



Review article: Herbal sources as a remedy for rheumatoid arthritis

R. S. Mohammed

Pharmacognosy Dept., National Research Centre, 33 Elbohouth St, (Former El-Tahrir St.), Dokki – Giza-Egypt. P.O.12622, ID: 60014618.

Introduction

Herbal sources of medicinal plants are in great demand for curing various diseases. Medicinal knowledge gained over trial and error over thousands of years in many regions around the world. Knowing that plants have a large number of phyto-constituents with different pharmacological activities, guide us towards the safe use of these plants in future for treatments of many diseases.

RA is the most common systemic inflammatory disorder that affects 1% of the population worldwide, it is more common among women than men(3:1), it may be mild self-limiting or may rapidly progress into a multisystem inflammation with irreversible joint destruction¹.

Rheumatoid arthritis (**RA**) is an autoimmune disease that occurs when the body's own immune system mistakenly attacks the synovium which is the cell membrane lining inside the joint leading to joint pain, swelling, stiffness and loss of joint function. **RA** mostly affect elderly people but nowadays it appear in younger age 20-40. Fortunately there are many herbs in nature which act as a remedy for this condition and reduce chronic joint inflammation in **RA**². Major pharmaceutical companies are currently paying attention on the research on plant materials for their potential medicinal value³

RA is an inflammation of synovial joint due to immune mediated response. Synovial inflammation, cartilage destruction, and bone erosions characterize the different stages of the disease in which several pro-inflammatory pathways are involved¹.

B cells and T cells lymphocytes are the main two components of the immune system that play an important role in inflammation associated with **RA**⁴. Herbal medicinal products (HMPs) that interact with the mediators of inflammation are used in the treatment of **RA**

1.The immune system

It is a defense system of the body that provides protection against foreign invaders, which are disease causing microbes (bacteria, parasite and fungi). Immune system works in two ways: identifying and killing pathogens. Detection of invaders is difficult because pathogen can modify rapidly according to environment effects and producing adaptation which is hard for the immune system to detect and allow the pathogens to successfully infects their host⁵, so there are multiple mechanisms involved in recognition and killing of pathogens⁶.

The immune system is a complex set-up of cells, tissues and organs that work together to provide protection against the threat of microorganisms and infections. White blood cells (Leukocytes) are the main components of the immune system as it can eliminate the germs or any other substances responsible for occurrence of diseases. Leukocytes are also called lymphoid organs as they are accumulated in thymus, spleen and bone marrow⁵. Leukocytes move throughout the body between organs and nodes by means of lymphatic vessels and blood vessels. So the defense system works in synchronized method to protect the body against disease causing agents.

1.1. Organs of the Immune System ^{7, 8,9}

1.1.1. Bone Marrow:

The bone marrow are responsible for creating all types of cells of the immune systems. It produces : B cells, immature T cells, natural killer cells (NKC), granulocytes and, it also generate red blood cells and platelets.

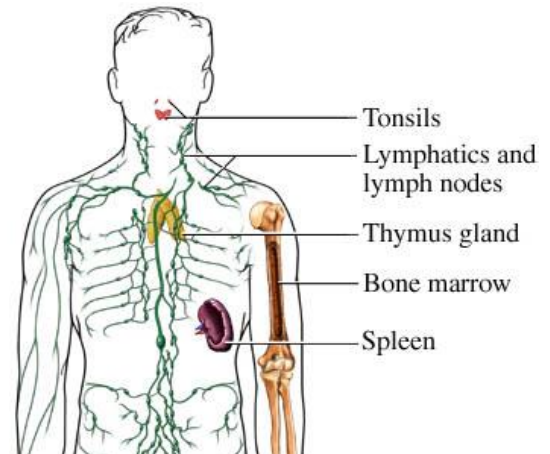
1.1.2. Thymus:

It produce mature T cells which are then migrate into the bloodstream.

1.1.3. Spleen:

It is an immunologic filter of the blood, capturing foreign materials (antigens) from the blood that passes through the spleen.

1.1.4. Lymph nodes: The lymph nodes has the same function of spleen it is responsible for filtering bodily fluid known as lymph. They are distributed throughout the body, it is composed mostly of T cells, B cells, dendritic cells and macrophages, antigens are filtered out of the lymph in the lymph node before returning the lymph to the circulation.



1.2. Disorders of immune system.

1.2.1 Acquired immune deficiency disorder: it is caused by virus which reduces efficiency of the immune system

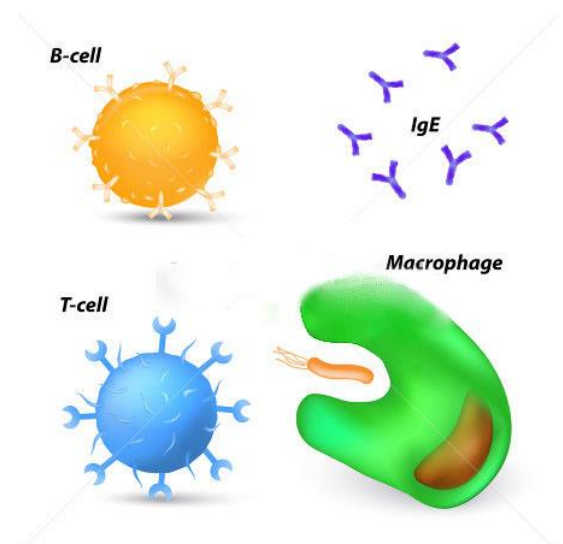
1.2.2. Autoimmune disorders: the body's own immune system cannot distinguish between foreign invader and its own tissues

1.2.3. An allergic disorder: the immune system over reacts is response to an antigen.

1.2.4. Cancers of the immune system

1.2.2. Autoimmune disease

The immune system in the body means protection against foreign substances. It has the ability to recognize tissues and cells that are its own (self) and differentiate them from those that are not (non-self). In autoimmune diseases, the body mistakenly attack its own tissues and consider it as foreign invader, the reason for this attack may be due to genetic, environmental factors or of unknown reasons



2. Pathophysiology of rheumatoid arthritis

RA starts in synovium which is the membrane lining sac which surround the joint. This sac containing synovial fluid which act as lubricant and cushioning the joints(Fig 1), the synovial fluid which coats the end of bones supplies cartilage with oxygen and nutrients ⁴

