressings and catheters silver nanoparticles are used as therapeutic agents. Even then it is controversial for its toxicological effects ³⁷. Vicia-MCN is carried out to evaluate the genotoxicity of silver nanoparticles. In this assay root tips of *V.faba* is treated with silver nanoparticles in four various concentrations. Finally, the results exposed increased number of chromosomal aberrations (CA), micronuclei (MN) and decreased mitotic index (MI) as a sign of genotoxicity exhibited by the silver nanoparticles. This study confirms that silver nanoparticles is rick for plants and the environment ³⁸. But seed germination is resistance and root length and mitotic cell cycle were susceptible to Ag NPs ³⁹.

Multi-walled carbon nanotubes (MWCNTs) toxicity evaluation

Multi-walled carbon nanotubes (MWCNTs) along with heavy metals were released from industries. These MWCNTs were investigated using Vicia-MCN, in this study, leaves of *V. faba*was damaged may be due to oxidative stress. It is also predicted that MWCNTs may cause phototoxicity and increases the ecological risks in combination with heavy metals. Vicia-MCNis an implemented as a test plant to determine the toxic effects of MWCNTs released from industries ⁴⁰.

Cyano bacteria toxicity evaluation

Microcystin-LR (MCY-LR) is a cyanobacteria toxin, in order to find out the toxity caused by it Vicia-MCN was carried out. The findings of the study show MCY-LR is toxic. The mode action could be by altering mitotic activity and inducing mitotic anomalies in the plant V.faba⁴¹.

Ionic solvent toxicity evaluation

Genotoxicity and cytotoxicity of ionic solvent, 1-butyl-3-methylimadazolium chloride [(C4MIM)Cl] was studied through Vicia-MCN. This evaluation (C4MIM)Cl exhibits genotoxicity and cytotoxicity of 1-butyl-3-methylimadazolium chloride . This study concluded that Ionic solvent1-butyl-3-methylimadazolium chloride could be unsafe to environment and harmful to human beings ⁴².

Toxicity Activation

It is believed that metabolites present in *V.faba* activate the herbicides which cause DNA damage. In a study, evaluation of DNA damage of three herbicides, such as, ametryn, metribuzintriazines, and EPTC thiocarbamate was treated with *V.faba* root metabolism in isolated peripheral human lymphocytes using alkaline single cell gel electrophoresis has been carried out. It is been observed in the study that the DNA damage is caused in human peripheral lymphocytes only when herbicides undergo *V.faba* metabolism. There is no damage was observed in control, which does not undergo *V.faba* metabolism. This study reveals that *V.faba* metabolites plays a key role in genotoxicity of herbicides, which induce DNA damage in isolated human peripheral lymphocytes ⁴³.

Breeding analysis

The damage caused by pollutants to plant *V.faba* can be prevented by developing new resistant varieties using breeding methods. Breeding methods can be improved by applying molecular tools. A whole genome analysis is required; it can be done using reliable markers. About 125,559 SSR (Simple Sequence Repeat) sequences were constructed in 32 varieties of faba bean. SSR novel markers are developed in *V.faba* genome using next generation sequencing. These markers can be used for the expression of genetic maps ¹⁰. In a study, *V.faba* and *trancatula*, both the genomes relationships were studied using different markers and no significant

