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Assessment of Children under than Five Ages toward the Diarrheal Cases with Antibacterial Effect of bacteria isolates in Babylon Province, Iraq

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Abstract : Background: Diarrheal disease is an infection caused by the presence and growth of microorganism in the intestine. **Objectives:** This study aims at determining the common organism that causes diarrhea in children less than 5 years and detection the most effective antimicrobial agents and plant extract that causative agents of diarrhea and select the best antibiotics to treat it. **Methods:** One hundred stool sample was collected from children have watery diarrhea in Al-Qassim hospital; during the period (January to April 2015). The microbial isolate was identified tested for antibacterial agents. **The results:** Sixty case From 100 samples have a bacterial infection (60%) represented by **E. coli** (75%) and **Salmonella** spp.(25%)).

Regarding Antibiotic sensitivity test, *E. coli* and *Salmonella* spp. showed highly sensitive to Imipenem (88.8%), (100%) respectively, followed by Amikacin (100%) for *E. coli* and Ciprofloxacin (100%) for *Salmonella* spp., while Rifampin were not effective against tested isolates, whereas the plants extract result revealed that lemon extract more effect on diarrheal bacteria than other one.

Conclusions: in this study I conclude that rate of bacterial diarrheal infection among children younger more than 5 years was high. *E. coli* was predominant isolate and impeneim was the most effective antibiotics on bacterial isolates. Among plants extract lemon were more effect on diarrheal bacteria

Keywords: Antibacterial effect, Diarrheal Children antibiotics and plant extracts.

Introduction:

The Diarrhea define according the Organization of World Health (WHO) as having three or more loose or stools liquid per day, which can be caused by a variety of parasitic, viral, bacterial and organisms. Infection is spread through contaminated water and food, and from patient to patient (fecal oral rout) as a result of poor hygiene.

Diarrhea remains one of the leading causes of mortality and morbidity among children, which causing more than 1 billion episodes of illness and 3-4 million deaths annually.

The major causes of diarrheal illness include, among others, limited access to or poor quality of poor food, and water hygiene and sanitation¹. The mechanisms of transmission of diarrheal pathogen are person to person through the fecal oral route or by ingestion of contaminated water or food ². The bacterial pathogens usually responsible for diarrheal illness include *Shigella*, *Escherichia coli*, *Aeromonas*, *Campylobacter Salmonella*, and *Yersinia*, ¹.

There are many risk factors that increase susceptibility to infection with enteric bacteria including; young age, immune deficiency, lack of breast feeding, malnutrition and others²⁹. Gastrointestinal tract of breastfed infant is colonized more easily with micro flora than are those of formula fed infants. Colonization is through to reduce infant diarrhea by inhibition of the growth of pathogenic organism³,³⁵.

The reason for the extensive use of plants as drugs may be the fact that medicinal plants are available, wide range of medicinal plants and their distinct form and thus are procured without any trouble. Plants extracts have a long history of use and their use is widespread in both developing and underdeveloped countries ⁴. The medicinal plants are very important to eradication of disease due to multidrug resistance bacteria and the plant extract without side effect to human ^{36, 37}.

This study aims at determining the common organism that causes diarrhea in children less than five years and detection the most effective antimicrobial agents and plant extract against the causative agents of diarrhea and select the best antibiotics to treat it and to identify the care level of diagnosis of bacteria.

Materials and methods:

Patients and Sample collection: A total of 100 specimens of stool were collected from children younger than five years suffering from diarrhea (watery diarrhea) in Al-Qassim hospitals in Babylon province, during a period of three months (January to April 2015). Sample ofFecal was collected from each patient. The sample plated onto MacConkey agar and incubated anaerobically and aerobically at 37 °C overnight.

Plants collection:

Fresh fruits (Pomegranate, banana and lemon) were obtained from the Hilla market, Iraq, 2015. It washed in running distill water in laboratory, sterilized the surface with 70% alcohol.

