



## **Enzymes From Actinomycetes – Review**

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**Abstract :** Actinomycetes are group of microorganisms produce valuable secondary metabolites like antibiotics, vitamins, organic acids and enzymes. Antibiotics from actinomycetes of different habitats have been employed extensively in pharmaceutical field. The enzymes produced by actinomycetes and applied in different industries are amylases, proteases, lipases, cellulases, xylanases, chitinases, gelatinases and keratinases. This review summarizes the application of both intracellular and extracellular enzymes of actinomycetes in different industries such as textile, biorefineries, food, pulp and paper, agriculture, detergent and pharmaceuticals.

**Keywords :** Actinomycetes, Enzymes, Chitinases, Amylases.

### **Introduction**

The micro-organisms which live in soil cannot transport complex molecules inside their cytoplasm, so, they depend on extracellular enzymes for breakdown of these molecules into useful and essential nutrients. Extracellular enzymes from microorganisms are important bio-catalysts with their widespread applications in industries such as textile, bio-refineries, food, pulp and paper, agriculture, detergent and pharmaceuticals. The active secondary metabolites produced by microorganisms are reported to be around 23,000. There are 10,000 active secondary metabolites are produced by actinomycetes. Among actinomycetes, approximately 7,600 bioactive compounds are produced by *Streptomyces* species<sup>[1]</sup>. Actinomycetes are of enormous importance since they possess a capacity to produce and secrete a variety of extracellular hydrolytic enzymes<sup>[2-4]</sup>. Many actinomycetes have been isolated from various natural sources, as well as in plant tissues and rhizospheric soil. Biological functions of actinomycetes mainly depend on sources from which they are isolated. Physiological, biochemical and molecular characteristics and metabolic pathway of aquatic actinomycetes are different from terrestrial actinomycetes. Saline actinomycetes produced a variety of biologically active enzymes than the terrestrial actinomycetes. This review summarized the studies on the extracellular and intracellular enzymes production by actinomycetes from different sources.

### **Amylases**

Amylases are categorized into exoamylases and endoamylases, these hydrolyze the starch molecules to variety of products including dextrans and smaller polymers composed of glucose units<sup>[5]</sup>. Actinomycetes secrete amylases to the outside of the cells to carry out extracellular digestion. Amylase starch degrading amyolytic enzymes play their major role in biotechnological applications such as food industry, fermentation and textile to paper industries<sup>[6]</sup> and having approximately 25% of the demand in the world enzyme market<sup>[7, 8]</sup>. Amylases can be derived from plants, animals and microbes. The enzymes from microbial origin generally meet great demand in the industries. Occurrence of amylases in actinomycetes is a characteristic commonly occurred in *Streptomyces*<sup>[9]</sup> and the genus considered as an active source of amylases. *Streptomyces avermitilis*,

*Streptomyces* sp. SLBA-08; *Streptomyces* strain A3; *Streptomyces rochei* BTSS 1001 are used in production of amylase in starch, detergent, food and textile industries. It is effectively used in field of medicinal research<sup>[10-12]</sup>. Industrial processes of starch degradation have been improved with the help thermostable amylolytic enzymes. Extracellular amylase production by a newly isolated alkali-thermotolerant strain *Streptomyces gulbargensis* DAS 131 was studied for the highest amylase production<sup>[13]</sup>. A haloalkaliphilic marine *Saccharopolyspora* sp. strain A9 with an ability to produce surfactants, oxidant and detergent stable amylase was isolated from marine sediments<sup>[14]</sup>. The surfactant, detergent stable and calcium ion independent amylase from strains A3 was isolated which has widespread applications for detergent and pharmaceutical industry<sup>[1]</sup>.  $\alpha$ - Amylases have potential and wide application not only in industrial processes but has been applied widely in many fields such as clinical, medicinal and analytical chemistry. Amylaes have been utilized effectively in starch saccharification and in the textile, food, brewing and distilling industries<sup>[15,16,6]</sup>.

