

## Level of Cortisol and Catecholamines using Enzyme Linked Immunosorbent Assay (ELISA) and Fourier Transform Infra Red (FTIR) in Dairy Cattle

\*<sup>1</sup>Risa Ummami, <sup>2</sup>Claude Mona Airin, <sup>2</sup>Pudji Astuti

<sup>1</sup>Departement of Biotechnology and Veteriner, Vocational School, Universitas Gadjah Mada, Yogyakarta, Indonesia

<sup>2</sup>Departement of Physiology, Faculty of Veterinary Medicine, Universitas Gadjah Mada, Yogyakarta, Indonesia

**Abstract :** Animal stress is commonly determined by measuring cortisol levels in blood, saliva, feces or urine. Samples used in this study were blood serum from cows that have already known their cortisol levels. Blood samples were taken from the three different times of milking process: one minute (just before milking), 50 minutes after milking, and 100 minutes after milking. The samples were measured at 4000-400cm<sup>-1</sup> wavelength using MB3000 FTIR. Data absorption band at a wavelength of FTIR was descriptively analyzed using Horizon MB<sup>TM</sup> FTIR software. The results of FTIR analysis were hydroxyl (-OH) at 3294-3321 cm<sup>-1</sup>, ketone (CO) at 1636 cm<sup>-1</sup> and the amine (-NH<sub>2</sub>) at 1551 cm<sup>-1</sup>. A methyl group (-CH<sub>3</sub>) was absorbed at a low level, thus it cannot get the absorbance peak at 2880 to 2950 cm<sup>-1</sup> wavelength. The mean of catecholamines levels using ELISA on  $\pm$  1 minute before milking, 50 minutes after milking and 100 minutes after milking were  $0.0469 \pm 0.00546$  ng/mL,  $0.0467 \pm 0.00263$  ng/mL and  $0.0502 \pm 0.00308$  ng/mL respectively. Two way ANOVA test results showed that the mean of catecholamines were not significantly different ( $p > 0.05$ ) among three milking treatments. Measurement of cortisol and catecholamine levels using ELISA and FTIR showed the same result, and no significant differences were found in ELISA assay and no wavelengths difference were found in FTIR.

**Keyword:** milking stress, FTIR, ELISA, cortisol, catecholamines.

### Introduction

Stress is a non-specific condition of discomfort causing some adverse impacts on animals such as decreased immunity, reproductive failure, decreased carcass weight, and even death<sup>18</sup>. Environmental stressors are not limited to climatic factors, but also from nutrition, housing and every stimulus that demands a response from animals to adapt to new circumstances<sup>14</sup>.

Milk is an almost ideal food. It has high nutritive value. It supplies body building proteins, bone forming minerals and health giving vitamins and furnishes energy giving lactose and milk fat. Besides supplying certain essential fatty acid<sup>20</sup>. Stress during milking process is not only violating animal welfare but also negatively affects milk ejection, which increasing the amount of residual milk that could have negative effects on health<sup>23</sup>. Furthermore, the milking process is also increasing risk of udder injury<sup>10,3</sup>.

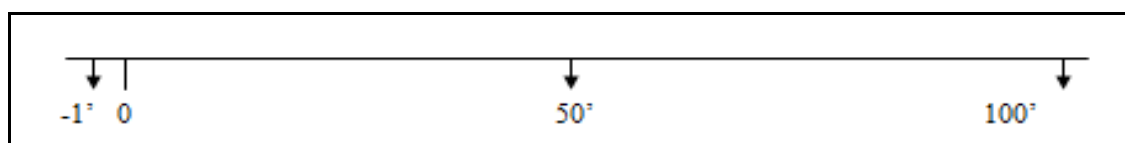
Detection of stress hormones using FTIR (Fourier Transform Infra Red) spectroscopy has previously conducted by other researcher in beef cattle<sup>1</sup>. Until now, FTIR has been applied to studies structural nickel