differences were observed. This genomic relationship data is a central reference for significance in legumes ⁴⁴.As 30% to 90% of *V.faba* yields are affected by aschochyte blight in different environmental conditions and found four genomic controlling resistance regions. It is the need of the hour to develop regions which control the resistance ¹¹.V.faba is studied for specialized nutrient transfer cell (TC) development, its cells are enriched with transporter proteins that enable nutrient transfer. Among legumes, V.faba has largest genome of about 13000 MB. However, only limited amount of genetic information is available. In order to provide a database for future transcript profiling of epidermal TC differentiation this study has been designed to map out de-novo assembly of a genome-wide transcriptome of *V.faba*⁴⁵. The breeding program enhances the quality of faba bean with high nutritional, resistant to stress and pathogens ⁴⁶. In breeding and out breeding studies on five genotypes of *V.faba* shows depression in first cycle selfing (S_1) and it is not observed in second depression (S_2) . RAPD marker analysis showed positive effects, it was not significant to use for evaluating its genetic diversity and in studying faba bean breeding patterns ⁴⁷. The mitochondrial DNA (mtDNA) of *V.faba* is about 588,000 bp. Gene sequences of V.faba were similar to Beta vulgaries, Nicotiana tabacum, Vitisvinifera, Oryza sativa, Zea mays and Medicago truncatula. In which 45% are homologous to Medicago truncatula. This breeding analysis may improve the characters of V.faba genetically, phenologically, morphologically, physiologically, and biochemical traits make it to adapt to target environments with resistance to biotic and abiotic stresses and understanding its genetic control, establishing a desired variety with weed and nutrient management ⁴⁸.

Expression of ClpB/Hsp by faba bean

In drought conditions, temperatures were elevated and less rainfall was reported. Plants growing in these harsh places leads to heat stress⁴⁹.Heat stress ultimately affects cellular, biochemical and physiologically properties of the plant, which affects the growth of the *V.faba* plant ^{50, 51}.Under such conditions Hsps expresses, in this condition protein misfolds and nonfunctional toxic aggregation takes place ⁵². Kumar et al in 2005, initially reported the expression of Hsps in *V.faba*⁵³. The expression of ClpB/Hsp 100 was observed in the leaves, seedlings, mature plants, and pollens of the plant. This experiment was carried out under heat stress conditions and the expression studies of ClpB/Hsp 100 were done using semi-quantitative RT-PCR. The RT-PCR was set at 33°C for 2 h and 4 h by using primers complementary to soya bean genome ⁵⁴.

References

- 1. Babu Nagati, V., Koyyati, R., Marx, P., Devi, V., 2015. Effect of heavy metals on seed germination and plant growth on Grass pea plant (Lathyrus sativus). Int. J. Chem. Tech. Res. 8, 333-345.
- 2. OmPrakash, M., Ashish, G., Mahipatsingh, R.P., 2011. Determination of toxic trace metals Pb, Cd, Ni, and Zn in soil by Polarographic Method. Int. J. Chem. Tech. Res. 3, 599-604.
- 3. Roya, A.Q., Akram, A., 2016. Impact of toxic heavy metals in food systems: A systemic Review. Int. J. Chem. Tech. Res. 9, 422-429.
- 4. Foltête, A.S., Dhyèvre, A., Férard, J.F., Cotelle, S., 2011. Improvement of Vicia-micronucleus test for assessment of soil quality: a proposal for international standardization. Chemosphere. 85, 1624-1629.
- 5. Wu, L., Yi, H., Yi, M., 2010. Assessment of arsenic toxicity using Allium/Vicia root tip micronucleus assays.J Hazard Mater. 176, 952-956.
- 6. EL-Qudah, J.M., 2015. Vitamin A Contents Per Serving of Eleven Foods Commonly Consumed in Jordan . Int. J. Chem. Tech. Res. 8, 83-88.
- 7. Akpinar, N., Akpinar, M.A., Türkoğlu, S., 2001.Total lipid content and fatty acid composition of the seeds of some *Vicia* L. species. Food Chem. 74, 449-453.
- 8. Darwish, D., Shrief, S., Fahmy, G., Madany, M., Saleh, Al-Juboori, R.M., 2016. Influence of three soil moisture levels on early growth and proline content of some faba bean genotypes. Int. J. Chem. Tech. Res. 9, 188-197.
- 9. Azaza, M., Wassim, K., Mensi, F., Abdelmouleh, A., Brini, B., Kraïem, M., 2009.Evaluation of faba beans (*Vicia faba* L. var. minuta) as a replacement for soybean meal in practical diets of juvenile Nile tilapia Oreochromis niloticus. Aquacult. 287, 174-179.
- 10. Chillo, S., Civica, V., Iannetti, M., Mastromatteo, M., Suriano, N., Del Nobile, M., 2010.Influence of repeated extrusions on some properties of non-conventional spaghetti. J food Eng. 100, 329-335.
- 11. Kaur, S., Kimber, R.B., Cogan, N.O., Materne, M., Forster, J.W., Paull, J.G., 2014.SNP discovery and high-density genetic mapping in faba bean (*Vicia faba* L.) permits identification of QTLs for ascochyta blight resistance. Plant Sci. 217, 47-55.

