

Co-Expressional Protein Products of BRCA-1 and EBV-EBNA-1 Genes in Tissues from Human Female Patients with Breast Cancers: An Immunohistochemical Screening Study

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Abstract : Back ground: Breast malignancies are most frequently diagnosed among women in many populations world-wide as well as in Iraq. Although the genetic mutations in BRCA -1 and BRCA - 2 genes are still constituting up to 90% of the total risk for breast cancers, yet many indirect evidences are supporting a role for an association of EBV with such cancers.

Objective:To analyze the rate of EBV infection in the breast tissues in association with defects and / or mutations in BRCA-1 gene, by assessing the endogenous levels of the total expressed BRCA-1- as well as EBNA-1 protein products, and their relations to the differentiation of primary invasive breast cancer tissues.

Patients and methods: Fifty-four (54) formalin-fixed, paraffin- embedded breast tissues were obtained in this study; (34) biopsies from breast cancers (BC) and (20) from apparently normal breast autopsies as a control group. Detection of protein expressional products of EBNA-1 gene of Epstein Barr Virus as well as BRCA-1 gene was done by HRP/DAB immune-enzymatic antigen detection system using specific rabbit-anti-human primary antibodies for EBV-EBNA -1as well as detected or mutated BRCA-1 protein products.

Results: Detection of EBNA-1 - immunohistochemical (IHC) reactions in tissues with BC was observed in 12 out of 34 (35.3%), while in healthy breast tissues in the control group was detected in 10% (2 out of 20). Detection of BRCA-1- protein- immunohistochemical (IHC) reactions in tissues with BC was observed in 16 out of 34 (47.1%), while none of the examined healthy breast tissues in the control group revealed such IHC-reactions. The difference between the percentages of BRCA-1- as well as EBNA1 proteins detection in BC tissues & control group was statistically significant (<0.05). Among breast cancer tissues that showed score I of IHC reactions for BRCA1, 68.8% have well differentiated grade; and 18.7% of those tissues that have score II- IHC reactions showed moderate differentiated grade and lastly, 12.5% of the BC tissues which showed score III have presented as poorly differentiated BC tissues. However, statistical significant differences between the frequencies of EBV-EBNA1 and BRCA-1- immunohistochemical reactions were neither observed in relation to the age of these breast cancer patients nor to the grade of invasive breast cancer tissues (P value > 0.05).

Conclusions: Our results indicate that the EBV might contribute to the development of subset of breast tumors. The present results of the rates of defects or mutations in the BRCA-1- genes in relation to the grade of breast cancer tissues also could point for their occurrence and contribution as early events in breast carcinogenesis.

Keywords : Breast cancer; Epstein Barr Virus; EBV - EBNA 1; Defects / Mutations; BRCA-1- Gene; Immunohistochemical technique.

Introduction:

Globally, breast cancers are the most frequent malignancies that affect women¹⁻³. In Iraq, Iraqi Cancer Board as well as Iraqi Cancer Registry Center in Iraqi Ministry of Health has recently demonstrated that female breast cancers constituted about 25% out of the total registry of cancers in Iraqi patients⁴.

A variety of reproductive and hormonal factors have been identified in the etiology of breast cancer constituting together fifty percent among all the etiologies of breast carcinogenesis and as such, the research works were prompted to unravel other risk factors in that critical issue⁵.

Genetic factors were noticed to play a recognized role in less than 5% of breast cancer leading to hereditary breast- ovarian cancer syndrome⁶. However, and on exclusion the risk of the familial history, the risk of breast carcinogenesis is significantly increased in relation to the occurrence of some mutations, particularly in BRCA-1 and BRCA-2 genes⁷.

Among human tumor suppressor genes, the BRCA-1 and BRCA-2 genes are ubiquitously found in all humans where their genes and proteins are called breast cancer type 1 and 2 susceptibility genes and proteins, respectively⁸.

Certain variations of the *BRCA1* gene lead to an increased risk for breast cancer as part of a hereditary breast - ovarian cancer syndrome. Researchers have identified hundreds of mutations in the *BRCA1* gene, many of which associated with an increased risk of cancer. Women with an abnormal BRCA1 or BRCA2 gene have up to an 80% risk of developing breast cancer by age 90; increased risk of developing ovarian cancer is about 55% for women with BRCA1 mutations⁹.

Approximately 50% to 65% of women who have a deleterious mutation in *BRCA1* could develop breast cancer by age 70, and 35% to 46% could develop ovarian cancer by age 70. Approximately 40% to 57% of women with a deleterious mutation in *BRCA2* will develop breast cancer by age 70, and 13% to 23% will develop ovarian cancer by age 70(10).

Viruses are involved in the development of various cancers¹¹. In 1995, the Epstein-Barr virus (EBV), and ubiquitous herpes virus, was found in 21% of 91 breast cancers¹².

Since then, a large number of studies have detected EBV infection in patients with breast carcinoma. A series of studies that adopted non-breast-cancer control groups have also been performed^{12,13}, and several mechanisms and hypotheses about the association between EBV infection and breast carcinoma have been developed^{11, 14,15}.

Some researchers believed that EBV infection may play a role in the early stages of breast carcinogenesis and elevate breast cancer risk¹⁶. Moreover, EBV infection might be a latent factor in the development of certain types of breast carcinoma (15). However, statistical data from studies have varied widely. This inconsistency could be largely attributable to several problems: technical challenges in detecting and localizing the EBV in tumor cells, study designs that involved a specific histological type of breast carcinoma, and the lack of an epidemiological perspective that could clarify the inconsistencies in EBV prevalence across studies¹⁷.