### Cellulases

Cellulases required for the hydrolysis of cellulose<sup>[17]</sup> and they are a collection of hydrolytic enzymes which hydrolyze the glucosidic bonds of cellulose and related cello-digosaccharide derivatives<sup>[18]</sup>. Actinomycetes are one of the known cellulase producers<sup>[19, 20]</sup>. *Streptomyces drozdowiczii*, *S. lividans*, *S. longispororuber*, *S. rugersensis*, *Streptomyces* sp. B-PNG23 are better examples for production of cellulase and used in industries such as pulp and paper, textiles, biorefineries, animal feedstocks, wine and brewing, baking<sup>[21-27]</sup>. Agro-industrial wastages are better utilized by actinomycetes in converting the wastages into cellulose by the enzyme cellulases. *Streptomyces viridobrunneus* SCPE-09 was selected as the active cellulolytic strain produces cellulose from agro- industrial residue<sup>[28]</sup>. Cellulolytic enzymes are employed in the color extractions of juices, in detergents causing color brightening and softening, in the biostoning of jeans, in the pretreatment of biomass that contains cellulose to improve nutritional quality of forage and in the pretreatment of industrial wastes<sup>[29-33]</sup>. Alkaline or alkalitolerant and cellulase producers are mainly found in the genera *Streptomyces* and *Thermoactinomyces*<sup>[34]</sup>. The cost of enzyme production can be reduced by using low value biological substrates (fruit processing waste)<sup>[35]</sup>.

### Xylanases

Xylan is the most dominating component of hemicelluloses, used in the pulp and paper industry<sup>[36]</sup>. Alkaliphilic and cellulase-free xylanases with an optimum temperature of 65°C from *Thermoactinomyces thalophilus* subgroup C was also reported recently<sup>[37]</sup>. Thermostable xylanase were isolated from a number of actinobacteria<sup>[38]</sup>. *Streptomyces spp* have been reported to produce xylanases which are active at temperatures between 50 and 80°C. Eighty eight actinomycetes were isolated from the soil samples, India for their production and characterization of xylanase<sup>[39]</sup>. *Actinomadura* sp. from compost in Thailand has been reported for the production of xylanase<sup>[40]</sup>. Thermophilic *Actinomadura* sp. from poultry compost has been reported the production and characterization of extracellular thermostable xylanase production<sup>[41]</sup>.

### Lipases

Lipases are produced from a variety of actinomycetes<sup>[42]</sup>. Lipases have broad applications in the detergent industries, foodstuff, oleochemical, diagnostic settings and also in pharmaceutical fields<sup>[43]</sup>. Lipases and esterases are a diverse group of enzymes that catalyze the hydrolysis of ester bonds in triacylglycerides to glycerol and fatty acids. Lipases have extensive range of enzymatic properties and substrate specificities. Lipases are used in processing of fat and oils, additives, detergents, cosmetics, paper manufacturing and pharmaceuticals.

### Proteases

Proteases, also known as peptidyl-peptide hydrolases, are important industrial enzymes and are extensively used in variety of industries including textiles, leather, detergents, meat tenderization, cheese making, dehairing, baking, organic synthesis, brewery and waste water treatment<sup>[44-45]</sup>. These enzymes also used in production of digestive aids and the recovery of silver from photographic film. Actinomycetes, particularly *Streptomyces* are known to secrete multiple proteases in culture medium<sup>[46]</sup>. Microbial alkaline proteases for manufacturing uses are produced mostly from *Streptomyces spp*. Several studies have been made on the proteolytic enzymes of mesophilic actinomycetes<sup>[47]</sup>. *Streptomyces thermonitrificans* showed maximum protease activity<sup>[48]</sup>. Recently, alkaline protease from *Nocardiopsis* sp. NCIM 5124<sup>[49]</sup> has been purified and

characterized. Alkaliphilic actinomycete from the soil and crude components such as molasses, wheat flour, and wheat bran were found to be effective for growth and protease production<sup>[50]</sup>. The high level of enzyme production using agro-industrial by-products is commercially significant due to cheap nature of these sources. *S. gulbargensis* DAS 131 was isolated from soil samples and that was proved to produce multiple proteases<sup>[51]</sup>. There are 46 strains of actinomycetes have been isolated from soil samples of Northern Himalayas and studied their culture characterization, protease production and cytotoxic effects on cancer cell line<sup>[52]</sup>.