Extracts of Pomegranate peels were prepared as 50% extract, hot water was primed, peel were added clean container and left to cool, the container was covered in order to keep all active elements. The mixtures were vigorously swirled by the blender. The mixtures were filtered using filter paper into a clean beaker.

After **Bananapeels**, sterilized, then peel was taken. Distilled water was boiled, peels were added to the distilled water and left to cool.

These contents were mixed by the blender and filtered to remove the large, UN homogenized particles to get clear aqueous extract, while **lemon** were cut open with a knife (sterile) and the juice pressed out into a sterile container and then filtered by filter paper (0.45Millipore) into another container (sterile) to remove the tissues plants and used as crude (freshly) without refrigeration. The Extract was stored at four °C until use.

Citrus lemon(dried fruits) extracts were according to 5,22.

Spice Collection and Extraction:

The plant material (Curcuma, cinnamon and Sumac extract) were brought from markets of Babylon Province. Ten grampowders of dried plant were boiled in 500 mL of distilled water 6,34

Methods:

After positive results of growth were appear, the bacteria recapture on EMB(eosin methylen blue agar) and SS(*Salmonella shigella* agar) then identified with Gram stain and Biochemical test⁷.

Antimicrobial susceptibility testing:

Susceptibility to antibacterial agents for all isolates were determined by the diffusion of standard disk method on Muller-Hinton agar incubated for twenty hours at 37 °C. The selection of discs of antibiotic was performed according to the guidelines recommended by ¹. The following disks of antibiotic were used as table 1. After twenty hours, the diameter of each zone of inhibition was measured with a pair of calipers& recorded in mm. The results then interpreted according to CLSI documentation ⁸.

In vitro antibacterial activities of plants extract testing using Agar diffusion well assay.

The screening of antimicrobial activities of each extract on the tested bacteria used in this investigation was determined on Muller Hinton agar media, by the agar diffusion techniques using agar well diffusion method. Six mm diameter of wells and 5 mm depth were made on the solid agar using a sterile glass borer 9,10,23,33. Approximately 20µl of each extract was inoculated onto wells were made in the pouring culture of each microorganism isolates. The plate was then allowed to incubate at 37°C for 18 hrs. After 18 hrs of incubation, each extract was zone of inhibition for all isolates. The diameters of the zone of inhibitions were measured by millimeter (mm).

Antibiotic disc	Symbol	Potency	Zone diameter nearest who		
	Symbol	Totaley	Sensitive	Medium	
Azithromycin	AZM	15 μg	>18	14-17	

Table 1:-Inhibition Zone of Different Antibiotics According To CLSI

ole mm Resistant <13 ΑK >17 15-16 <14 Amikacin 30µg CTX 30µg >23 15-22 <14 Cefotaxime С >18 13-17 <12 Chloramphenicol 30 µg CN 10μg >15 13-14 <12 Gentamicin FOX 30µg >18 15-17 <14 Cefoxitin 17-19 RA >20 <16 5 μg Rifampin IPM >16 14-15 <13 **Imipenem** 10 μg CIP >21 16-20 <15 Ciprofloxacin 5 μg

Results and discussion:

Rate of infection:

The rates of bacterial infection among diarrheal children were (60%) the rest (40%) maybe due to parasite or viral infection.

Bacterial isolates:

This study show that *E. coli* was predominant isolate in 75%, while *Salmonella* spp. in 25% as shown in fig (1). *E. coli* is the most commonly recognized bacterial pathogen in infantile gastroenteritis ¹¹.

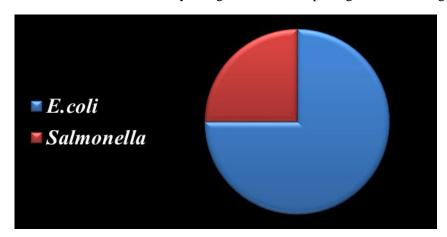


Fig (1): Rate of bacterial isolates among diarrheal children

Age group:

The high infection of diarrhea was recorded in age groups from 1 month to 1 year (table 2).