ferrite<sup>19</sup>, to research modification of magnetic nanoparticles with CMC<sup>11</sup>, to investigate the spectra in recycled and virgin resin<sup>28</sup>, to identify the presence of functional groups in the grown crystal<sup>35</sup>, to identify *Nicotiana tabacum* as biodiesel production<sup>9</sup>, to know nanostructure of phosphosilicate<sup>21</sup>, to analyse anti bacterial in bioactive compound<sup>32</sup>, to analyse spectra of antibiotic drug<sup>17</sup> also to estimate quantification of Clonidine hydrochloride in bulk<sup>38</sup>.

Researches on cortisol and catecholamine hormones analysis using FTIR and ELISA in dairy cows has not been conducted. There are a number of similar research that have been investigating the mechanism of cortisol and catecholamines with ELISA<sup>27</sup>, while FTIR measurements is widely used on human samples.

## Materials and Methods

Blood samples were collected three times: shortly before the portable milking machine was installed (approximately a minute before milking), 50 minutes after milking, and 100 minutes after milking ( Figure 1) . Each blood sample then collected into two venoject tubes without EDTA. To reduce the effects of stress due to blood collection, the blood was taken from coccigeal vein .



**Figure 1. Collected blood sample from minute 1 (before milking), 50 minutes after milking and 100 minutes after milking.**

### ELISA Analysis.

A commercial kit of TSZ ELISA Bovine catecholamine (Biotang, USA) was used to determine the catecholamine levels. The kit consist of a microplate (96 wells), a bottle of sample buffer (12 ml), a bottle of standards (100 ng), a bottle of biotinylated antibody (6 ml), a bottle of enzyme conjugate (12 ml), a bottle of wash solution (50 ml), a bottle of substrate solution (12 ml), and a bottle of stop solution (12 ml).

### FTIR Analysis.

The FTIR spectrum for serum specimens were recorded in 400-4000  $\text{cm}^{-1}$  frequency range using MB3000 FTIR spectrophotometer. Wavelengths produced were analyzed using Horizon MB<sup>TM</sup> FTIR software. The MB3000 FTIR spectrophotometer equipped with DTGS detector which cooled with air and thallium bromide crystal window (transparent IR).

### Calculation and statistical analysis.

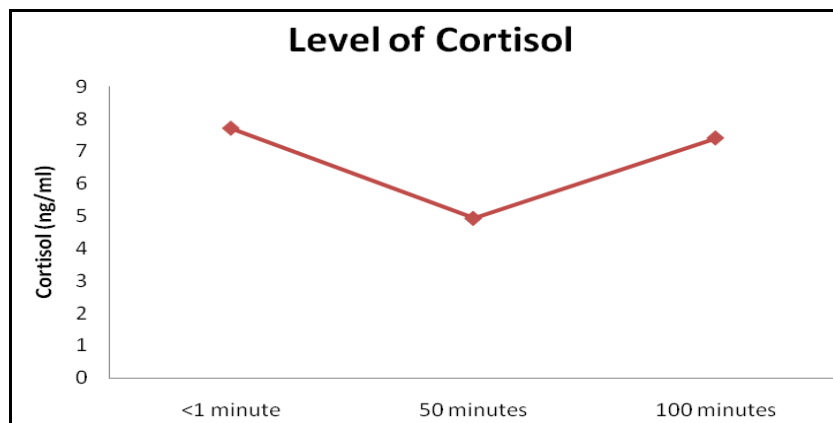
The data of absorption band at FTIR wavelength were analyzed descriptively with Horizon MB<sup>TM</sup> FTIR software. Quantitative confirmation test was conducted by measuring the levels of cortisol and catecholamines with ELISA. The differences among three groups were analyzed by two way Analysis of Variance (ANOVA), if there were any significant differences ( $p < 0.05$ ) Duncan test would be conducted<sup>5</sup>.