### Keratinases

Keratinases are extra cellular enzymes used for the bio degradation of keratin<sup>[53]</sup>. Keratinases are produced only in the presence of keratin substrate. Some microbes have been reported to produce keratinases in the presence of keratin substrate. Keratinase producing microorganisms have ability to degrade chicken feathers, hairs, nails, wool etc<sup>[54-55]</sup>. Mostly protease positive actinomycetes are useful for studying the production of proteases. Actinomycetes, particularly streptomycetes are known to secrete multiple proteases in culture medium<sup>[46]</sup>. The promising applications of keratinolytic proteases include enzymatic dehairing of leather, detergent industry and development of biodegradable films<sup>[56]</sup>. There is a great demand for developing biotechnological alternatives for recycling of keratin wastes, converting unused chicken feather to useful value added products with help of actinomycetes keratinases<sup>[57]</sup>. Different studies on keratinase activity of *Streptomyces sp* have been reported<sup>[58-60]</sup>.

### L-asparaginase

Actinomycetes are the excellent resource for the production of L-asparaginase (L-asparagine amino hydrolase). A range of soil actinomycetes, *Streptomyces griseus*, *S. karnatakensis*, *S. albidoflavus* and *Nocardia sp.* have abilities to produce L-asparaginase enzyme<sup>[61, 62]</sup>. Microbial L-asparaginases have been used as a therapeutic agent in the cure of certain human cancers, mostly in acute lymphoblastic leukemia<sup>[63]</sup>.

### Chitinases

Chitin is an insoluble linear 1, 4-linked polymer of N-acetylglucosamine. It is found in the cell walls of fungi and exoskeleton of insects and the shells of crustaceans. Chitinases are produced by viruses, bacteria, actinobacteria, higher plants and animals and they play important physiological and ecological roles<sup>[64]</sup>. Chitinases hydrolyze the 1, 4 linkages in chitin, yielding predominantly N-Ndiacetylchitobiose, which is further degraded by N-acetylglucosaminidases to the N-acetylglucosamine monomer<sup>[65]</sup>. Amongst actinomycetes, the genus streptomycetes is the best studied for chitinases<sup>[66]</sup>. Chitinolytic activity of culture filtrates of *S. griseus* has been reported<sup>[67, 68]</sup>. *Streptomyces thermoviolaceus* OPC-520 was isolated to extract the thermophilic chitinases<sup>[69]</sup>. Chitinase was isolated from the culture filtrate of *Streptomyces sp.* M-20<sup>[70]</sup>. *Nocardiopsis prasina* showed chitinase activity<sup>[65]</sup>. Chitinase is the potential antifungal agent through its chitin degradation activity<sup>[71]</sup>. Endophytic *Streptomyces aureofaciens* CMUAc130 produced chitinase and showed antagonism against phytopathogenic fungi<sup>[72]</sup>. *Streptomyces griseolobus* JCM4480, *Streptomyces Clauifer* JCM5059, *Streptomyces anulatus* NBRC13369 and *S. griseus* that produced chitinase compounds, showing selective inhibition of the insect GlcNAcase<sup>[73]</sup>. *Streptomyces hygrosopicus* was isolated from Thailand and studied chitinase activity against phytopathogenic fungi<sup>[74]</sup>. *S. griseus* strain (MTCC) was studied for its chitinase enzyme activity against some soil borne plant pathogens<sup>[75]</sup>. Chitinase activity against *Sclerotinia sclerotiorum* was studied with 186 endophytic actinomycetes from nine kinds of plants<sup>[76]</sup>. *Streptomyces tendae* strain TKVL 333 was isolated from laterite soils of the Guntur region, India, for chitinase production<sup>[77]</sup>.

### Conclusions

Recent studies on importance and application of microbial enzymes in industries proved that the enzymes from microbial origin generally meet great demand in the industries. Actinomycetes are of enormous importance since they possess a capacity to produce and secrete a variety of extracellular hydrolytic enzymes that are safer to environment. Extracellular enzymes from actinomycetes are important biocatalysts with their widespread applications in industries. Since the actinomycetes play their major role in industrial enzymes production, these could occupy their priority in different industries for giving solutions to many challengeable problems in the diverse field like textile, biorefineries, food, pulp and paper, agriculture, detergent and pharmaceuticals.