Epstein-Barr virus (EBV) is a large double-stranded DNA virus that is classified as a gamma-1 herpes virus of the lymphocrypto- genus. EBV has infected greater than 90% of the world's population and is the etiologic agent of infectious mononucleosis¹⁸.

In the growth program of EBV expresses all the nine known latent proteins: the six EBV nuclear antigens (EBNA1, EBNA2, EBNA3A, EBNA3B, EBNA3C, and EBNA3L), three latent membrane proteins (LMP1, LMP2A, and LMP2B), and the non-polyadenylated EBV RNAs (EBERs)¹⁹.

In the default program, EBNA1, LMP1, LMP2A, and the EBERs are expressed, providing necessary signals that are thought to allow infected lymphoblasts to differentiate into memory B cells²⁰. In the latency program, which has a much more restricted pattern of viral gene expression, very few viral genes are expressed. The transcript for LMP2A has been consistently detected, and recent reports suggest the EBERs are also expressed. This low level of viral gene expression allows persistence of the virus in resting recirculating memory cells in a way that is nonpathogenic and not detectable by the immune system²¹.

Production of infectious virus is the essential feature of fourth and final gene expression program found in humans latently infected with EBV²⁰.

This study was proposed to unravel the rates of both EBV infection as well as defects and / or mutations in BRCA-1 gene in the breast tissues and their relations to the differentiation grades of primary invasive breast cancer tissues.

Materials and Methods:

Study Groups:

This study was designed as a retrospective research; a number of (54) formalin-fixed, paraffin embedded breast tissue blocks enrolled in this study which comprised both patients and control samples that their age ranged from 32 to 71 years. These retrospective paraffin-embedded samples were retrieved from the archives of the period from 2011 till 2016 belonging to major hospitals and private histopathological laboratories in Kerbela, Babylon, and Al-Najaf provinces. The diagnoses were based on their accompanied pathological reports of the corresponding patients. These blocks included a group of (34) biopsies from patients who had undergone surgical operation or biopsies for their breast cancers (BC) and (20) autopsies from apparently normal breast tissues control group. These breast tissues were properly subjected to fixation as well as paraffin embedding and used for this research work as an age- and grade- matched groups.

Laboratory methods:

Slide Preparation:

Tissue sectioning was conducted following trimming process of the tissue blocks at the histopathological department of Teaching laboratories / Al- Sadar Medical City (Al- Najf) & Al- Hilla hospital teaching and a second confirmatory histopathological re-evaluation of each obtained tissue blocks was done by a consultant pathologist. One paraffin embedded (4 mm) thick-tissue section was prepared and mounted on ordinary glass slide and stained with hematoxyline and eosin, while other (4 mm) thick-tissue sections were stuck onto positively charged slide to be used for detection of EBV-EBNA 1 & BRCA1 – antigen using Mouse and Rabbit Specific HRP/DAB (ABC) Detection IHC kit (Lot. Number: ab64264) that was purchased from (Abcam, UK) , an immunoenzymatic antigen detection system for immunohistochemistry techniques, using specific Rabbit Monoclonal primary Anti-EBV Nuclear Antigen antibody [E1-2.5] (ab8329) [Lot. Number: [E1-2.5] ab8329], was also purchased from (Abcam, UK) and BRCA1 [Lot. Number: (ab191042)], also purchased from (Abcam, UK) .The details of methods for performing IHC reaction with these antibodies were conducted according the instructions of that manufacturing company, and were done in the Research Laboratories of the Clinical Communicable Diseases Research Unit, at College of Medicine, University of Baghdad as well as in the Advanced Microbiology Research Laboratory at College of Science, University of Babylon.

Histopathological Analysis:

According to the specification of the kit, proper use of this IHC detection system gives an intense brown signal at specific sites of the expression protein in positive test tissues (by using light microscope).

The signal was evaluated under light microscopy using $\times 100$ lens for counting the positive cells. The IHC results were given intensity and percentage scores based on intensity of positive signals and number of cells that gave these signals, respectively.

Positive cells were counted in 10 different fields of 100 cells for each sample and the average percentage of positive cells within the 10 fields was determined. A scale of 0-3 was used for relative intensity with 0 corresponding to no detectable IHC reaction, and 1, 2, 3 equivalents to low, moderate, and high intensity of reaction respectively. Cases were assigned to one of the following percentage score categories: 1%–25% (score 1), 26%–50% (score 2) or > 50% (score 3)²².

Statistical Analysis:

T test, ANOVA test, and Chi square were applied for statistical examination of results obtained in our research. All these statistical analysis were done by using Pentium-4 computer through the SPSS program (version-19) and Excel application.

Results

Archival specimens enrolled in this study were related to female patients with breast cancer whom mean age was (52.7+ 8.5 years) while the mean age of those who have apparently healthy tissues was (62.7 + 7.2 years) (Table 1).