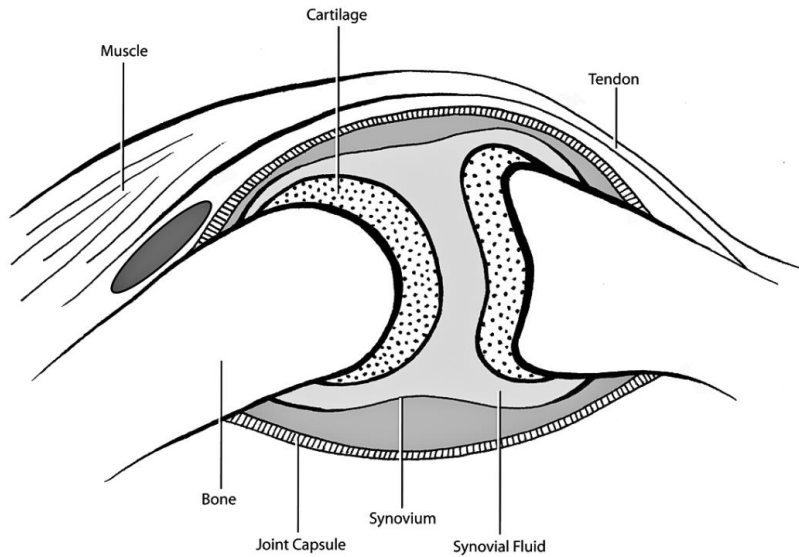


Fig 1

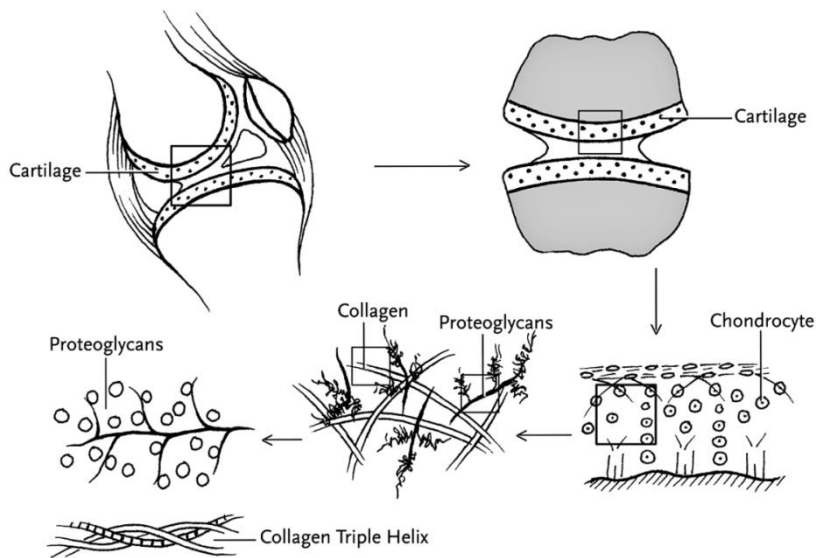


Fig 2

Cartilage is made of collagen which gives the joint support and flexibility. In RA harmful molecules produced by an abnormal immune system response cause inflammation of the synovium in turn collagen begin to destroy leading to narrowing the joint space and finally damaging bone. In a progressive rheumatoid arthritis, destruction of the cartilage accelerates(**Fig 3**).

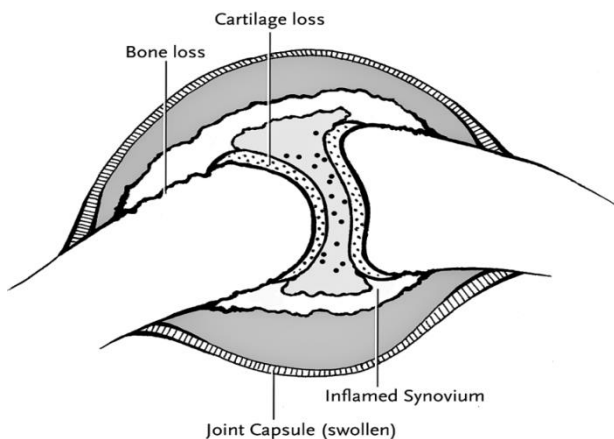
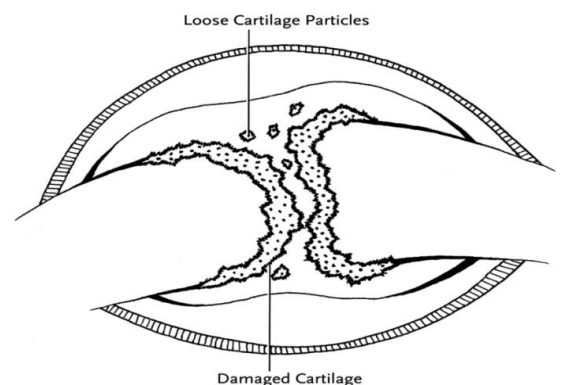


Fig 3



Which joints are affected

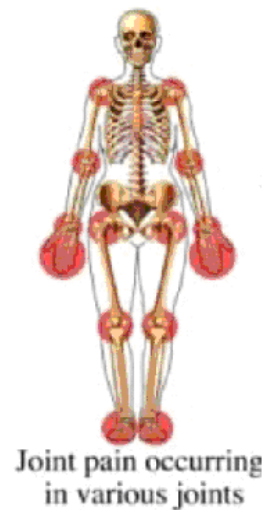
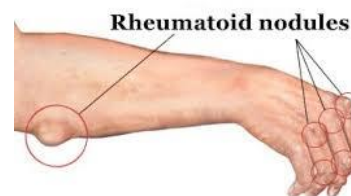
RA affects any joints of the body but the most affected joint are those of hands wrists and feet

Symptoms of RA ¹⁰

Small joints in the body are early affected and symptoms are often felt in:

fingers and toes, shoulders and knees can also be affected

1. Morning stiffness that last for at least 1 hr.
2. Pain, swelling, tenderness and stiffness of the joint after resting
3. Low-grade fever.
4. Appearance of painless small nodules under the skin which results from inflammation of small blood vessels



3.Diagnosing rheumatoid arthritis

It is hard to diagnose RA in its early stages, diagnosis is based on clinical assessment, lab. tests and X-rays.