## References

1. Berdy J. Bioactive microbial metabolites, J. Antibiot., 2005. 58(1): 1-26.
2. Saadoun I. Rawashdeh R. Dayeh T. Ababneh Q and Mahasneh A. Isolation, characterization and screening for fiber hydrolytic enzymes-producing streptomycetes of Jordanian forest soils, Biotechnology, 2007. 6 (1), 120-128.
3. Tan H. Deng Z and Cao L. Isolation and characterization of actinomycetes from healthy goat faeces, Letters in Appl Microbio, 2009. 49 (2), 248-253.
4. Sathya R. Ushadevi T. Industrially Important Enzymes Producing *Streptomyces* Species From Mangrove Sediments. Int J Pharm Pharm Sci, 2014. 6(10), 233-237.
5. Windish WW. Mhatre NS. Microbial amylases. In: Wayne WU, editor. Advances in applied microbiology, 7. New York: Academic Press; 1965. 273-304.
6. Pandey A. Nigam P. Soccol CR. Soccol VT. Singh D. Mohan R. Advances in microbial amylases. Biotechnol Appl Biochem. 2000; 31(2):135-152.
7. Rajagopalan G. Krishnan C. Alpha-amylase production from catabolite derepressed *Bacillus subtilis* KCC103 utilizing sugarcane bagasse hydrolysate. Biores Technol. 2008; 99:3044-3050.
8. Reddy NS. Nimmagadda A. Sambasiva Rao KRS. An overview of the microbial  $\alpha$ -amylase family. Afr. J. Biotechnol. 2003; 2:645-648.
9. Vigal T. Gil JF. Daza A. Garcia-Gonzalez MD. Martin JF. Cloning characterization and expression of an alpha amylase gene from *Streptomyces griseus* IMRU 3570. Mol Gen Genet; 1991. 225:278-288.
10. Hwang SY. Nakashima K. Okai N. Okazaki F. Miyake M. Harazono K. Ogino C. and Kondo A. Thermal stability and starch degradation profile of  $\alpha$ -amylase from *Streptomyces avermitilis*. Biosci Biotechnol Biochem, 2013. 7, 2449-2453.
11. Chakraborty S. Raut G. Khopade A. Mahadik K. Kokare C. Study on calcium ion independent amylase from haloalkaliphilic marine *Streptomyces* strain A3. Ind J Biotech 2012; 11:427-437.
12. Acharyabhatta A. Kandula SK. and Terli R. Taxonomy and polyphasic characterization of alkaline amylase producing marine actinomycete *Streptomyces rochei* BTSS 1001. Int Jour of Micro, 2013, p. 1-8.
13. Syed DG. Agasar D. Pandey A. Production and partial purification of  $\alpha$ -amylase from a novel isolate *Streptomyces gulbargensis*. J Ind Microbiol Biotech 2009a; 36(2):189-194.
14. Chakraborty S. Khopade A. Biao R. Jian W. Liu XY. Mahadik K. et al. Characterization and stability studies on surfactant, detergent and oxidant stable  $\alpha$ -amylase from marine haloalkaliphilic *Saccharopolyspora sp.*A9. J Mol Catal B: Enzymat 2011; 68(1):52-58.
15. Gupta R. Gigras P. Mohapatra H. Goswami VK. Chauhan B. Microbial  $\alpha$ -amylases: a biotechnological perspective. Process Biochem. 2003; 38:1599-1616.
16. Kandra L.  $\alpha$ -Amylases of medical and industrial importance. Journal of Molecular Structure (Theochem) 2003:666-667. 487-498.
17. Matsui I. Sakai Y. Matsui E. Kikuchi H. Kawarabayasi Y. Honda K. Novel substrate specificity of a membrane-bound  $\alpha$ -glycosidase from the hyperthermophilic archaeon *Pyrococcus horikoshii*. FEBS Lett; 2000. 467(2):195-200.
18. Ito S. Alkaline cellulases from alkaliphilic Bacillus: enzymatic properties, genetics, and application to detergents. Extremophiles. 1997. 1:61-66.
19. Jang HD and Chenks. Production and characterisation of thermostable cellulase from *Streptomyces* transformant T3-1. World J. Microbiol. Biotechnol; 2003.19:263-268.
20. Arunachalam R. Wesley EG. George J and Annadurai G. Novel approaches for Identification of *streptomyces nobortoensis* TBGH-V20 with cellulase production. Curr. Res, Bacteriol. 2010. 3(1): 15-26.
21. Yassien MAM. Jiman-Fatani AAM. and Asfour HZ. Production, purification and characterization of cellulase from *Streptomyces sp.* Afri Journal of Microbio, 2014, 4. 348-354.
22. Javmen, A. Grigiskis, S. Rudenkov M. and Mauricas, M. 2013. Purification and partial characterization of a novel  $\beta$ -1,3-endoglucanase from *Streptomyces rutgersensis*. Protein J, vol. 32, 411-417.
23. Cirigilano MNF. Rezende RC. Oliveira MPG. Pereira PHF. Nascimento RP. Bon EPS Macrae A. and Coelho RRR. *Streptomyces misionensis* PESB-25 produces a thermoacidophilic endoglucanase using sugarcane bagasse and corn steep liquor as the sole organic substrates. BioMed Research International, 2013.1-9.