They are most groups exposed to this disease. The reason of higher infection in infants less than 6 months may be due to low immunity, as the amount of trance placental antibodies of the child starts dwindling after 6 months of age.

Those infants also may not have been breast-fed but bottle- fed instead, which is a source of infection and contamination ¹².

Table (2): Number of diarrheal infections according to children's age

Age(years)	No. of infections	%
lmonth tol year	40	40
1-2	10	10
2-3	20	20
3-4	25	25
4-5	5	5
Total	100	100

Antibiotics sensitivity:

In this study, Antibiotics sensitivity was studied. *Salmonella* spp. and *E. coli* showed highly sensitive to Imipenem(100%) and (88.8%) respectively. This results in agreement with ¹³, and ¹⁴ and ¹⁵ in Iraq, this study demonstrated that all (100%) *E. coli* isolates were susceptible to Imipenem (Fig: 2). Imipenem showed promising results to treat enteric fever ¹⁶. The high efficiency of these drugs may be the usage rarely in studied area and high cost and non-availability in oral forms ¹² and ¹⁶.

Moreover, the result of sensitivity of Amikacin and Ciprofloxacin were studies. The bacterial isolates were highly sensitive to Amikacin and and Ciprofloxacin (100%) for *E. coli* and (100%) for Salmonella spp. (fig 2). Amikacin show more effect on *E. coli* in 100% Sensitive rate that result similar to Baghdad study (CLSI, formerly NCCLS, 2010) and Ciprofloxacin is the first-line drug for the treatment of enteric fever ¹⁷while the Rifampin were not effective against tested isolates. The reason of high resistance of antibiotics in area of study might be to the overuse of drug in the treatment of diarrhea could lead to an increase of antibiotic resistance ¹².

Antibacterial activity of plant extracts on diarrheal bacteria:

On the other hand, the present study investigated of the **antibacterial activity** of some plant products {Curcuma,cinnamon, pomegranate peel, Limon juice,lemon (dry fruit) and banana peel} against diarrheal bacterial isolates as result shown as **fig:3**.

The results indicate that the **Curcuma** aqueous extract have **antibacterial activity** against *Escherichia* coli and *Salmonella* ¹⁸ and ⁶.

Also **cinnamon** reveal antibacterial effect against bacterial isolates more than other spice on bacterial isolates. Several studies ¹⁹ and ³¹ have shown that cinnamon oil had good inhibitory effects against various pathogens as we found. The antibacterial activity has been due to the presence of active constituents in the oils. The study revealed cinnamaldehyde to be the major constituent of cinnamon oil ³⁰.

Furthermore, the Sumac extract has less effect against bacterial isolates and its action of inhibiting gram negative bacteria related to its contain active component like alkaline and alcoholic toxic materials which inhibit and kill microorganism by destroy plasmic membrane, act on nucleic acid and protein denaturation ²⁰. This result agree with ²¹ in effect on inhibition growth of *E.coli*.

Moreover, the antimicrobial activities of juice Limon and lemon (dry fruit) on bacterial isolates were studied. **Limon juice** reveals high antibacterial activity of this extract against bacterial isolates; 30 and 35mm for E.coli and salmonella respectively. That result similar to ²²which found high antibacterial activity on gram

negative bacteria. Antibacterial activities of lemon (dry fruit) on bacterial growth were 22 and 28 mm for *E.coli* and *Salmonella* respectively. This could be due to the acidic pH of this juice that will affect the charges of the amino acids that constitute the peptidoglycan, and it may affect the active sites of enzymes leading to defect in their activity ²³.