Lab test:

1. Rheumatoid factor (RF) is an antibody in the blood it is linked with RA, 80% of people having RA give positive test.
2. Anti-CCP (anti-cyclic citrullinated peptide antibody) test may be also done to give better confirmation when combined with RF test ¹¹. Other way of diagnosis include X-rays and magnetic resonance imaging (MRI) scans ¹².

4.Etiology:

The etiology of RA is still unknown, a genetic susceptibility in combination with environmental factors may be the reasons for the onset of RA.

Factors which are responsible for the disease

4.1. Genetic factors:

RA has a genetic link, and the disease can run in families. Human leukocyte antigen (HLA) genes is linked with people have the chance of developing RA than people who do not have the HLA genes. Still, not everyone with the HLA genes develops RA People with specific.

4.2. Environmental factors:

Smoking may be linked to with the development of RF positive, RA has been associated with exposure to mineral oils, silica exposure, diet factors, and blood transfusion

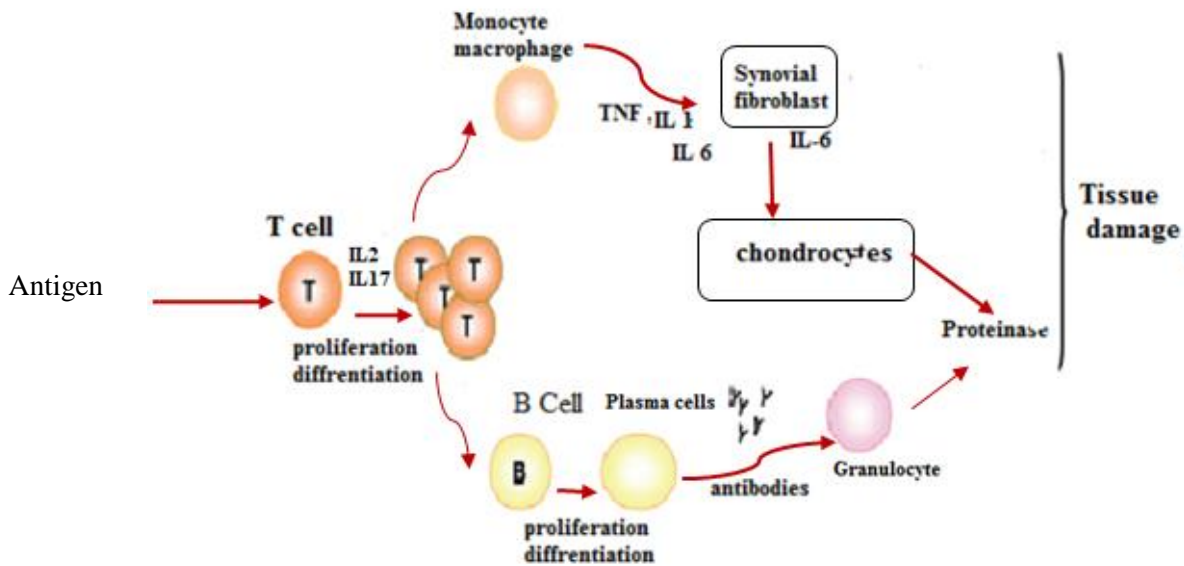
4.3. Effect of hormones :

Women were found to be more affected by RA than men , particularly at younger ages. In women, peak incidence is observed in the pri-menopausal, postpartum period and pregnancy.

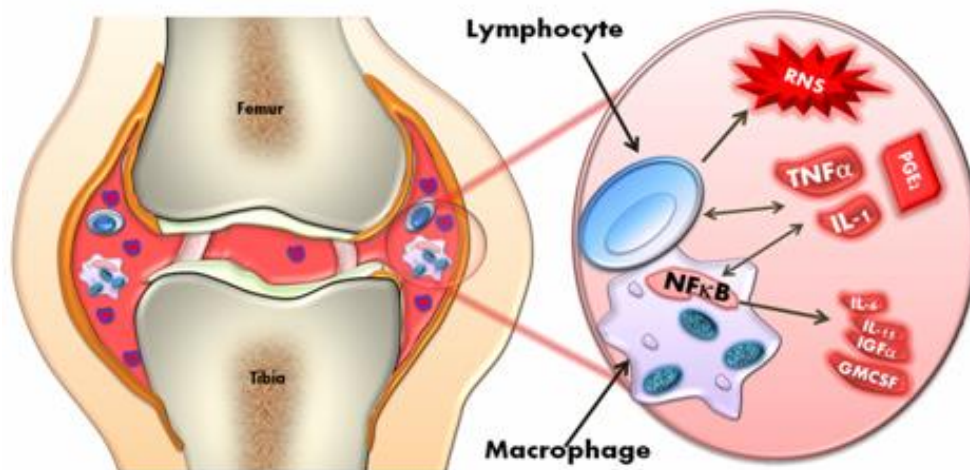
5.Mechanism of action of the immune system in RA

The T cell is activated when it recognize an antigen (non-self) it produces chemicals known as cytokines which cause B cells to multiply and release antibodies which circulate in the blood stream, recognizing the foreign particles(Fig 4,5) and causing inflammation to protect the body from invasion ¹³.

Proinflammatory cytokines: interleukin-1(IL-1)¹⁴ and tumor necrosis factor (TNF- α), are central mediators in RA, the regulation of these mediators and modulation of arachidonic acid metabolism by inhibiting enzymes like COX (cyclooxygenase) and LOX (lipoxygenase) are the potential target for chronic inflammation¹⁵ (Fig 6). There is a relationship between the concentration of IL-1 in and disease activity, it is noticed that patients with erosive RA have higher synovial and circulating levels of IL-1 than patients without erosions.