24. Kuhad RC. Gupta R. and Singh A. Microbial cellulases and their industrial applications. Enzyme Research, 2011. 1-10.
25. Azzeddine B. Abdelaziz M. Estelle C. Mouloud K. Nawel B. Nabila B. Francis D. and Said. B. 2013. Optimization and partial characterization of endoglucanase produced by *Streptomyces* sp. B-PNG23. Arch Biol Sci, vol. 65, p.549-558.
26. Shweta A. Cellulases of bacterial origin and their applications: A review. Inter Journal of Sci and Res, 2012. 3, 1652-1655.
27. Bagewadi ZK. Vernekar AG. Patil AY. Limaye AA. and Jain VM. Biodegradation of industrially important textile dyes by actinomycetes isolated from activated sludge. Biotechnol Bioinf Bioeng, 2011. 1, 351-360.
28. Da Vinha FNM. Gravina-Oliveira MP. Franco MN. Macrae A. da Silva Bon EP. Nascimento RP, et al. Cellulase production by *Streptomyces viridobrunneus* SCPE- 09 using lignocellulosic biomass as inducer substrate. Appl Biochem Biotech. 2011. 164(3):256-67.
29. Buchert J. Pere J. Oijusuoma L. Rahkamo L. Viikari L. Cellulases-tools for modification of cellulosic materials. In: Niches in the world of textiles world conference of the textile institute, Manchester, England; 1997. 284-290.
30. Niehaus F. Bertoldo C. Kähler M. Antranikian G. Extremophiles as a source of novel enzymes for industrial application. Appl Microbiol Biotechnol 1999; 51(6):711-729.
31. Bhat M. Cellulases and related enzymes in biotechnology. Biotechnol Adv 2000; 18(5):355-383.
32. Nakamura H. Kubota H. Kono T. Isogai A. Onabe F. Modification of pulp properties by cellulase treatment and application of cellulase to wastepaper deinking and mechanical pulp refining. In: 68th pulp and paper research conference proceedings of the pulp and paper research conference; 2001. 18 - 19.
33. Van Wyk J. Mogale A. Seseng T. Bioconversion of wastepaper to sugars by cellulose from *Aspergillus niger*. *Trichoderma viride* and *Penicillium funiculosum*. J Solid Waste Technol Manage 2001; 27(2):82-86.
34. Techapun C. Pooaran N. Watanabe M. Sasaki K. Thermostable and alkaline-tolerant microbial cellulase free xylanases produced from agricultural wastes and the properties required for use in pulp bleaching bioprocess: a review. Process Biochem 2003; 38(1):1327-1340.
35. Kumar D. Yadav KK. Singh M. Garg N. Biochemical utilization of agro-industrial lignocelluloses rich waste for cellulase production. Res J Agri Sci, 2012. 44(2):184-191.
36. Chen CC. Adolphson R. Dean JFD. Eriksson KEL. Adams MWW. Westpheling J. Release
37. of lignin from kraft pulp by a hyperthermophilic xylanase from *Thermatoga maritima*. Enzyme Microbiol Technol 1997; 20(1):39-45.
38. Kohilu U. Nigam P. Singh D. Chaudhary K. Thermostable alkaliphilic and cellulase free xylanases production by *Thermoactinomyces thalophilus* subgroup C. Enzyme Microb Technol 2001; 28:606-610.
39. Bode W. Huber R. Natural protein proteinase inhibitors and their interaction with proteinases. Euro J Biochem 2005; 204(2):433-451.
40. Ninawe S. Lal R. Kuhad R. Isolation of three xylanase-producing strains of actinomycetes and their identification using molecular methods. Curr Microbiol, 2006; 53(3):178-182.
41. Sriyapai T. Somyoonsap P. Matsui K. Kawai F. Chansiri K. Cloning of a thermostable xylanase from *Actinomadura* sp. S14 and its expression in *Escherichia coli* and *Pichia pastoris*. J Biosci Bioeng, 2011; 111(5):528-536.
42. Taibi Z. Saoudi B. Boudelaa M. Trigui H. Belghith H. Gargouri A. et al. Purification and biochemical characterization of a highly thermostable xylanase from *Actinomadura* sp. strain Cpt20 isolated from poultry compost. Appl Biochem Biotechnol 2012; 166(3):663-679.
43. Kulkarni N and Gadre RV. Production and properties of an alkaline, thermophilic lipase from *Pseudomonas fluorescens* NS2W. J. Ind. Food. Microbiol; 2002. 28: 344-348.
44. Schmid RD and Verger R. Lipases: Interfacial enzymes with attractive applications. Angew. Chem. Int. 1998. 37: 1608-1633.
45. Kalisz HM. Microbial proteinases. Adv Biochem Eng Biotechnol 1988; 36:1-65.
46. Kumar CG. Takagi H. Microbial alkaline proteases: from a bioindustrial viewpoint. Biotechnol Adv, 1999; 17(7):561-94.