Table (1): Studied groups according to the mean age

Study Groups	N	Mean (years)	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
Patients	34	52.7	8.5	1.5	49.5	55.9	38.00	69.00
Control	20	62.7	7.2	2.2	58.1	67.3	54.00	76.00
Total	54	55.6	9.3	1.5	52.7	58.4	38.00	76.00
t-test for Equality of Means								
		Independent Samples Test:					95% Confidence Interval of the Difference	
		t	df	Sig. (2-tailed)	Mean Difference	Std. Error Difference	Lower	Upper
Age		-3.579	40	.001	-10.00000	2.79444	-15.64777	-4.35223

The signals of EBV-EBNA1 immunohistochemical (IHC) reactions were detected as brown discoloration at the antigenic sites that were detected by their specific primary antibodies (Figure 1). Table (2) shows the positive results of EBNA 1 - IHC reactions, where 35.3% (12 of total 34) breast cancers showed positive signals while 2 out of 20 (10%) in control group has presented such positive signals for IHC test. While, the highest percentage of EBNA 1 - IHC score signaling (50%: 6 out of 12 cases) was found in the moderate score (score II), whereas 41.7% (5 out of 12 cases) and 8.3% (1 out of 12 cases) were found within low (score I) and strong (score III) scores, respectively. Statistically, significant differences ($p < 0.05$) were found on comparing the percentage of EBNA1 in the BC group according to their positive signal scoring.

Table (2): Signal scoring of EBNA 1- immunohistochemical reactions among the breast cancers tissues.

P	Normal Breast Tissues (n=20)		Malignant Breast Tumors (n=34)		EBNA-1 Signal scoring	
	%	N	%	N		
0.001 significant	80	18/20	64.7	22/34	Negative	
	10	2/20	35.3	12/34	Positive	
	100	2	41.7	5/12	I	Scoring
	0	0	50	6/12	II	
	0	0	8.3	1/12	III	

The highest percentage of EBNA 1 signal intensity in the present study was (41.7%: 5 out of 12 cases) that is related for each weak and moderate intensity, while 16.6% (2 out of 12) of the breast cancers tissues were presented with strong intensity. Significant statistical differences ($p < 0.05$) were found among breast cancers tissues according to their EBNA 1- scoring intensities (Table3).

Table (3): Signal Intensities of Positive EBNA1- IHC Reactions.

Chi-square Tests	Negative EBNA1 signaling	Signal Intensity			positive EBNA1 signaling	Studied Groups
		High	Moderate	Weak		
<0.001 significant	22/34 (64.7%)	2/12 (16.6%)	5/12 (41.7%)	5/12 (41.7%)	12/34 (35.3%)	Malignant Br. Tumors (n=60)
	18/20 (80%)	0/2 (0.00%)	1/2 (50%)	1/2 (50%)	2/20 (10%)	Healthy Br. Tissues (n=20)

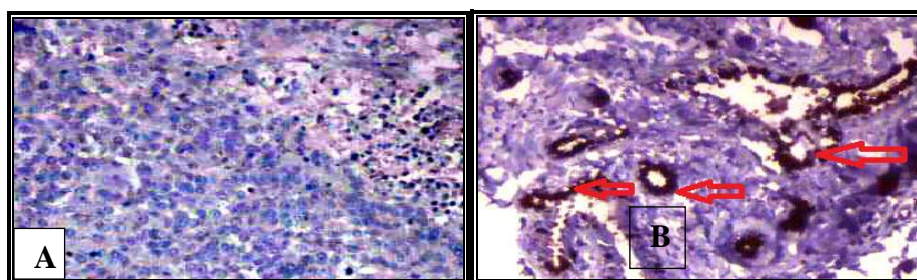


Figure (1): Infiltrative Breast Carcinoma Showing The Results of Immunohistochemistry Staining Protein Over Expression Using Biotinylated Anti-EBNA1 Protein Antibody; Stained By DAB-Chromogen (Brown) and Counter Stained By Mayer's Heamatoxylin (Blue).

A- Breast Cancer with negative staining with EBNA1 .

B-EBNA1-IHC-reaction with high signal score and strong signal intensity (4x)

It was noticed that 12 cases out of 34 (35.3%) of EBNA1-IHC test showed positive EBNA1-IHC reactions .It was found that 66.7% of breast cancer tissues that showed score I of IHC reaction for EBNA1 have well differentiation; and 75% of breast tissues that have score II of IHC reaction for EBNA1 have moderate differentiation. Lastly, 60% of the BC tissues cases that showed score III have presented with poor differentiation. Statistically, the overall EBNA1 –IHC scoring on comparing to tumor grading showed non- significant differences ($P>0.05$) Table (4) .

Table (4): The correlation of EBNA1-IHC score with grading of breast carcinoma

P	Breast Cancer Grades						EBNA1 score	
	Poorly differentiated (n=15)		Moderately differentiated (n=11)		Well differentiated (n=8)			
	%	N	%	N	%	N		
0.1[N.S]	66.7	10/15	63.6	7/11	62.5	5/8	Negative	
	33.3	5/15	36.4	4/11	37.5	3/8	Positive	
	20	1/5	25	1/4	66.7	2/8	I	Scoring
	20	1/5	75	3/4	33.3	1/8	II	
	60	3/5	0.00	0/4	25.0	0/8	III	

The signaling results of immunohistochemical reactions for BRCA-1 antigenic detection were observed as brown discoloration at the specific antigenic sites of these reactions with their specific primary antibodies Figure (2).