Moreover, the result show that the **pomegranate peel extract** have active action on bacterial growth were 30 mm to both studied bacteria. This result agree with 24 who found that antibacterial effect on E.coli at 20mm and disagree with 22 which not found any effect of pomegranate peel on bacterial growth.

The consumption of **banana** was high contributed to nutritional value; its shell has been studied for treatment of gastrointestinal disorders ²⁵additionally, ²⁶ who found banana peel gel inhibited the growth of enterobactericea. These studies support this result when found banana juice were effect on diarrheal bacterial growth at (18, 16 mm) to *E.coli* and *Salmonella* respectively.

The prevalence of resistance antibiotic is a continual problem due to the evolution of a potent defense mechanism against antibiotics.

Therefore, it is necessary to exploit and develop a novel inhibitory agent against those bacteria ²⁷. Plants and plant products have been used extensively throughout history to treat medical problems. Numerous studies have been carried out to plant extract various natural active products for screening antimicrobial activity ²⁸.

The medicinal plants are very important to eradication of disease due to multidrug resistance bacteria and the plant extract without side effect to human^{36, 37}.

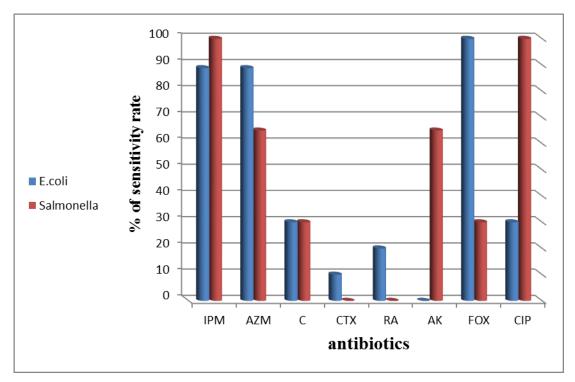


Fig 2: Sensitivity Rat of E. coli and Salmonella to different Antibiotics

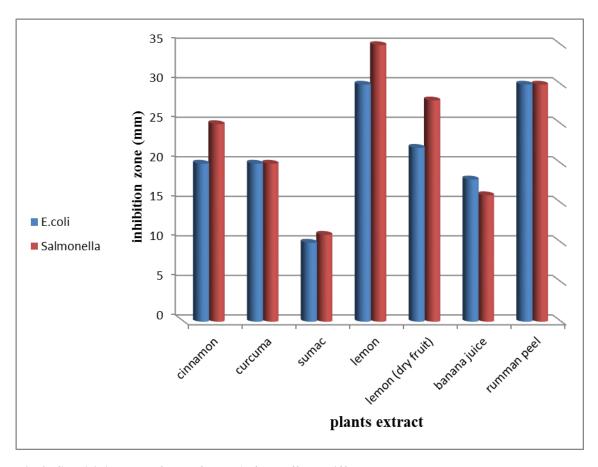


Fig 3: Sensitivity Rat of E. coli and Salmonellat o different plants extract

Conclusion:

In this study we conclude that rate of bacterial diarrheal infection among children younger than 5 years was high. *E. coli* was predominant isolate and impeneim was the most effective antibiotics on bacterial isolates. Among plants extract lemon were more effect on diarrheal bacteria.

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References:-

- 1. Clinical and Laboratory Standards Institute (CLSI, formerly NCCLS), Performance standards for antimicrobial susceptibility testing, Seventeenth informational supplement. 2010, 27 (1).
- 2. Richard E, Bchrman MD, Robert Kliegman M, Hal MD, Jenson, B, . Nelson Textbook of pediatrics, Elsivier, 18th Ed.,2007
- 3. Wolin, MJ, ZhangY, Bank, Yerry S, Miller, TL, NMR detection of Nifedido bacterium fermentation in the intestinal tract. J. Nutr. 1999, 128(1): 91-6.
- 4. Kinghorn AD,Kaneda N, Baek NI, Kennelly EJ, SoejartoJahan M, Warsi M.K., Khatoon F.D. Concentration influence on antimicrobial activity of banana blossom extract-incorporated chitosan-polyethylene glycol (CS-PEG) blended film. J Chem Pharm Res, 2010, 2: 373-378.
- 5. Hindi, N. Antimicrobial Activity of Different Aqueous Pomegranate (Rumman) Extracts against Different Human Pathogens. International Journal of Indigenous Medicinal Plants, 2013, 46: (2)
- 6. Niamsa N. and Sittiwit, C. Antimicrobial activity of Curcuma longa aqueous extract. J. pharmacol.Toxicol. 2009,4(4):173-177.