47. Sharmin S. Hossain MT and Anwar MN. Isolation and characterization of a protease producing bacteria *Bacillus amonvivorus* and optimization of some factors of culture conditions for protease production, Journal of Biological Sciences, 2005. 5(3), 358-362,
48. Boeckle B. Galunsky B. Mueller R. Characterization of a keratinolytic serine proteinase from *Streptomyces pactum* DSM 40530. Appl Environ Microbiol. 1995. 61(10):3705–10.
49. Mohamedin A. Isolation, identification and some cultural conditions of a proteaseproducing thermophilic *Streptomyces* strain grown on chicken feather as a substrate. Inter Biodeter Biodegrad 1999; 43(1):13–21.
50. Dixit V. Pant A. Comparative characterization of two serine endopeptidases from *Nocardiosis* sp. NCIM 5124. Biochim Biophys Acta 2000; 1523(2): 261–268.
51. Mehta V. Thumar J. Singh S. Production of alkaline protease from an alkaliphilic actinomycete. Bioresour Technol 2006; 97(14):1650–1654.
52. Dastager S. Dayanand A. Li WJ. Kim CJ. Lee JC. Park DJ, et al. Proteolytic activity from an alkali-thermotolerant *Streptomyces gulbargensis* sp. nov. Curr Microbiol 2008; 57(6):638–642.
53. Balachandran C. Duraipandiyar V. Ignacimuthu S. Purification and characterization of protease enzyme from actinomycetes and its cytotoxic effect on cancer cell line (A549). Asian Pac J Trop Biomed 2012; 2(1):392–400.
54. Shigeri Y. Matsui T. Watanabe K. Decomposition of intact chicken feathers by a thermophile in combination with an acidulocomposting garbage-Treatment process. Bioscience Biotechnology Biochemistry. 2009. 73 (11):2519- 2521.
55. Gradisar J. Friedrich I. Krizaj and Jerala R. Applied Environmental Microbiology. 2005. 71. 3420 - 3426.
56. Cai B. Lou and Zheng X. Zhejiang. J. University Science, 2008, B 9, 60.
57. Jeong JH. Lee OM. Jeon YD. Kim JD. Lee NR. Lee CY. Son HJ. Production of keratinolytic enzyme by a newly isolated feather-degrading *Stenotrophomonas maltophilia* that produces plant growth promoting activity. Process Biochemistry. 2010. 45: 1738-1745.
58. Williams CS. Richter JM. Mackenzie JC. Shih H. Isolation, identification and characterization of a feather degrading bacterium. Appl Envir Microbiol. 1990.56:1509-1515.
59. Al-Zarban SS. Al-Musallam AA. Abbas IH. Fasasi YA. Noteworthy salt-loving actinomycetes from Kuwait. Kuwait J Sci Eng, 2002;29: 99–109.
60. Chitte R. Nalawade V. Dey S. Keratinolytic activity from the broth of a featherdegrading thermophilic *Streptomyces thermoviolaceus* strain SD8. Lett Appl Microbiol. 1999. 28(2):131–6.
61. Janaki T. Keratinase activity (Feather) of *Streptomyces cacaoi subsp. cacaoi-M20*. Int Journal of Pharma Sci and Research, 2016d, 1(2); 25-27.
62. Narayana KJP et al. L-asparaginase production by *Streptomyces albidoflavus*. Indian J. Microbiol., 2007, 48(3): 331-336.
63. Mostafa SA and Salama MS. L-asparaginase producing *Streptomyces* from soil of Kuwait. Zentralbl Bakteriell Naturwiss., 1979, 134(4): 325 - 334.
64. Verma N. et al. L-asparaginase: a promising chemotherapeutic agent. Crit. Rev. Biotechnol., 2007. 27(1): 45-62.
65. Gooday GW. The ecology of chitin decomposition. Adv Microb Ecol, 1990;11: 387–430.
66. Tsujibo H. Kubota T. Yamamoto M. Miyamoto K. Inamori Y. Characterization of chitinase genes from an alkaliphilic actinomycete. *Nocardiosis prasina* OPC-131. Appl Environ Microbiol 2003; 69(2):894–900.
67. Miyashita K. Fujii T. Sawada Y. Molecular cloning and characterization of chitinase genes from *Streptomyces lividans*. J Gen Microbiol; 1991, 137(9):2065–2072.
68. Ralph Berger L. Reynold DM. The chitinase system of a strain of *Streptomyces griseus*. Biochim Biophys Acta, 1958; 29(3):522–534.
69. Beyer M. Diekmann H. The chitinase system of *Streptomyces* sp. ATCC 11238 and its significance for fungal cell wall degradation. Appl Microbiol Biotechnol 1985; 23(2):140–146.
70. Tsujibo H. Okamoto T. Hatano N. Miyamoto K. Watanabe T. Mitsutomi M. et al. Family 19 chitinases from *Streptomyces thermoviolaceus* OPC-520: molecular cloning and characterization. Biosci Biotechnol Biochem 2000; 64(11):2445–53.
71. Kim KJ. Yang YJ. Kim JG. Purification and characterization of chitinase from *Streptomyces* sp. M-20. J Biochem Mol Biol 2003; 36(2):185–9.