The positive- signal results of BRCA-1 - immunohistochemical reactions were found in 47.1% % (16 out of total 34) breast cancers while no tissue in the control group has showed such IHC signals. The statistical Pearson Chi-Square analysis shows significant difference between the patients and control groups regarding BRCA-1 immunohistochemical results (<0.05) Table (5).

Table (5): Frequency of BRCA-1 immunohistochemical reactions among the study groups

			GROUP			Total	Valid Percent	Cumulative Percent	
The Marker			Patients	Control					
BRCA-1	Positive	Count	16	0	16				
		%	47.1%	0.0%	33.3%	47.1	47.1		
	Negative	Count	18	20	38				
		%	52.9%	100.0%	66.7%	52.9	100.0		
	Total	Count	34	20	54				
		%	100.0%	100.0%	100.0%	100.0			
				Value	df	Asymp. Sig. (2-sided)			
BRCA-1	Pearson Chi-Square		8.400	1	.004				

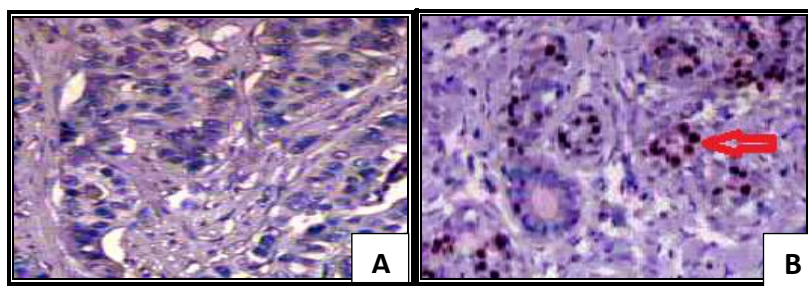


Figure (2): Infiltrative Breast Carcinoma Showing The Results of Immunohistochemistry Staining Protein Over Expression Using Biotinylated Anti-BRCA1 Protein Antibody; Stained By DAB-Chromogen (Brown) and Counter Stained By Mayer's Heamatoxylin (Blue).

A- Breast Cancer with negative staining with BRCA1 .

B-BRCA1-IHC-reaction with moderate signal score and strong signal intensity (4x)

In the present study, 16 tissues out of 34 showed positive EBV-EBNA 1 -IHC reactions .It was found that 62.5% of breast cancer tissues that showed score I of IHC reaction for EBNA- 1 have well differentiation; and 63.6% of breast tissues that have score II of IHC reaction for EBNA 1 have moderate differentiation. Lastly, 26.7% of the BC tissues cases that showed score III have presented with poor differentiation. Statistically, the overall EBNA 1- IHC scoring on comparing to tumor grading showed non- significant differences ($p>0.05$) Table (6).

Table (6):Correlation of EBNA 1 -IHC scoring with the grading of breast cancers .

P	Breast Cancer Grading						State Of Signal Scoring	
	Poorly differentiated (n=15)		Moderately differentiated (n=11)		Well differentiated (n=8)			
	%	N	%	N	%	N		
0.54 [N.S]	73.3	11/15	36.4	4/11	37.5	3/8	Negative	Scoring
	26.7	4/15	63.6	7/11	62.5	5/8	Positive	
	25	1/4	71.4	5/7	20	1/5	I	
	25	1/ 4	14.3	1/7	80	4/5	II	
	50	2/ 4	14.3	1/7	0.00	0/5	III	

Discussion

Breast cancers have ranked the top of the commonest ten cancers in the Iraqi provinces and districts, accounting for about one third of the registered female cancers²³. The role of EBV in breast cancer etiology is still controversial. Unraveling the relationship of EBV in such cancer is potentially important as to a better understanding of breast carcinogenesis as well as for early detection and the prevention of such cancer²⁴.

Reviewing the 34 breast cancers patients included in this study, it was found that their age was ranging between 38-69 years and their mean age was 52.8 + 8.6 years. These results are consistent with those Iraqi as well as world-wide reported results which have found that breast malignant tumors are usually affecting females aged over forty years²⁵⁻²⁷.

Generally, aging is a risk factor that increase the possibility of malignant changes noticed in breast epithelial tissues which was found to increase with age. The present data could also point for the importance of such risk factor in the tumorigenesis of the studied BC tissues and might be related to the effect of long exposure of these breast epithelial tissues to the hormonal changes²⁸.

In the current study ,the results have shown EBV-EBNA1 – IHC reactions positivity in 35.3% (12 out of 34 cases) of those breast tissues obtained from a randomly collected Iraqi patients with breast cancers , while it was detected in only 2 out of 20 (10%) of breast tissues in the control group. The importance of such findings was their relevance and in line with many observations which are indirectly supporting an association of EBV with breast cancer : (a) EBV is detected in breast milk of some women²⁹ transfection of EBV DNA stimulates growth of human breast milk cells³⁰; (c) some EBV-associated lymphomas have occurred in breast^{31,32}; (d) epidemiological similarities of breast cancer to young-adult Hodgkin's lymphoma³³; (e) EBV detection in benign breast tumors in immunosuppressed women³⁴; (f) EBV-lymphoblastoid cell lines can directly infect breast epithelial cells on contacting them and (g) serological evidence of anti-EBNA-1 antibodies in stored sera of breast cancer Indian women³⁵.

IARC Working Group has classified EBV as a group-1 carcinogen³⁶. However, the associated cancers vary markedly in their viral prevalence (nearly 100% in nasopharyngeal carcinoma to about 10% in gastric carcinomas)^{37,38}.