(Fig 4)



(Fig 5) In RA of joints the immune cells lymphocytes produces inflammatory cytokinins reactive oxygen and reactive nitrogen species (ROS/RNS)

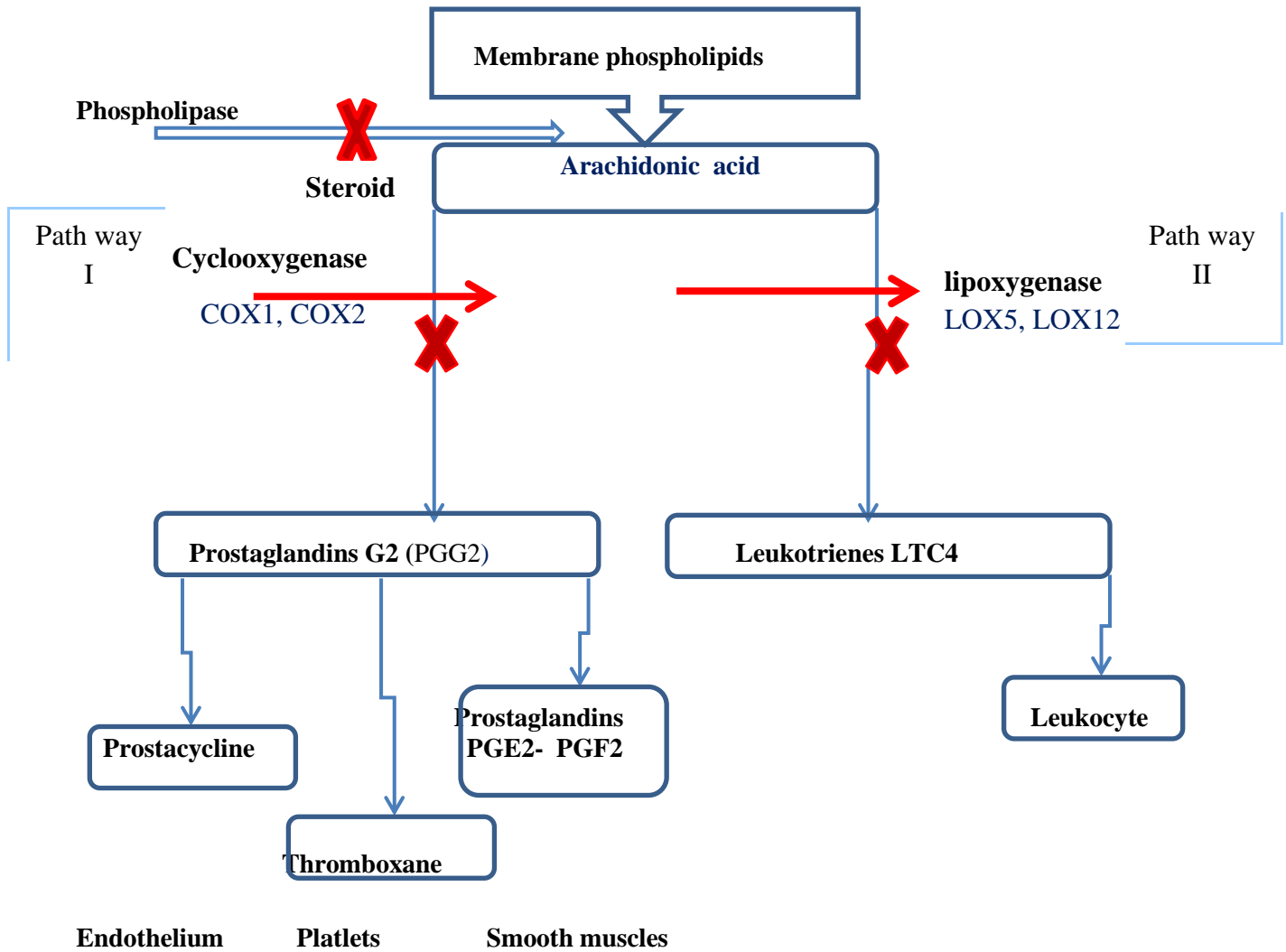


Fig 6 :Arachidonic acid path way

6. Methodology

Animal model

Induction of arthritis in animal models

6.1. Complete Freund's Adjuvant Induced arthritis (CFA)¹⁶

Freund's complete adjuvant induced arthritis in rat model is the best and most widely used experimental model for arthritis. This model is sensitive to anti-inflammatory and immune inhibiting medicines and best for the study of pathophysiological and pharmacological control of inflammation process.

Design of experiments

Animals are randomly divided into five groups each of six animals.

1. Group I served as controlled received normal saline.
2. Standard and test groups are further divide into two sections of treatment i.e.

3. Group II (Prophylactic P group, before induction of disease) and Group III (Therapeutic T group, after induction of disease). were standard groups received Indomethacin (10 mg/kg body weight).
4. Group IV(P) and V (T) were test groups that received the test drug¹⁷

On day zero animals are injected into the sub plantar region of the left hind paw with 0.1 ml of complete Freund's adjuvant (FA) Which consists of 6mg Mycobacterium butyric suspended in heavy paraffin oil by through grinding with mortar and pestle to give a concentration of 6 mg/ml. Drug treatment is started from the 14th day i.e. from the day of adjuvant injection and continued till 28th day. The paw oedema and joint thickness is measured on 7th, 14th, 21st and 28th day by using digital vernier callipers or plethys mometer. The mean changes in injected paw edema and joint thickness with respect to initial paw volume and joint thickness, are calculated on respective days and % inhibition of paw edema and joint thickness with respect to untreated group are calculated using following formula.

Inhibition in paw edema / joint thickness = $100 \times (1 - V_t/VC)$

VC = Mean paw edema volume/ joint thickness in control group

VT = Mean paw edema volume/ joint thickness in the drug treated group

Radiological analysis: radiographs are taken by using X-ray apparatus. X-rays are taken at the joint of hind paw of the animal for evaluating the bone damage before sacrificing the animal.

Histopathological analysis: the ankle joint of rats are removed and separated from the surrounding tissues. To examine the histopathological changes during the experimental period in all the groups under the light microscope, the joints are fixed in 10% formalin and decalcified, sectioned and finally stained with eosin and hematoxyline¹⁸

Other method were used:

6.2. Carrageenan induced paw edema in rats^{19, 20}

6.3. Formaldehyde Induced Arthritis^{19, 21}.

6.4. Collagen Type II Induced Arthritis (CIA) In Rats^{16, 22}

6.5. Pristane induced arthritis (PIA)²³

6.6. Oil-induced arthritis (OIA)²⁴

6.7. Streptococcal cell wall-induced arthritis²⁵

7. Treatment of RA

The main idea of treatment is focused on decreasing the disease activity along with minimizing joint destruction which will lead to improvement of the physical activity and quality of life.