## Result and Discussion

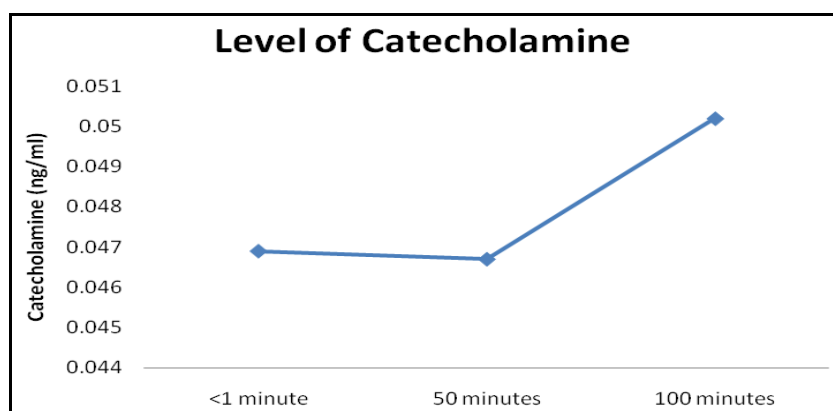
Catecholamines (epinephrine and norepinephrine) and its metabolites have been used to evaluate several types of animal behaviors, neuroendocrine disorders and stress conditions, from the point of physiology and pathology as a neurotransmitter or a hormone<sup>4</sup>. Serum samples used in this study have already measured its cortisol level in the previous study (Tabel 1), while the catecholamine levels detected using ELISA in this study were 0.04-0.058 ng/ml. The mean and standard deviation of catecholamines shortly before milking, 50 minutes after milking, and 100 minutes after milking were  $0.0469 \pm 0.00546$  ng/ml,  $0.0467 \pm 0.00263$  ng/ml, and  $0.0502 \pm 0.00308$  ng/ml respectively (Table 2).

**Tabel 1. The mean and standard deviation of cortisol levels in the blood serum Frisien types Holstein dairy cows 1 minute before milking, 50 minutes after milking and 100 minutes after milking (27).**

Treatment	Cortisol level (ng/ml) (27)
1 minute before milking	7,27 ± 4,64
50 minutes after milking	7,42 ± 5,17
100 minutes after milking	4,94 ± 1,76

**Figure 2. Cortisol level in the blood serum Frisien Holstein dairy cow**

The result of catecholamine measurements using ELISA were still considered as normal and were not much different compared to the results obtained by other researcher, i.e. 0.05 ng/ml<sup>16</sup>. Another study conducted by Lay et. al.<sup>15</sup> stated that catecholamine levels in dairy cows were not affected by lactation process. This founding was supported by Pesce et. al.<sup>25</sup>, who found that adrenal gland demodulation in mice that were lactating did not alter the normal levels of oxytocin, but significantly increases the feeding induction which further stimulates the release of oxytocin, given that the action of adrenal catecholamines are inhibited in central neurohypophysis.