The patterns of viral genes expression also differ, suggesting that EBV may affect cell growth in more than one way³⁹. Although in the current study the detected EBV infection could represent an important step in carcinogenesis, yet is not a sufficient one , and as such we believe that an additional epidemiological risk factors, and stated by Sally L. Glaser *et al.*⁴⁰, could have played a critical role in this process. However, our results are comparable to the results reported by Zekriet *et al.*,²⁴ where PCR for EBNA1 has detected EBV infection in 23 (57.5%) out of those 40 studied Egyptian cases and in 16 (32%) out of those 50 studied Iraqi cases with invasive breast carcinoma.

The current finding is in close to the previous study done by Joshi *et al.*⁴¹ where they found that about 55% of breast cancer Indian women cases showed EBNA-1 expression in tumor cells by IHC, while all the controls

with benign breast disease were negative. The results of the current study are also compatible with and / or consistent to the findings of numerous studies, as those done by Chu *et al.*⁴²: 25%, Murray *et al.*⁴³: 31%, Preciado⁴⁴: 38/102 (37%), Fawzyet *al.*,⁴⁵: 10/40(25%), Lorenzetti *et al.*⁴⁶: 22/71(31%) of EBNA-1 expression in breast cancer cells by IHC.

However, the present findings are lower than a study done by Bonnet *et al.*,⁴⁷ where they found that 9/9 (100%) of EBNA-1 expression in breast cancer cells by PCR. In addition, The obtained results in the present study are in disagreement with the study done by Brink *et al.*,⁴⁸ who found that about 2% (2 out of 115 cases) EBNA-1 expression in tumor cells by IHC.

Lin *et al.*,⁴⁹ demonstrated that EBV infection promotes tumorigenic activity of breast cancer cells. It is suggested entry of EBV into epithelial cells involves an epithelial-specific receptor or cell–cell contact with infected lymphocytes .

In the present study, the results have shown BRCA-1- protein- IHC reactions positivity in 16 out of 34 (47.1%), while none of the examined healthy breast tissues in the control group revealed such IHC- reactions. The BRCA-1 gene spans approximately 80 kb of genomic DNA region make it relatively unstable and prone to deletions and rearrangements (50). Indeed, 36% of mutations in Dutch families are due to three different, large genomic deletions in BRCA-1(51), and duplication of exon 13 has been reported in a BCLC family linked to BRCA1.

Several studies have compared survival of BRCA1-associated breast cancer to that of BRCA1-negative cases with conflicting results^{52,53}.

However, the pathological features of BRCA1-associated tumors would predict a poorer survival in this group of patients. The mutation prevalence estimates may also be biased by preferential survival between cases with mutations and those without. For many oncologists it is not yet possible to draw evidence-based conclusions about the association between *BRCA1* and/or *BRCA2* mutation carrier ship and breast cancer prognosis, relating that to the heterogeneity of the reported results which precluded a conclusion regarding the contribution of *BRCA1/2* status and tumor features to a worse survival. However, primary breast cancer treatments may be different for *BRCA1* and *BRCA2* mutation carriers compared to non-carriers, mostly related to different pathological features of tumors in carriers (S10 Supporting Information, part A)^{54,55}. In addition, therapy response of tumors in *BRCA1/2* mutation carriers might be better compared to that in non-carriers⁵⁶.

In the current study, it was found 68.8% of breast cancer tissues that revealed BRCA1 mutation have well grades, followed by moderate grade (18.7%) and poor grade (12.5%). In this study, the percentage of BRCA1 was found to decrease with the proceeding of the grading of breast cancer. Their BRCA1-negative BC counterparts tissues were found to have a similar decreasing trend of grades of BC.

The grade of BRCA1-related breast cancers has been shown to be consistently elevated with grade 3 comprising 61.3–81.5% of cases versus 22–27% of hospital-based comparison series^{57,58}. In this report, 65% of tumors from BRCA1 patients were high grade. Although a higher proportion of grade 3 tumors has been reported for BRCA1- related breast cancers^{57,58}, survival has been reported as comparable or better^{58,59}. Additionally, hereditary breast cancer presents at a slightly younger age⁶⁰, and a young age has been demonstrated to be an independent adverse risk factor⁶⁰⁻⁶³. Thus, grade 3 in BRCA1-related breast cancer may have a different implication for prognosis than in other breast cancer cases⁶⁴.

According Bordeleau and colleagues review⁶⁵, the overall prognosis of *BRCA*-associated breast cancer was similar to that of breast cancer not associated with *BRCA* mutations. For studies published in the 1990s, several methodological limitations have led to an inconclusive results whereas for more recently published studies, with an improved methodology, still failed to demonstrate a significant overall survival difference. In 2010, Lee and colleagues⁶⁶, found that *BRCA-1* mutation carriers had significantly lower short-term and long-term overall survival rates (OSR) relative to non-carriers while both short-term and long-term OSR of *BRCA2* carriers did not differ from non-carriers.