72. Janaki T. Nayak BK. Ganesan T. Antifungal activity of soil actinomycetes from the mangrove *Avicennia marina*. *Journal of Medicinal Plants Studies*, 2016b; 4(2): 05-08.
73. Taechowisan T. Peberdy JF. Lumyong S. Isolation of endophytic actinomycetes from selected plants and their antifungal activity. *World J Microbiol Biotech*, 2003;19(4):381–385.
74. Usuki H. Nitoda . Okuda T. Kanzaki H. Screening and partial characterization of inhibitors of insect. beta.-N-acetylglucosaminidase. *J Pesticide Sci*, 2006;31(1):41–6.
75. Prapagdee B. Kuekulvong C. Mongkolsuk S. Antifungal potential of extracellular metabolites produced by *Streptomyces hygroscopicus* against phytopathogenic fungi. *Int J Biol Sci* 2008; 4(5):330–337.
76. Anitha A. Rebeeth M. In vitro antifungal activity of *Streptomyces griseus* against phytopathogenic fungi of tomato field. *Acad J Plant Sci*, 2009; 2(2):119–123.
77. Xiao-ning G. Gulpiye W. Li-li H. Xuan T. Zhen-sheng K. Screening of plant endophytic actinomycetes producing chitinase and its antagonistic activity against *Sclerotinia sclerotiorum*. *J Zhejiang Univ (Agric Life Sci)* 2010; 36(6):615–622.
78. Kavitha A, Vijayalakshmi M. Partial purification and antifungal profile of chitinase produced by *Streptomyces tendae* TK-VL 333. *Ann Microbiol* 2011; 61(3):597–603.

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