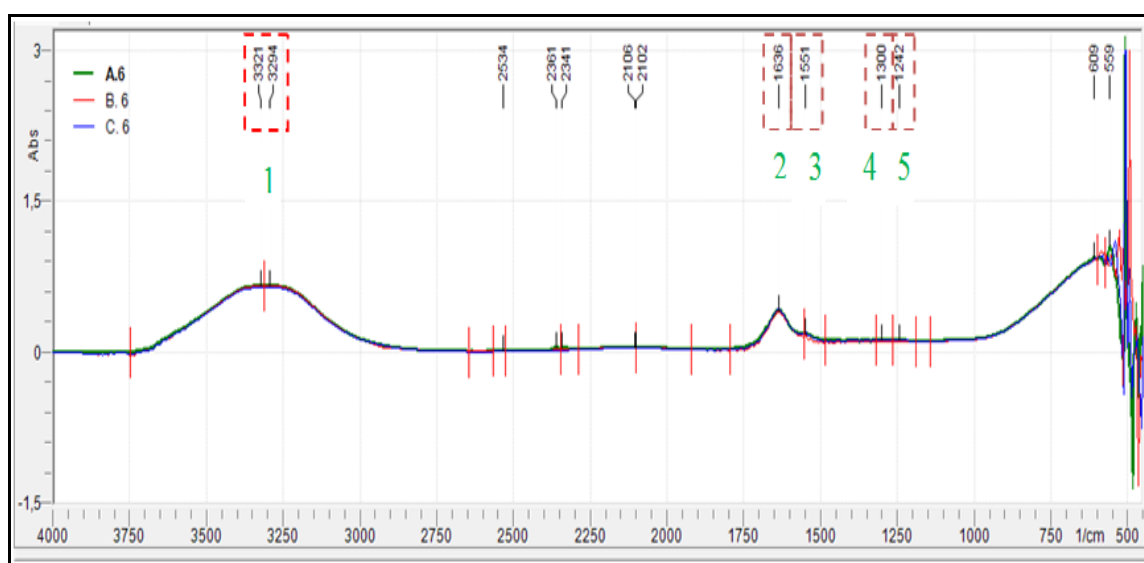
**Figure 3. Cathecolamine level in the blood serum Frisien Holstein dairy cow****Tabel 2. The mean and standard deviation of catecholamine levels in the blood serum Frisien Holstein dairy cows 1 minute before milking, 50 minutes after milking and 100 minutes after milking**

Treatment	Cathecolamine level ng/ml
1 minute before milking	0,0469 ± 0,00546
50 minutes after milking	0,0467 ± 0,00263
100 minutes after milking	0,0502 ± 0,00308

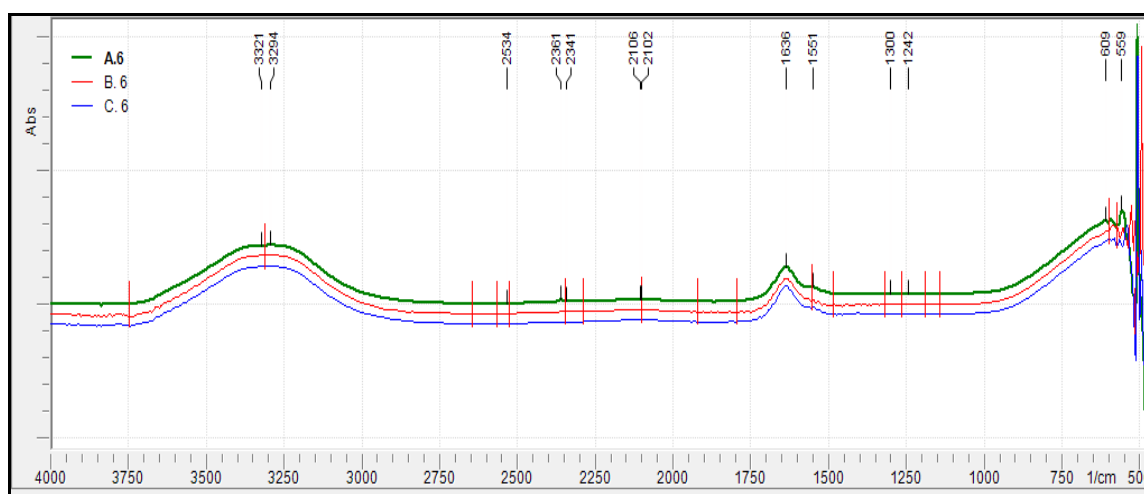
The result of two way ANOVA test showed that the difference of mean in three milking treatments: shortly before milking, 50 minutes, and 100 minutes after milking were not significant ( $p > 0.05$ ). Similar results were also obtained on cortisol level measurements that have been conducted previously by Titisari<sup>36</sup>, who found that there were no differences on cortisol levels shortly before milking, 50 minutes after milking, and 100 minutes after milking. These findings showed that catecholamines and cortisol had a same pattern, so that measuring catecholamine or cortisol levels could be a good indicator of stress response in animals.