On the basis of systematic and evidence-based analysis of all studies published to date, to explain the large heterogeneity between the reported results, surprisingly, only two factors seem to explain part of the heterogeneity; misclassification bias; when a study had not tested the comparison (‘non-carriers’) group, and the

proportion of incident cases (S11 Supporting Information, panels C and D). Other reasons for the large heterogeneity and generally weak associations observed might be population differences (i.e. different mutations), differences in completeness of follow-up (often not reported), differences in consideration of contralateral breast cancer and prophylactic surgeries (usually not reported). Publication bias is unlikely to play a large role, because of the low prevalence of *BRCA1/2* mutations in populations as well as included published studies with only a small number of carriers⁶⁷.

From our results we can conclude that EBV might contribute to the development of subset of breast tumors. The present results of the rates of defects or mutations in the *BRCA-1*- genes in relation to the grade of breast cancer tissues also could point for their occurrence and contribution as early events in breast carcinogenesis.

References

1. Dumitrescu RG, Cotarla I. Understanding breast cancer risk – where do we stand in 2005? *J Cell Mol Med*. 2005;9:208–221
2. Globocan (2008) International Agency for Research on Cancer, WHO, Lyon, IARC Press, 2010..
3. Glaser SL, Hsu JL, Gulley ML. Epstein-Barr virus and breast cancer: state of the evidence for viral carcinogenesis. *Cancer Epidemiol Biomarkers Prev* 2004;13:688–97
4. Alwan N (2010). Breast Cancer: Demographic Characteristics and Clinicopathological Presentation of Patients in Iraq. *Eastern Mediterranean Health Journal*, 16:1073–1078.
5. Familial breast cancer: collaborative reanalysis of individual data from 52 epidemiological studies including 58,209 women with breast cancer and 101,986 women without the disease. *Lancet*. 2001; 358:1389–1399.
6. Miki Y, Swensen J, Shattuck-Eidens D, et al. A strong candidate for the breast and ovarian cancer susceptibility gene *BRCA1*. *Science*. 1994;266(5182):66–71. [PubMed]
7. Walsh T, Lee MK, Casadei S, et al. Detection of inherited mutations for breast and ovarian cancer using genomic capture and massively parallel sequencing. *Proceedings of the National Academy of Sciences of the United States of America*. 2010;107(28):12629–12633. [PMC free article] [PubMed]
8. Pal T, Permuth-Wey J, Betts JA, et al. *BRCA1* and *BRCA2* mutations account for a large proportion of ovarian carcinoma cases. *Cancer* 2005; 104(12):2807–16.
9. NanciePetrucelli , Mary B Daly, and Gerald L Feldman.(2013).*BRCA1* and *BRCA2* Hereditary Breast and Ovarian Cancer.Copyright © 1993-2016, University of Washington, Seattle.GeneReviews is a registered trademark of the University of Washington, Seattle. All rights reserved.
10. Serraino D, Piselli P, Scognamiglio P. Viral infections and cancer: epidemiological aspects. *J BiolRegulHomeost Agents*.2001;15:224–228.
11. Labrecque LG, Barnes DM, Fentiman IS, Griffin BE. Epstein-Barr virus in epithelial cell tumors: a breast cancer study. *Cancer Res*. 1995;55:39–45.
12. Bonnet M, Guinebretiere JM, Kremmer E, Grunewald V, Benhamou E, et al. Detection of Epstein-Barr virus in invasive breast cancers. *J Natl Cancer Inst*. 1999;91:1376–1381.
13. Subramanian C, Robertson ES. The metastatic suppressor Nm23-H1 interacts with EBNA3C at sequences located between the glutamine- and proline-rich domains and can cooperate in activation of transcription. *J Virol*.2002;76:8702–8709.
14. Trabelsi A, Rammeh S, Stita W, Mokni M, Mourou A, et al. Detection of Epstein-Barr virus in breast cancers with lymphoid stroma. *Ann BiolClin*. 2008;66:59–62.
15. Yasui Y, Potter JD, Stanford JL, Rossing MA, Winget MD, et al. Breast cancer risk and “delayed” primary Epstein-Barr virus infection. *Cancer Epidemiol Biomarkers Prev*. 2001;10:9–16.
16. Hsu JL, Glaser SL. Epstein-barr virus-associated malignancies: epidemiologic patterns and etiologic implications. *Crit Rev OncolHematol*.2000;34:27–53.
17. Levin LI, Munger KL, Rubertone MV, Peck CA, Lennette ET, Spiegelman D, et al. Multiple sclerosis and Epstein-Barr virus. *JAMA*.2003;289(12):1533–6.
18. Nanbo A, and Takada K. The role of Epstein-Barr virus-encoded small RNAs (EBERs) in oncogenesis.*RevMed Virol*2002; 12: 321-326.