7.1. NSAIDs: Paracetamol, Aspirin, Indomethacin

7.2. Corticosteroids: Prednisone, Prednisolone, Methyl prednisolone

7.3. Disease Modifying Anti-rheumatic Drugs (DMARDs): Methotrexate, cyclosporine, Leflunomide, Hydroxychloroquine, chloroquine, sulfasalazine, gold salts.

DMARDs: These are collection of various heterogeneous agents collected together according to their use, they reduce joint swelling, pain, decrease acute phase markers, limit progressive of joint damage, and improve function

7.4. Biological therapy

TNF-inhibitor, T- cell blockers, B- cell depletion molecules, IL-1 receptor antagonist were used. TNF α inhibitors : these medication lessen the signs and symptoms of RA, reduce progression of structural damage, and improves physical function , the clinical response can be observed within 2 weeks of treatment BUT:

1. There is a risk of getting microbial infection with the use of TNF inhibitors medication (tuberculosis and sepsis fungal infections)
2. A risk of development of cancer has been observed with the use of TNF- α . (incidence is very low), these compounds are usually administered in combination with other treatments ²⁶

Most of drugs which are used as anti –arthritic and anti- inflammatory did not suppress T-cell and B-cell mediated response ²⁷.

7.5. Herbal sources in RA

All the above mentioned therapies, which are used in the treatment of arthritis such as : NSAIDs, corticosteroids, DMARDs and biological therapy are helpful in decreasing the joint stiffness, pain.

But: the main drawbacks while they reduce the symptoms of the disease the progression of the disease continues, they also have a common side effects which includes: gastrointestinal ulcers, osteoporosis, serious infections like sepsis, tuberculosis, development of various lymphomas²⁸. So there is a hope to shift towards the use of herbal therapy in the treatment of RA

7.5.1. Ginger

Ginger is obtained from the rhizomes (underground stems) of *Zingiber officinale* (Zingiberaceae), it is well known medicinal herb and spices²⁹

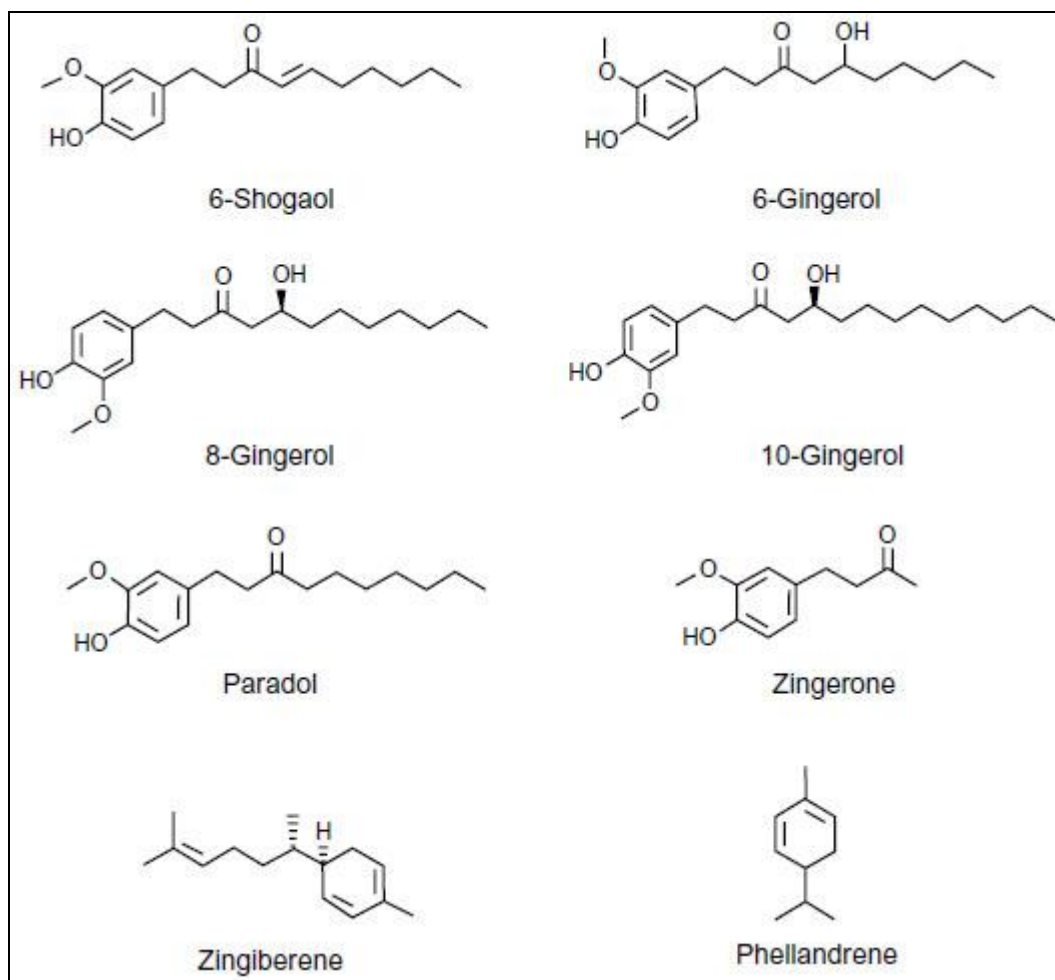


Active constituents

Ginger contains a large number of phytochemical constituents which were revealed to have valuable effect in RA ³⁰.

Phenolic compounds: Shogaols, paradols, and gingerols ³¹, Sesquiterpenes: Bisabolene, zingiberene, zingiberol, sesquiphellandrene, and curcurnene ³²,

Vitamins: VitaminA, vitamin C, Thiamine, riboflavin, niacin, pyridoxine, and vitaminE ³³ and other compounds. ³⁴



Mechanism of action on RA

Ginger showed beneficial effects of reducing the pain associated with RA which is due to inhibition of prostaglandin and leukotriene biosynthesis (block path way I, II), its mechanism of action as anti-inflammatory through blocking the activities of both COX-1 and COX-2, it suppress leukotriene biosynthesis through inhibition of 5-lipoxygenase³⁵.