### The measurement of stress levels using FTIR

Stress level was measured by determining the wavelength and evaluating the absorption level of functional groups composing cortisol and catecholamines on FTIR at 400-4000  $\text{cm}^{-1}$  wavelength (Figure 5 and figure 6). Serum spectrum contains several absorption bands in mid-IR region (600-4000  $\text{cm}^{-1}$ ) which is typical for biological samples<sup>24</sup>.



**Figure 5. Results and rate of absorption wavelength component of serum cortisol and catecholamines in the time before treatment, 50 minutes after treatment and 100 minutes after treatment.**



**Figure 6. Three-wavelength spectral separation, no visible difference from the third treatment**

The peak of absorption bands was at 3321  $\text{cm}^{-1}$  and 3294  $\text{cm}^{-1}$  wavelength (number 1), 1636  $\text{cm}^{-1}$  (number 2), 1551  $\text{cm}^{-1}$  (number 3), 1300  $\text{cm}^{-1}$  (number 4) and 1242  $\text{cm}^{-1}$  (number 5). Each of those wavelength was an absorption area for  $\nu\text{O-H}$  (3570-3120  $\text{cm}^{-1}$ ),  $\delta\text{C=O}$  (1720-1600  $\text{cm}^{-1}$ ),  $\delta\text{NH}_2$  (1600-1480  $\text{cm}^{-1}$ ),  $\nu\text{C-O}$  and  $\nu\text{C-O-C}$  (1300-900  $\text{cm}^{-1}$ ) and  $\nu\text{C-O}$  (1270-1200  $\text{cm}^{-1}$ ). The possibility of functional groups in number 1 and 2 were hydroxyl and ketone which are the constituent group of the cortisol hormone, while the functional group

at number 3 was amide group II, number 4 was the lactate group, and number 5 was a long chain fatty acid group.

The absorption rate in all three milking treatments showed that there was no peak for methyl ( $\text{CH}_3$ ) and methylene group ( $\text{CH}_2$ ). The absence of the peak of methyl signifies that  $\text{CH}_3$  and  $\text{CH}_2$  groups in the sample was absorbed at a low level absorbance. If infrared light was passed through organic compounds, then there were several frequencies absorbed and several other forwarded or transmitted without being absorbed. The absorption of light by molecules depends on the structure of the molecules. In the energy absorbing molecules, there were changes in vibration energy and rotational energy level. At room temperature, molecules of the organic compound are stable, each bond has a frequency that is characteristic for the stretching vibration and bending vibration where infrared light can be absorbed at these frequencies. The stretch energy of a bond is greater than its bending energy so that the stretch absorption of a bond appeared at a higher frequency in the infrared spectrum compared to the bending absorption from the same bond. Infrared spectrum graphics were created from the percentage of absorption (absorbance) against its characteristics frequency. The shape of light spectrum from organic compounds is closely related to transitions among electronic energy levels<sup>26</sup>.

The role of cortisol in helping the body to cope with stress is likely related to its metabolic effects. Cortisol increases blood glucose levels by utilizing protein and fat deposits. It is a logical assumption that the glucose, amino acids, and fatty acids deposits are always available to use anytime when needed, for example in a stress condition<sup>8</sup>.

The  $3600\text{-}3000\text{ cm}^{-1}$  region is the primary and secondary amine absorption band ( $\text{NH}$ ,  $\text{NHR}$ ) and also  $\text{OH}$ , while the peak at  $3371\text{ cm}^{-1}$  was placed for  $\text{VNH}$  or  $\text{VOH}$ <sup>13</sup>. Amino acids are utilized in other processes beside growth, such as if the animal is under the effect of stress. Injury, infection, and stress increase the need of amino acids for protein synthesis, selective catabolism, or synthesis of other specific molecules<sup>18;22</sup>.