- 12. El-Rokiek, K.G., El-Metwally, I., Messiha, N.K., Amin, S.E.D.S., 2015. Controlling Orobanche crenata in Faba bean using the Herbicides Glyphosate and Imazapic with some additives. Int. J. Chem. Tech. Res. 8, 18-26.
- 13. Manal, F.M., Thalooth, A., Amal, G.A., 2012.Performance of Wheat Plants in Sandy Soil as Affected by Foliar Spray of Potassium and Zinc and their combination. Chem Anal.Int. J. Chem. Tech. Res. 9, 715-725.
- 14. Khattab, E., Badr, E.A., Afifi, M.,2016. Response of Some Varieties of Faba bean (Vicia faba L.) to Boron and Potassium.Int. J. Chem. Tech. Res. 9, 98-103.
- 15. Shafeek, M., Ali, A.H., Asmaa, R.M., 2016.Foliar application of amino acids and bio fertilizer promote execution of broad bean plant (Vicia faba L) under newly reclaimed land conditions. Inter. J. of Chem. Tech. Res. 9, 100-109.
- 16. Mekki, B.E.D., 2016. Growth and Yield of Mungbean (Vigna radiataL.) in Response to Gibberellic Acid and Uniconizole Foliar Application. Inter. J. of Chem. Tech. Res. 9, 100-109.
- 17. El-Metwally, I., A.E. El-Saidy. Physiological and chemical response of faba bean to water regime and cobalt supplement in sandy soils. Inter. J. of Chem. Tech. Res. 9, 76-82.
- 18. Talaat, I.M., El-Wahed, M.A., El-Awadi, M., El-Dabaa, M., Bekheta. M., 2015. Physiological Response of Two wheat Cultivars to α-tochopherol.Inter. J. of Chem. Tech. Res. 9, 342-350.
- 19. Sadak, M.S., Orabi, S.a., 2015.Improving thermo tolerance of wheat plant by foliar application of citric acid or oxalic acid.Inter. J. of Chem. Tech. Res. 8, 333-345.
- 20. Dhyèvre, A., Foltête, A.S., Aran, D., Muller, S., Cotelle, S., 2014.Effects of soil pH on the Viciamicronucleus genotoxicity assay.Mutat Res Genet Toxicol Environ Mutagen. 774, 17-21.
- 21. Béraud, E., Cotelle, S., Leroy, P., Férard, J.F., 2007.Genotoxic effects and induction of phytochelatins in the presence of cadmium in *Vicia faba* roots.Mutat Res Genet Toxicol Environ Mutagen. 633, 112-116.
- 22. Minouflet, M., Ayrault, S., Badot P.M., Cotelle, S., Ferard, J.F., 2005. Assessment of the genotoxicity of 137Cs radiation using *Vicia*-micronucleus, Tradescantia-micronucleus and Tradescantia-stamen-hair mutation bioassays. J Environ Radioact. 81, 143-153.
- 23. Sta, C., Ledoigt, G., Ferjani, E., Goupil, P., 2012.Exposure of *Vicia faba* to sulcotrione pesticide induced genotoxicity.Pestic Biochem Phys.103, 9-14.