- 7. MacFaddin, JF. Ed. Biochemical Tests for Identification of Medical Bacteria. 3rd ed. Philadelphia: Lippincott. Williams and Wilkins. 2000.
- 8. Al-Hilli, Z. B. Dissemination of β-lactamases in *Escherichia coli* and *Klebsiella* spp.isolated from Merjan teaching hospital in Hilla city. M. Sc.Thesis. Kufa University, College of Science, 2010.
- 9. NCCLS (National Committee for Clinical Laboratory Standards), Methods for dilution antimicrobial susceptibility tests of bacteria that grow aerobically. Approved Standard M100-S12. Wayne. PA, NCCLS, 2002.
- 10. Prescott LM, Harley J. and Klein, DA. Microbiology 5th.ed, McGraw-Hill NewYork, 2002,809-811
- 11. Khan MM, Igbal J, Chafour N, and Burney M.T., 1988. Etiological agents in diarrial disease in hospitalized children in Rawalpindi, Pakistan. Journal Diarrhoeal disease Res, 6:228-31.
- 12. Shamki J, Al-Charrakh A, Al-Khafaji J. Detection of ESBLs in Enteropathogenic E. coli (EPEC) Isolates Associated with Infantile Diarrhea in Kut City. Medical Journal of Babylon, 2012, 9(2).
- 13. Hadi Z J. Detection of extended-spectrum beta-lactamases of *Escherichia coli* and *Klebsiella* spp. isolated from patients with significant bacteriuria in Najaf. M. Sc. Thesis. College of Medicine.Kufa University, 2008.
- Al-Hilali SAM. Occurrence and Molecular characterization of Enteropathogenic *Escherichia coli* (EPEC) Serotypes isolated from children with diarrhea in Najaf. M. Sc. Thesis. University of Kufa, College of Medicine, 2010.
- 15. Qaisr S, SeemaIrfan E, Khan, TanwirAhsan A. In Vitro Susceptibility of typhoidal Salmonellae against newer antimicrobial agents: A search for alternate treatment options. JPMA 2011, 61:462.
- 16. Kadhim SA, Hassan L, Shoukat D. Antimicrobial susceptibility patterns against Escherichia coli and prevalence of extended–spectrum β-lactamases. Mustansiriya Medical Journal, 2011, 10 (1).
- 17. John T, Jeremy A, Skinner and Linda R Ward. Detection of decreased *in vitro* susceptibility to ciprofloxacin in *Salmonella enterica* serotypes Typhi and Paratyphi A. J. Antimicrob. Chemother.2001,48 (5): 740-741.
- 18. Gupta S. and SRavishankar. A comparison of the antimicrobial activity of garlic, ginger, carrot and turmeric pastes against Escherichia coli O157: H7 in laboratory buffer and ground beef. Foodborne.Pathog. Dis., 2005, 2: 330-340.
- 19. Livak KJ, and TD.Schmittgen. Analysis of relative gene expression data using real-time PCR and the 2_CT method. Methods, 2001, 25:402–408.
- 20. Cowman MM. Plant products as antimicrobial agents. Clin.Microbial. Rev. 1999, 12 (4): 564-582.
- 21. Alwan BH, Esmaal S, Alwan AH. effect of crude extract of sumac plant and inhibition of bacterial attachment which isolated from patients with urinary tract infection, 2008.
- 22. Hindi NKK and Chabuck ZA. Antimicrobial Activity of Different Aqueous Lemon Extracts. Journal of Applied Pharmaceutical Science, 2013, 3 (06): 074-078.
- 23. Abdullah NY. Effect of some plant extracts against Staphylococcus aureus and Klebsiellapneumoniae. Iraqi academ SC J, 2009, 1(2): 32-36.
- 24. Dahham SS, Ali N, Tabassum H. and Khan M. Studies on Antibacterial and Antifungal Activity of Pomegranate (*Punica granatum* L.). American-Eurasian J. Agric. & Environ. Sci., 2010, 9 (3): 273-281
- 25. Pannangpetch P, Vuttivirojana A, Kularbkaew C, Tesana S, Kongyngyoes B, Kukongviriyapan V. The antiulcerativeeffectof Thai Musa species in rats.Phytother Res, 2001, 15, 407-10.
- 26. Lino PB, Corrêa CF, Archondo ME, Dellovam DC. Evaluation of post-surgical healing in rats using a topical preparation based on extract of Musa sapientumepicarp. Braz J Pharmacogn, 2011, 21, 491.
- 27. Cabello FC. Heavy use of prophylactic antibiotics in aquaculture: a growing problem for human and animal health and for the environment. Environ Microbiol, 2006, 8: 1137-1144.
- 28. Nita T, Arai T. and Takamatsu H. Antibacterial activity of extracts prepared from tropical and subtropical plants on methicillin-resistant Staphylococcus aureus. J Health Sci. 2002, 48: 273-276.
- 29. Richard J.Schanler. The pediatrics clinic of America, feeding, part II, The management of Breast feeding, Guest Ed, 2001.
- 30. Prabuseenivasan S, Jayakumar M, and Ignacimuthu S.In vitro antibacterial activity of some plant essential oils. BMC Complementary and Alternative Medicine.2006, 6:39
- 31. Mah TC., and TooleGAO. Mechanism of biofilm resistance to antimicrobial agents. Trends BHM BBNJJ NMicrobiol. 2001, 9:34–39.
- 32. Hindi N K H, Yasir A A AL-MAHDI ZKA, Jebur MH. Evaluation of Anti-Bacterial Activity: Anti adherence, Anti Biofilm and Anti Swarming of the Aquatic Extract of Black Raisins and Vinegar of Black Raisins in Hilla City, Iraq. International Journal of PharmTech Research. 2016; 9(9): 271-280.