19. Deshpande, S Badve, N Kidwai and R Longnecker .(2002). Lack of Expression of the Epstein-Barr Virus (EBV) Gene Products, EBERs, EBNA1, LMP1, and LMP2A, in Breast Cancer Cells. *Lab Invest*, 82:1193–1199.
20. Thorley-Lawson D.A. EBV persistence in vivo. Invading and avoiding the immune response. In: Medveczky P.G., Friedman H., Bendinelli M., editors. *Herpesviruses and Immunity*. Plenum Press; New York: 1998. pp. 207–229.
21. Blancato J, Singh B, Liu A, Liao DJ and Dickson RB : Correlation of amplification and overexpression of the c-myc oncogene in high-grade breast cancer: FISH, in situ hybridization and immunohistochemical analyses. *British Journal of Cancer* (2004) 90, 1612 – 1619.
22. Iraqi Cancer Board; Iraqi Cancer Registry Center: Results of Iraqi Cancer Registry. Iraqi Ministry of Health (editor). Baghdad. Iraq. 2012: Pp.44 and 131.
23. Zekri AR, Bahnassy AA, Mohamed WS, El-Kassem FA, El-Khalidi SJ, Hafez MM, Hassan ZK.(2012). Epstein-Barr virus and breast cancer: epidemiological and molecular study on Egyptian and Iraqi women. *J Egypt NatlCanc Inst.*;24(3):123-31.
24. Al-Alwan NA. DNA proliferative index as a marker in Iraqi aneuploid mammary carcinoma. *East Mediterr Health J*. 2000 Sep-Nov ;6(5-6):1062–72.
25. Elkum N, Dermim S ,Ajarim D ,et al ; Being 40 or younger is an independent risk factor for relapse in operable breast cancer patients :The Saudi Arabia experience *BMC Cancer* ., 2007.
26. Al-Khafaji K, Attu G. and AL –Kurri j .Pathological staging of breast cancer .A study of 220 patients. *The Iraqi post graduate medical journal*. 2004; January. 3(1). P:38-4.
27. Pike MC, Spicer DV, Dahmouch L, Press MF. Estrogens, progestogens normal breast cell proliferation, and breast cancer risk. *Epidemiol Rev*. 1993;15(1):17–35.
28. A.K. Junker, E.E. Thomas, A. Radcliffe, R.B. Forsyth, A.G. Davidson, L. Rymo Epstein-Barr virus shedding in breast milk *Am J Med Sci*, 302 (1991), pp. 220–223
29. S.A. Xue, I.A. Lampert, J.S. Haldane, J.E. Bridger, B.E. Griffin Epstein-Barr virus gene expression in human breast cancer: protagonist or passenger? *Br J Cancer*, 89 (2003), pp. 113–119
30. R. Giardini, C. Piccolo, F. Rilke Primary non-Hodgkin's lymphomas of the female breast *Cancer*, 69 (1992), pp. 725–735
31. S.H. Abhyankar, K.Y. Chiang, J.P. McGuirk, *et al*. Late onset Epstein-Barr virus-associated lymphoproliferative disease after allogeneic bone marrow transplant presenting as breast masses *Bone Marrow Transplant*, 21 (1998), pp. 295–297
32. Y. Yasui, J.D. Potter, J.L. Stanford, *et al*. Breast cancer risk and “delayed” primary Epstein-Barr virus infection *Cancer Epidemiol Biomarkers Prev*, 10 (2001), pp. 9–16
33. C.G. Kleer, M.D. Tseng, D.E. Gutsch, *et al*. Detection of Epstein-Barr virus in rapidly growing fibroadenomas of the breast in immunosuppressed hosts. *Mod Pathol*, 15 (2002), pp. 759–764.
34. Joshi, M. Quadri, N. Gangane, R. Joshi, N. Gangane. Association of Epstein Barr Virus Infection (EBV) with Breast Cancer in Rural Indian Women. *Plos One*, 4 (12) (2009), p. e8180.
35. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Epstein - Barr virus and Kaposi's sarcoma *Herpesvirus/Human Herpesvirus 8*. Lyon: IARC; 1997.
36. J.L. Hsu, S.L. Glaser Epstein-Barr virus-associated malignancies: epidemiologic patterns and etiologic implications *Crit Rev Hematol Oncol*, 34 (2000), pp. 27–53
37. M.L. Gulley Review Molecular diagnosis of Epstein-Barr virus-related diseases *J Mol Diagn*, 3 (1) (2001), pp. 1–10
38. G. Niedobitek, N. Meru, H.J. Delecluse . Epstein-Barr virus infection and human malignancies. *Int J Exp Pathol*, 82 (2001), pp. 149–170.
39. Sally L. Glaser, Joe L. Hsu and Margaret L. Gulley. (2004). Epstein-Barr Virus and Breast Cancer: State of the Evidence for Viral Carcinogenesis. *Cancer Epidemiol Biomarkers Prev* 2004;13(5).
40. Joshi, M. Quadri, N. Gangane, R. Joshi, N. Gangane. Association of Epstein Barr Virus Infection (EBV) with Breast Cancer in Rural Indian Women. *Plos One*, 4 (12) (2009), p. e8180
41. Chu, K.L. Chang, Y.Y. Chen, W.G. Chen, L.M. Weiss. No significant association of Epstein-Barr virus infection with invasive breast carcinoma. *Am J Pathol*, 159 (2001), pp. 571–578.
42. Murray PG, Lissauer D, Junying J, Davies G, Moore S, et al. (2003) Reactivity with A monoclonal antibody to Epstein-Barr virus (EBV) nuclear antigen 1 defines a subset of aggressive breast cancers in the absence of the EBV genome. *Cancer Res* 63: 2338–2343.
43. Perrigoue JG, den Boon JA, Friedl A, Newton MA, Ahlquist P, et al. (2005) Lack of association between EBV and breast carcinoma. *Cancer Epidemiol Biomarkers Prev* 14: 809–814.