Ginger was screened for anti-inflammatory activity using ibuprofen as standard, both showed similar anti-inflammatory activities indicating that ginger as a potential anti-inflammatory agent³⁶. The crude extract containing essential oils and more polar compounds exhibited better activities in preventing joint inflammation and bone destruction compared to the essential oils only. It was concluded that gingerol and non-gingerol compounds of ginger had considerable anti-arthritis activity³⁰.

7.5.1.1. Ginger therapy

Ginger was used traditionally by the Chinese in a variety of external application including muscular tension, disorder in metabolism and infection in the chest³⁷.

Transdermal delivery

Topical application of gingerols and/or shogaols to the skin is also tested and proved its efficiency. External application of dry ginger extracts in the form plasters showed promising anti-inflammatory activity indicating that the active constituents of ginger could penetrate through the skin³⁸.

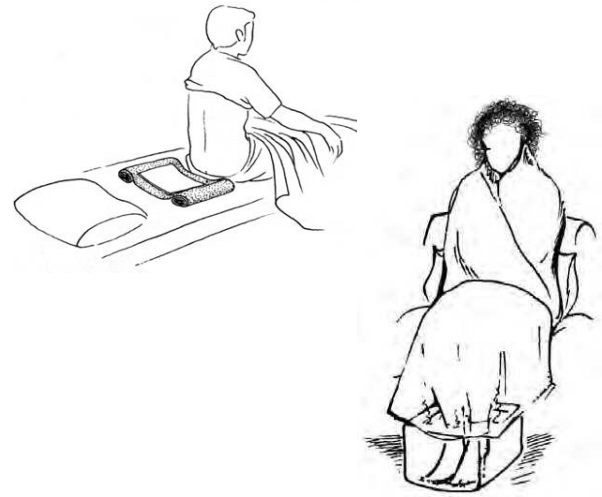
Ginger compress

Ginger compresses to the kidney region warm and reactivate the metabolism of the body. These application have an efficacy in some cases of arthritis, chest and psychiatric conditions³⁹.

Ginger compresses provide the body with heat and relaxation⁴⁰.

Ginger footbath

Ginger footbath is another method of decreasing muscle tension and relaxation of musculoskeletal fatigue



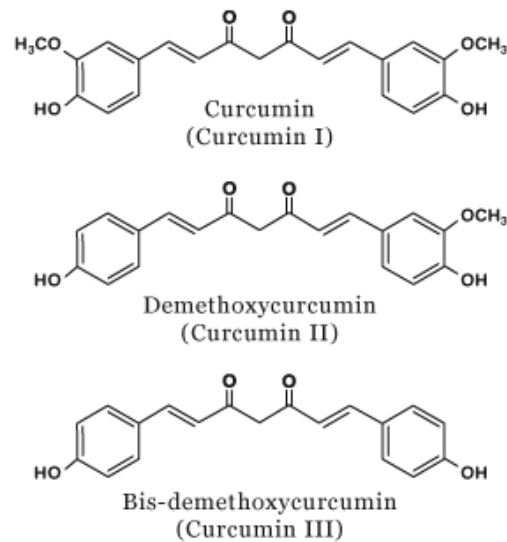
7.5.2.Tumaric

It is the rhizome of *Curcuma longa* L

(Zingiberaceae). It is used in Indian traditional medicine in curing inflammation⁴¹

Active constituents

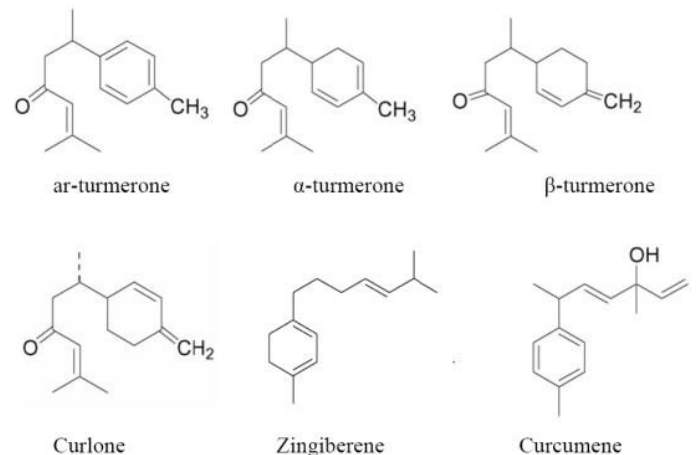
The major active constituents was known to be curcuminoids which comprises into three compounds (curcumin, demethoxy-curcumin, and bisdemethoxy curcumin), it also contain volatile oils constituted about 3-7%, its major compouds were zingiberone tumerone and atlantone ⁴².



Curcumin is the major curcuminoid in *Curcuma longa* it constitute 2-6%, it has a strong free radical scavenging activity so it can help in preventing disease result from free radical damage

Uses

- Curcuma has been used for centuries as spices , it is usually combined with several other spices for vegetable and meat preparations, it adds a characterstic fragrance and yellow color to foods.
- Curcuma was used traditionally in Asian and Indian medicine in various pharmacological activities : biliary disorders, intestinal disorders, hepatic disorder, diabetic wounds, anorexia, cough, rheumatic pain cancer, and Alzheimer's disease.



- The antioxidant activity of curcumin is important criteria in chronic inflammation⁴³.

Intraperitoneal injection(IP) of curcuminoids extract before arthritis induction inhibited acute (75%) and chronic (68%) inflammation of joint⁴⁴

Mechanisms of action

Curcumin is an alternative TNF- α blocker, it regulates the inflammatory response by inhibiting the activity of COX-2, lipoxygenase, it also decrease the production of cytokines (TNF- α) and Interlukin IL(1, 2, 6, 8, and 12)⁴⁵.

- It also inhibit the metabolism of arachidoic acid in mouse experimental model of skin inflammation through regulation of COX and LOX pathways(I &II)⁴⁶.
- 20-80 mg/kg bodyweight of Curcumin was successfully inhibited edema in experimental animals, it is nearly effective as phenylbutazone or cortisone at the same dose,40 mg/kg body weight of Curcumin showed anti-arthritic activity in formaldehyde induced arthritis model in rats .