In wavelengths analysis using FTIR,  $1300\text{-}900\text{ cm}^{-1}$ ,  $1630\text{-}1570\text{ cm}^{-1}$ ,  $1720\text{-}1600\text{ cm}^{-1}$  and  $2880\text{-}2850\text{ cm}^{-1}$  wavelengths are typically used to identify functional groups and each of its characteristics, from  $\text{CO}$  (lactate, glycerol, saccharides),  $\text{NH}_2$  (amine),  $\text{CH}$  (amide) and  $\text{CH}_3$  (methyl) bending vibration<sup>24</sup>. Tsunoda<sup>37</sup> who measured catecholamine levels in mice using HPLC (High Performance Liquid Chromatography) method stated that it can be used to determine the mechanism of catecholamines in regulating blood pressure.

According to Prabahari<sup>26</sup>, FTIR method is able to recognize an increase in the absorption of functional group in cortisol and serum, such as the methyl group ( $=\text{CH}_3$ ), methylene group ( $=\text{CH}_2$ ), hydroxyl group ( $-\text{OH}$ ), carbonyl group (ketone) ( $\text{C}=\text{O}$ ), primary amines ( $\text{NH}$ ), secondary amines ( $\text{NHR}$ ), carboxyl group ( $\text{COO}^-$ ), glucose and urea, which increased due to stress. In this study, not all of the functional groups increase. These findings were in accordance with the result of ELISA test which showed that the catecholamine levels were slightly below normal, and also with the result of cortisol test by Titisari<sup>36</sup> who stated that the cortisol levels in all three milking treatments were normal.

The physiological response to stress is more than just behavioral changes. However, there are no physiological parameter to measure the stress response that have been identified yet<sup>7</sup>. In humans and animals, some parameters that have been used to study the stress response were measuring the hormone levels which were released from brain and other organs, white blood cells fluctuations, changes in heart rate and blood vessel elasticity. A stress stimulus that lasts a few seconds up to one minute is able to increase heart rate, respiratory rate and causing indigestion or decreasing food intake<sup>25</sup>.

Animal response toward stress depends on genetic inheritances, lifetime experiences and physiological responses. This is related to the function of HPA (hypothalamic-pituitary-adrenal) axis, sympathetic nervous system, and the immune system<sup>2</sup>. Animals acquire stress stimulus from the environment through vision, hearing, smelling and physical contact. In some situation like the first lactating experience and being in unknown environment, animals take these experiences as negative stimuli which is able to create stress<sup>10</sup>. During these stressful conditions, there are some nervous and hormonal changes in the animals that might be causing some changes in their behavior, reproduction, productivity and immunity.