- 24. Qari, S.H., 2008.In vitro evaluation of the anti-mutagenic effect of *Origanum majorana* extract on the meristemetic root cells of *Vicia faba*.J Taibah Univ Sci.1, 6-10.
- 25. Duquesnoy, I., Champeau, G.M., Evray, G., Ledoigt, G., Piquet-Pissaloux, G., 2010.Enzymatic adaptations to arsenic-induced oxidative stress in *Zea mays* and genotoxic effect of arsenic in root tips of *Vicia faba* and *Zea mays*. Comptes rendus biologies. 333, 814-824.
- 26. Foltête, A.S., Masfaraud, J.F., Férard, J.F., Cotelle, S., 2012.Is there a relationship between early genotoxicity and life-history traits in *Vicia faba* exposed to cadmium-spiked soils? Mutat Res Genet Toxicol Environ Mutagen. 747, 159-163.
- 27. Fatnassi, I.C., Chiboub, M., Saadani, O., Jebara, M., Jebara, S.H., 2015.Impact of dual inoculation with Rhizobium and PGPR on growth and antioxidant status of *Vicia faba* L. under copper stress. Comptes rendus biologies. 338, 241-254.
- Ning, W., Chengrun, W., Xia, B., Yueyun, L., Liumin, T., Huixian, Z., Xiaorong, W., 2011.Toxicological effects and risk assessment of lanthanum ions on leaves of *Vicia faba* L. seedlings. J Rare Earths. 29, 997-1003.
- 29. Pourrut, B., Jean, S., Silvestre, J., Pinelli, E., 2011.Lead-induced DNA damage in *Vicia faba* root cells: potential involvement of oxidative stress.Mutat Res Genet Toxicol Environ Mutagen. 726, 123-128.
- 30. Marienfeld, S., Schmohl, N., Klein, M., Schröder, W.H., Kuhn, A.J., Horst, W.J., 2000.Localisation of aluminium in root tips of Zea mays and *Vicia faba*. J Plant Physiol. 156, 666-671.
- 31. Yi, H.,Si. L., 2007. *Vicia* root-mirconucleus and sister chromatid exchange assays on the genotoxicity of selenium compounds. Mutat Res Genet Toxicol Environ Mutagen. 630, 92-96.
- 32. Yi, H.,Z. Meng. 2003.Genotoxicity of hydrated sulfur dioxide on root tips of *Allium sativum* and *Vicia faba*.Mutat Res Genet Toxicol Environ Mutagen. 537, 109-114.
- 33. Raoof, K.A., Siddiqui, M., 2013. Allelotoxic effect of parthenin on cytomorphology of broad bean (*Vicia faba* L.). J Saudi Soc. Agri. Sci. 12, 143-146.
- 34. Sang, N.,Li., G., 2004.Genotoxicity of municipal landfill leachate on root tips of *Vicia faba*.Mutat Res Genet Toxicol Environ Mutagen. 560, 159-165.