7.5.3. Boswellia

Boswellia serrata : Part used is the oleo gum resin obtained by bark injury or natural crack

Active constituents

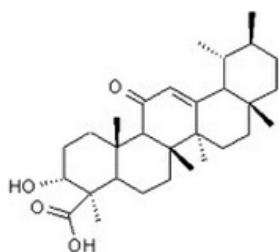
It include, oils, terpenoids and gum.

Essential oil is obtained in yield of 16% from oleo-gum-resin by steam distillation.: α -thujene , α -pinene , limonene , p-cymene , cadinene , geraniol and elemol constitute the major compounds in the essential oil ⁴⁷ boswellic acids was found to be the major terpenoid in the terpenoid part of the oleogum resin in *Boswellia serrata*.

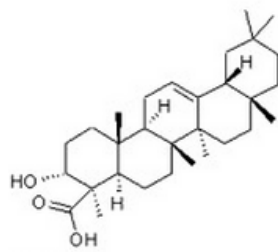


Boswellic acids are series of pentacyclic triterpenes which were used as anti-inflammatory through inhibition of biosynthesis of leukotriene ⁴⁸

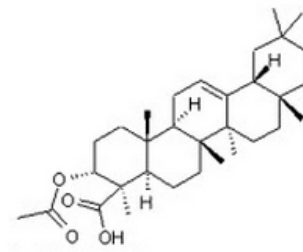
Clinical studies using *Boswellia* have yielded good results in both osteoarthritis and rheumatoid arthritis ^{49, 50}.



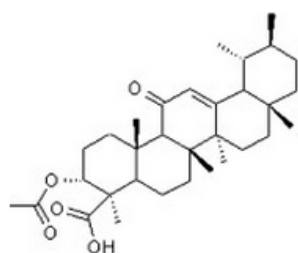
11-keto- β -Boswellic Acid



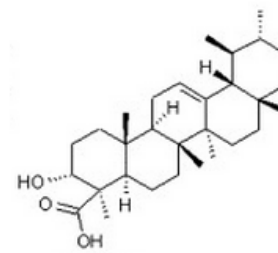
α -Boswellic Acid



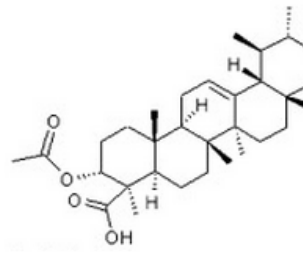
Acetyl- α -Boswellic Acid



3-O-Acetyl-11-keto- β -Boswellic Acid



β -Boswellic Acid



Acetyl- β -Boswellic Acid

Mechanism of action

Boswellic acids inhibit the leukotriene synthesis through its action on 5-LOX enzyme but it has no effect on 12-LOX and the COX activities^{51, 52}. 11-keto-boswellic acid and acetyl-11-keto-boswellic were found to be the most potent triterpenoids in the series of pentacyclic triterpene, They inhibit HLE (human leukocyte elastase) which is associated with rheumatoid arthritis and respiratory illnesses all of which are linked by inflammation.

- These two triterpenoid acids are potent elastase inhibitor compared with ursolic acid, which did not affect 5-LOX^{53, 54, 55, 56}.

7.5.4. Rosemary

Rosmarinus officinalis (Lamiaceae)

Part used whole plant and essential oils

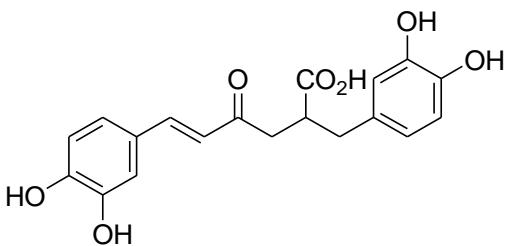


Active constituents

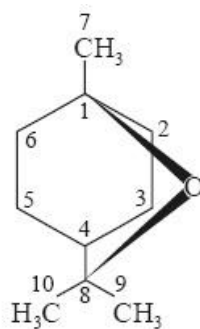
Carnosic acid, methylated carnosic acid, carnosol and rosmanol were the major compounds.

Water-soluble extract from rosemary leaves was rich in rosmarinic acid and flavonoids, oxygenated monoterpenes were also found : 1,8 cineole, borneol and terpineol.

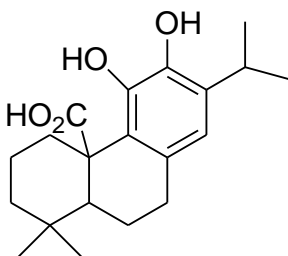
The main constituents in the essential oil of *Rosemary* dried leaves was 1,8 cineole (monoterpene oxide), monoterpene and sesquiterpene hydrocarbons (32.2%) á pinene, â-pinene, â -caryophyllene, camphene, limonene, myrcene p-cymene monoterpene ketonec amphor (12.8%) and monoterpene alcohols, borneol .All these compounds amounted to 93.4% of essential oil constituents⁵⁷.



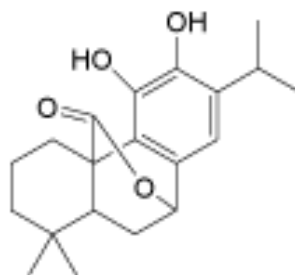
Rosmarinic acid



1,8 cineole



Carnosic acid



Carnosol

Uses

Rosemary essential oil is well known in aromatherapy due to its various reported health benefits, from stimulating hair growth to clearing the respiratory tract