## References

1. Astuti P., C.M. Airin, S. Widiyanto, A. Hana , H. Maheswari, L. Sjahfirdi. Penentuan Kadar Stres Sapi Menggunakan Fourier Transform Infrared (FTIR) : Studi awal pembuatan detektor stres. 2013. Jurnal Fakultas Kedokteran Hewan Universitas Gadjah Mada : Yogyakarta.
2. Borell, V.E., H. Dobson, A. Prunier. Stress, Behavior and Reproductive Performance in Female Cattle and Pigs. *J Hormones and Behavior*. 2007. 52(1). Pp 130-8
3. Bruckmeier, R.M. Normal and Disturbed Milk Ejection in Dairy Cows. 2005. *J. Dom Anim End*, 29. Pp 268-273.
4. Connolly, C.C., K.E. Steiner, R.W. Stevenson, D.W. Neal, P.E. Williams, K.G.M.M. Alberti, A.D. Cherrington,. Regulation of glucose metabolism by norepinephrine in conscious dogs. 1991. *Am.J.Physiol*. 261. E764-E772.
5. Crowther, J.R.. *The ELISA Guidebook*. Humana Press. Totowa, New Jersey. 2001.
6. Dimitrova M., D. Ivanova, I. Karamancheva, A. Milev, I. Dobrev. Application of FTIR-spectroscopy for diagnosis of breast cancer tumors, *Journal of The University of Chemical Technology and Metalurgy*. 2009. 44(3), 297-300.
7. Friend, T. The feasibility of using vocalization scoring as an indicator of poor welfare during slaughter. 1980. *Appl.Anim.Behav.Sci*.
8. Guyton. *Fisiologi Manusia dan Mekanisme Penyakit*. EGC: Jakarta; 2000.
9. Hariram, V. and Gowtham R. A., Chemometric Analysis of *Nicotiana tabacum* FAME's using GC/MS, FTIR and NMR Spectroscopic Studies, *International Journal of ChemTech Research*, 2016, 9 (04), 171 – 176.
10. Hemsworth, P.H., Barnett, J.L., Coleman, G.J., Hansen, C. A study of the relationships between the attitudinal and behavioural profiles of stockpersons and the level of fear of humans and reproductive performance of commercial pigs. *Applied Animal Behaviour Science* . 1989. 23, 301-314.
11. Herrera, A., Vela, L., and Morales, G., Synthesis of Magnetic Nanoparticles Coated with Covalently Bonded Carboxymethyl Cellulose, *International Journal of ChemTech Research*, 2016, 9 (05), 602 – 607.
12. Hopster, H. Coping strategies in dairy cows. Ph.D. Thesis. Agricultural University of Wageningen, Wageningen. 152 pp.1998.
13. Khaustova S., Shkurnikova M., Tonevitsky E., Artyusenkoa V., Tonevitsky A. Noninvasive biochemical monitoring of physiological stress by Fourier Transform Infrared saliva spectroscopy. 2010. Vol.135, pp 3183-3192.
14. Kleiner, J. Ramesh, M. Huleihel, B. Cohen, K. Kantarovich, C. Levi, B. Polyak, R.S. Marks, J. Mordechai, Z. Cohen, S. Mordechai. A comparative study of gallstones from children and adults using FTIR spectroscopy and fluorescence microscopy, *BMC Gastroenterology*. 2002. 2(3), 1-14.
15. Lay D.C.T.H., Friend K.E., Grissom, C.L., Bowers, M.E. Mal. 1992. Effect of freeze or hot-iron branding of Angus calves on some physiological and behavioral indicator of stress. *Appl. Anim. Behav.Sci*. 1992. 33:137.
16. Lee, C. N. Environmental stress effects on bovine reproduction. *Vet. Clin. North America. Food Anim. Pract*. 1993. 9(2): 263-273.
17. Leela, J.S.P.P., Hemamalini, R., and Muthu, S., The Spectroscopic (FTIR, FT-Raman and UV), first order hyperpolarizability and HOMO-LUMO analysis of an Antibiotic drug, *International Journal of ChemTech Research*, 2015, 8 (6), 203 -215.
18. Luthfirda, S, Aziz NS, Maheswari H, Astuti P, Suyatna FD, Nasikin M. Estrus Period Determination of Female Rats by FTIR through Identification of Reproductive Hormones Metabolites in Urine Samples. *International Journal of Basic and Applied Science IJBAS-IJENS*. 2011. Vol.11 N0.03.
19. Mahalakshmi, S. and Manja, K.S., Spectroscopic and Structural Studies of Nickel Ferrite Doped With Rare Earth Ions, *International Journal of ChemTech Research*, 2015, 7 (3), 1460 – 1464.
20. Makadiya, J. and Pandey, A., Quality Assessment and Detection of Adulteration in Buffalo Milk Collected From Different Areas of Gandhinagar by Physico-Chemical Method, *International Journal of PharmTech Research*, 2015, 8 (4), 602 – 607.
21. Mohamed, M. E.O., Metawe, F., Amany M.E., Osman, B.A.A., Enhanced Structural And Spectroscopic Properties Of Phosphosilicate Nanostructures By Doping With Al<sub>2</sub>O<sub>3</sub> Ions And Calcinations Temperature, *International Journal of ChemTech Research*, 2016, 9 (05), 228 – 234.