- 35. Chen, X., Schluesener, H., 2008.Nanosilver: a nanoproduct in medical application. Toxicol lett. 176, 1-12.
- 36. Tripathi, A., Chandrasekaran, N., Raichur., A.M., Mukherjee, A., 2009. Antibacterial applications of silver nanoparticles synthesized by aqueous extract of *Azadirachta indica* (Neem) leaves. J Biomed. Nanotechnol. 5, 93-98.
- 37. AshaRani, P., Low, K.M.G., Hande, M.P., Valiyaveettil, S., 2008.Cytotoxicity and genotoxicity of silver nanoparticles in human cells. ACS nano. 3, 279-290.
- 38. Patlolla, A.K., Berry, A., May, L., Tchounwou, P.B., 2012.Genotoxicity of silver nanoparticles in *Vicia faba*: a pilot study on the environmental monitoring of nanoparticles. Int J Environ Res Public Health. 9, 1649-1662.
- 39. Abdel-Azeem, E.A., Elsayed., B.A., 2013. Phytotoxicity of silver nanoparticles on *Vicia faba* seedlings. New York Sci J. 6, 148-156.
- 40. Wang, C., Liu, H., Chen, J., Tian, Y., Shi, J., Li, D., Guo, C., Ma, Q., 2014.Carboxylated multi-walled carbon nanotubes aggravated biochemical and subcellular damages in leaves of broad bean (*Vicia faba* L.) seedlings under combined stress of lead and cadmium. J hazard. mater. 274, 404-412.
- 41. Beyer, D., Tándor, I., Kónya, Z., Bátori, R., Roszik, J., Vereb, G., Erdődi, G., Vasas., G Márta, M., Jambrovics, K., 2012.Microcystin-LR, a protein phosphatase inhibitor, induces alterations in mitotic chromatin and microtubule organization leading to the formation of micronuclei in *Vicia faba*. Annals of botany. 110, 797-808.
- 42. Liu, T., Zhu, L., Wang, J., Wang, J., Xie, H., 2015. The genotoxic and cytotoxic effects of 1-butyl-3methylimidazolium chloride in soil on *Vicia faba* seedlings. J Hazard. mater. 285, 27-36.
- 43. Calderón-Segura, M.E., Gomez-Arroyo, S., Molina-Alvarez, B., Villalobos-Pietrini, R., Calderon-Ezquerro, C., Cortés-Eslava, J., Valencia-Quintana, P.R., López-González, L., Zúñiga-Reyes, R., Sánchez-Rincón, J., 2007.Metabolic activation of herbicide products by *Vicia faba* detected in human peripheral lymphocytes using alkaline single cell gel electrophoresis. Toxicol. In Vitro. 21, 1143-1154.
- 44. Ellwood, S.R., Phan, H.T., Jordan, M., Hane, J., Torres, A.M., Avila, C.M., Cruz-Izquierdo, H., Oliver, R.P., 2008.Construction of a comparative genetic map in faba bean (*Vicia faba* L.); conservation of genome structure with Lens culinaris. BMC genomics. 9, 1-11.
- 45. Arun-Chinnappa, K.S., McCurdy., D.W., 2015.De novo assembly of a genome-wide transcriptome map of *Vicia faba* (L.) for transfer cell research. Front Plant Sci. 6.1-9.
- 46. Rispail, N., Kaló, P., Kiss, G.B., Ellis, T.N., Gallardo, K., Thompson, R.D., Prats, E., Larrainzar, E., Ladrera, R., González, E.M., 2010.Model legumes contribute to faba bean breeding. Field Crops Res. 115, 253-269.
- 47. Obiadalla, A.H.A., Mohamed, N.E., Khaled, A.G., 2015.Inbreeding, outbreeding and RAPD markers studies of faba bean (*Vicia faba* L.) crop. J Advan. Res. 6, 859-868.
- 48. Singh, A.K., Bhatt, B., Upadhyaya, A., Kumar, S., Sundaram, P., Singh, B.K., Chandra, N., Bharati, R., 2012.Improvement of faba bean (*Vicia faba* L.) yield and quality through biotechnological approach: A review. African J Biotech. 11, 15264-15271.
- 49. Stoddard, F., Balko, C., Erskine, W., Khan, H., Link, W., Sarker, A., 2006.Screening techniques and sources of resistance to abiotic stresses in cool-season food legumes. Euphytica. 147, 167-186.
- 50. Adisarwanto, T., Knight, R., 1997.Effect of sowing date and plant density on yield and yield components in the faba bean. Australian J Agri. Res. 48, 1161-1168.
- 51. Sarkar, N.K., Kim, Y.K., Grover, A., 2014.Coexpression network analysis associated with call of rice seedlings for encountering heat stress. Plant Mol. Bio. 84, 125-143.
- 52. Lavania, D., Siddiqui, M.H., Al-Whaibi, M.H., Singh, A.K., Kumar, R., Grover, A., 2015.Genetic approaches for breeding heat stress tolerance in faba bean (*Vicia faba* L.). Acta Physiologiae Plantarum. 37, 1-9.
- 53. Kumar, R., Singh, A.K., Lavania, D., Siddiqui, M.H., Al-Whaibi, M.H., Grover, A., 2015.Expression analysis of ClpB/Hsp100 gene in faba bean (*Vicia faba* L.) plants in response to heat stress. Saudi Journal of Biological Sciences. 23, 243-247.
- 54. Negruk, V., 2013.Mitochondrial genome sequence of the legume *Vicia faba*. Frontiers in plant science. 4, 1-11.