44. Fawzy S, Sallam M, Awad NM (2008) Detection of Epstein-Barr virus in breast carcinoma in Egyptian women. *ClinBiochem* 41: 486–492.
45. Lorenzetti, E. De Matteo, H. Gass, P. Martinez Vazquez, J. Lara, P. Gonzalez, M.V. Preciado, P.A. Chabay. Characterization of Epstein Barr virus latency pattern in Argentine breast carcinoma. *PLoS One*, 5 (10) (2010), p. e13603.
46. Bonnet M, Guinebretiere JM, Kremmer E, Grunewald V, Benhamou E, et al. (1999) Detection of Epstein-Barr virus in invasive breast cancers. *J Natl Cancer Inst* 91: 1376–1381.
47. Brink AA, van Den Brule AJ, van Diest P, Meijer CJ (2000) Re: detection of Epstein-Barr virus in invasive breast cancers. *J Natl Cancer Inst* 92: 655–656; author reply 656.
48. Lin , Ching-Hwa Tsai, Jan-Show Chu , Jeou-Yuan Chen, Kenzo Takada, and Jin-YuhShew.(2007). Dysregulation of HER2/HER3 Signaling Axis in Epstein-Barr Virus-Infected Breast Carcinoma Cells. *J. Virol.* June 2007 vol. 81 no. 11 5705-5713
49. Smith TM, Lee MK, Szabo CI, Jerome N, McEuen M, Taylor M, Hood L and King M-C. (1996) . Complete genomic sequence and analysis of 117kb of humanDNA containing the gene *BRCA1*. *Genome Res* 6: 1029–1049.
50. Petrij Bosch A, Peelen T, van Vliet M, van Eijk R, Olmer R, Drusedau M, Hogervorst FB, Hageman S, Arts PJ, Ligtenberg MJ, MeijersHeijboer H, Klijn JG, Vasen HF, Cornelisse CJ, van't Veer LJ, Bakker E, van Ommen GJ and Devilee P (1997) *BRCA1* genomic deletions are major founder mutations in Dutch breast cancer patients. *Nat Genet* 17: 341–345.
51. Ansquer Y, Gautier C, Fourquet A, Asselain B and Stoppa-Lyonnet D (1998).Survival in early-onset *BRCA1* breast cancer patients. *Lancet* 352: 541.
52. Watson P, Marcus JN and Lynch HT (1998) Prognosis of *BRCA1* hereditary breast cancer. *Lancet* 351: 304–305
53. Honrado E, Osorio A, Palacios J, Benitez J. Pathology and gene expression of hereditary breast tumors associated with *BRCA1*, *BRCA2* and *CHEK2* gene mutations. *Oncogene*. 2006;25: 5837–5845. [PubMed]
54. Phillips KA. Current perspectives on *BRCA1*- and *BRCA2*-associated breast cancers. *Intern Med J*. 2001;31: 349–356.
55. Foulkes WD. *BRCA1* and *BRCA2*: chemosensitivity, treatment outcomes and prognosis. *Fam Cancer*. 2006;5: 135–142.
56. Plakhins G, Irmejs A, Gardovskis A, Subatniece S, Liepniece-Karele I, Purkalne G, et al. Underestimated survival predictions of the prognostic tools Adjuvant! Online and PREDICT in *BRCA1*-associated breast cancer patients. *Fam Cancer*. 2013.
57. Bayraktar S, Gutierrez-Barrera AM, Liu D, Tasbas T, Akar U, Litton JK, et al. Outcome of triple-negative breast cancer in patients with or without deleterious *BRCA* mutations. *Breast Cancer Res Treat*. 2011;130: 145–153.
58. Eccles D, Simmonds P, Goddard J, Coultas M, Hodgson S, et al. Familial breast cancer: an investigation into the outcome of treatment for early stage disease. *Fam Cancer*. 2001;1: 65–72.
59. Robson ME. Clinical considerations in the management of individuals at risk for hereditary breast and ovarian cancer. *Cancer Control*. 2002;9: 457–465.
60. Budroni M, Cesaraccio R, Coviello V, Sechi O, Pirino D, Cossu A, et al. Role of *BRCA2* mutation status on overall survival among breast cancer patients from Sardinia. *BMC Cancer*. 2009;9: 62 .
61. Chappuis PO, Kapusta L, Begin LR, Wong N, Brunet JS, Narod SA, et al. Germline *BRCA1/2* mutations and p27(Kip1) protein levels independently predict outcome after breast cancer. *J ClinOncol*. 2000;18: 4045–4052.
62. Dersimonian R, Laird N. Meta-analysis in clinical trials. *Control Clin Trials*. 1986;7: 177–188.
63. Narod SA, Foulkes WD. *BRCA1* and *BRCA2*: 1994 and beyond. *Nat Rev Cancer*. 2004;4: 665–676.
64. Bordeleau L, Panchal S, Goodwin P. Prognosis of *BRCA*-associated breast cancer: a summary of evidence. *Breast Cancer Res Treat*. 2010;119: 13–24.
65. Lee EH, Park SK, Park B, Kim SW, Lee MH, Ahn SH, et al. Effect of *BRCA1/2* mutation on short-term and long-term breast cancer survival: a systematic review and meta-analysis. *Breast Cancer Res Treat*. 2010;122: 11–25.
66. Broek AJ, Schmidt MK, van 't Veer LJ, Tollenaar RA, van Leeuwen FE.(2015). Worse Breast Cancer Prognosis of *BRCA1/BRCA2* Mutation Carriers: What's the Evidence? A Systematic Review with Meta-Analysis. *PLoS One*. 2015 Mar 27;10(3):e0120189.