22. Naik SV, Mahendra S, Sharma HD. Short term changes in plasma hormones, metabolites, milk yield and physiological responses in epinephrine administration cows.2014. J.Bio.Innov.3(2) pp:63-72.
23. Obled, C, Papet, I, Breuillé, D. Metabolic bases of amino acid requirements in acute diseases. 2002. Curr Opin Clin Nutr Metab Care 5: pp. 189-197.
24. Petibois C, Cazorla G, Cassaigne A, Deleris G., FT-IR Spectroscopy Utilization to Sportsmen Fatigability Evaluation and Control. Medicine and Science in Sports and Exercise. 2000. Pp 1804.
25. Pesce G, Guillaume V, Jezova D, Faudon M, Grino M, Oliver C. Epinephrine in rat hypophysial portal blood is derived mainly from the adrenal medulla. 1990. Neuroendocrinology 52, 322-327.
26. Prabahari CP. Perbandingan Proses Penyembelihan Sapi Ditinjau Dari Tingkat Stres Menggunakan Fourier Transform Infrared Spectroscopy (FTIR). Thesis. 2014.
27. Prodjodiharjo S. Pengelolaan Daging. Jakarta: Direktorat Jenderal Peternakan, Departemen Pertanian;2002.
28. Realpe, A., Acevedo, M.T., Ricardo, R., Mechanical Properties of Laminate of Residual Polyester Resin Reinforced with Recycled Newspaper, International Journal of ChemTech Research, 2016, 9 (02), 257 – 262.
29. Reeds PJ, Jahoor F. The amino acid requirements of disease. Clin Nutr 20 2001: pp. 15-22.
30. Rushen J, Munksgaard L, Marnet PG, De Pesille AM. Human Contact and the Effect of Acute Stress on Cows at Milking. J of App An Behav Sci, 2001 73(1).Pp 1-14.
31. Shaw R A dan Mantsch H. Infrared spectroscopy in clinical and diagnostic analysis. Institute for Biodiagnostic, National Research Council of Canada. Winnipeg 1997 Canada.
32. Shree Devi, M.S., Rajeswaran, S.P., and Kannan, M., GC-MS, FTIR Analysis and Anti Bacterial Study of Bioactive Coumpounds of Chundaivattral Chooranam – A Siddha Poly Herbal Formulation, International Journal of PharmTech Research, 2015, 8 (10), 204 – 209.
33. Stull CL. Stress and Dairy Calves. Davis: University of California 1997 Davis.
34. Sudjadi. Penentuan Struktur SenyawaOrganik. Ghalia Indonesia. Jakarta;1983.
35. Thirumalaiselvam, B., Kanagadurai, R., Jayaraman, D., and Natarajan, V., Growth, XRD, Optical Absorption Spectrum, FTIR Spectroscopy and Z – scan study of L – Serine, International Journal of ChemTech Research, 2015, 8 (8), 363 – 370.
36. Titisari N., Fauzi A., Astuti P., Physiological and Behavioral Responses of Dairy Cows which Milked with Portable parlor. International Journal of PharmTech Research, 2016, 9 (6), 172-177.
37. Tsunoda M. Development of an analytical method for cathecolamines with HPLC-Chemiluminesce detection and its application. Chromatography 2005 Vol.26.No.3.
38. Wadher, S.J., Kalyankar, T.M., Puranik, M.P., and Jayshri S., A Validated FTIR Method for the Quantification of Clonidine Hydrochloride in Bulk and Tablet Formulation, International Journal of ChemTech Research, 2015, 8 (11), 93-101.

\*\*\*\*\*