- 33. Hindi N K H. In vitro Antibacterial activity of Plants extracts against Porphyromonas gingivalis, Prevotella intermedia and Aggregatibacter actinomycetem comitans Streptococcusmutanus, Isolated from Periodontitis Patients in Babylon province, Iraq. International Journal of PharmTech Research. 2016;9(10).
- 34. Ibrahim M E. Essential oils Isolated From Leaves of Egyptian Verbena triphylla L Herb Using Different Extraction Methods. International Journal of PharmTech Research. 2016, 9(4):01-07.
- 35. Hemaia M, Motawe L, Ibrahim F M, Ibrahim ME, Mahmoud EA, Aly H F. Isolation and Identification of Terpenoids and Sterols of Nepetacataria L. International Journal of PharmTech Research. 2015, 8(10): 10-17.
- 36. Hafiz I, Silalahi RJ. Antioxidant and Anti-inflammatory Activity of Pagoda Leaves (Clerodendrum paniculatum L.)Ethanolic Extract in White Male Rats (Rattus novergicus), International Journal of PharmTech Research. 2016, 9 (5):165-170.
- 37. Manoppo H, Magdalena EF, Kolopita, Rotina Malatunduh Growth promoter effect of garlic (Allium sativum) on carp (Cyprinus carpio L), International Journal of PharmTech Research, 2016,9(4): 283-288,