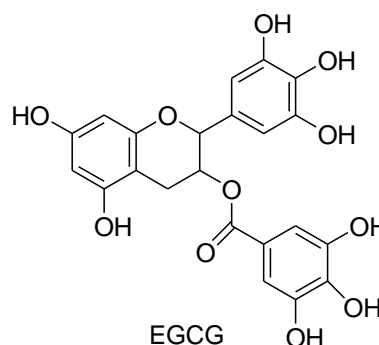
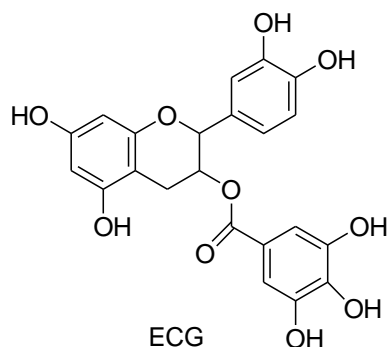
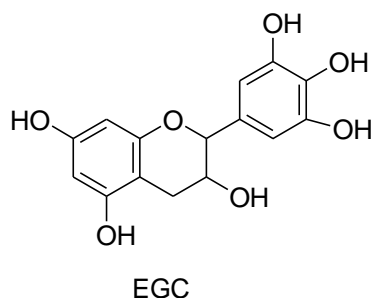
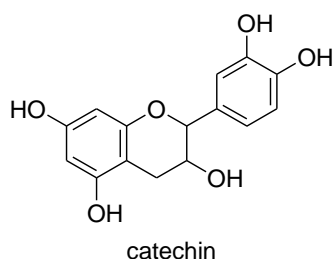
- The essential oil of *Rosemary* exert strong antioxidant activity in *in vitro* free-radical scavenging test, this activity is related to their phenolic compounds present in the essential oil, including rosmarinic acid, chlorogenic acid, caffeic acid, rosmanol, carnosolic acid, and carnosol.
- The antioxidant activity are useful for preventing tissue damage from the produced free radicals during inflammation, this explain the usefulness use of the antioxidant in treating musculo-skeletal problems, joints affected by rheumatoid arthritis.
- The high level of free radicals due to the immune system attacking the collagen and other proteins in the joint and as a result, the antioxidant constituents in *Rosemary* oil may scavenge free radicals, reducing tissue damage⁵⁸.
- Treatment with *rosemary* decrease the flow of white blood cells and mediators of inflammation⁵⁹.
- *Rosemary* essential oil is a pure herbal spray was used extensively for rheumatoid arthritis, joint and muscle pain without any side effects.
- It can be used in treating joint inflammations of rheumatic characteristic⁶⁰.Rosmary oil inhibit bone resorption in ovary ectomized rats⁶¹



7.5.5. *Camellia sinensis* Linn

The part used : leaves

The active constituents are polyphenols catechins, which is the major constituents : epicatechin (EC), epicatechin gallate (ECG), epigallocatechin (EGC), epigallocatechin gallate (EGCG), catechin (C), and gallocatechin (GC) and flavonols⁶².



Uses

- ✓ It has potent antioxidant activity due to high polyphenol contents, it reduces the risk of getting CVD and cancer⁶³.
- ✓ Black tea contain theophylline which is used as a therapy for respiratory tract diseases⁶⁴.

Mechanism of action

- ✓ The methanol extract of the leaves reduces inflammation and edema produced by inflammatory mediators. It improves the movement ability experimentally in induced arthritis in rats.
- Green tea reduced inflammation in the collagen-induced arthritic rats model, it reduces the inflammatory cytokines TNF- α & γ -interferon and COX-2^{63, 65}.
- EGCG found in green tea has a powerful scavenging activity to reactive oxygen species (ROS) which has a pathogenic role in rheumatoid arthritis and excessive quantities of oxygen free radicals have been identified in synovial fluid of 90% of patients with RA^{66, 67}.
- EGCG was found to have anti-arthritis activity which was first discovered from the finding that consumption of EGCG-containing green tea decrease collagen-induced arthritis in mice, inhibit the inflammatory mediators COX-2, Interferon gamma (IFN γ), and tumor necrosis factor alpha (TNF α) in arthritic joints
- *In vitro* and *in vivo* study supported the use of EGCG as anti-inflammatory and anti-arthritic effects indicating that EGCG or EGCG containing green tea can regulate the expression of cytokines, chemokines and other inflammatory mediator⁶⁸
- EGCG has been reported to have bone-preserving and synovial fibroblast regulatory effect .

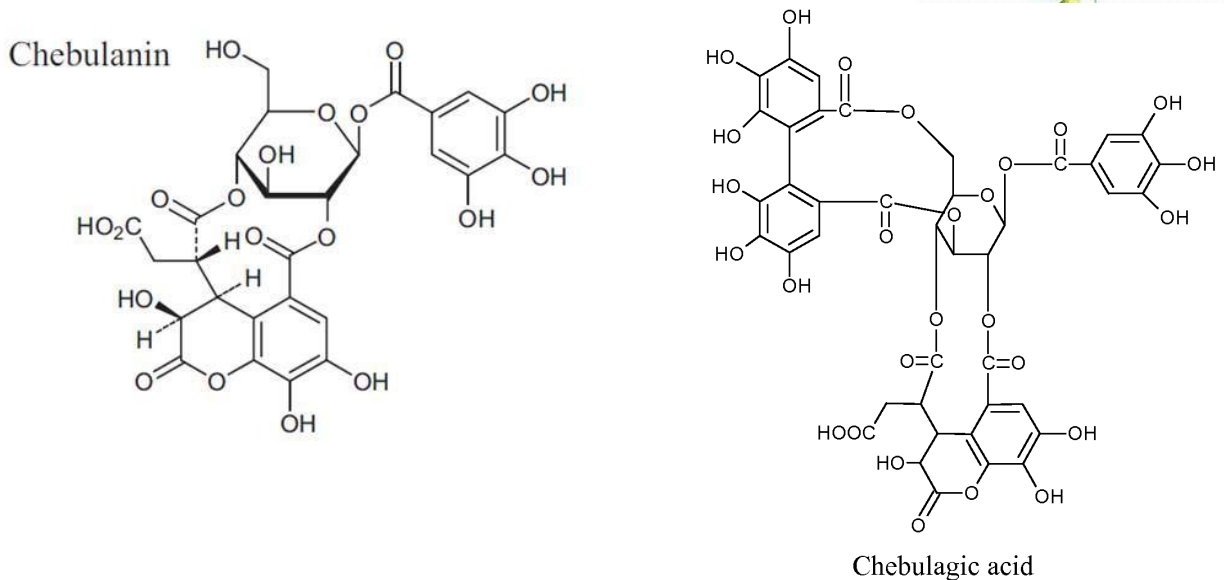
Water extract of black tea lessens in the serum levels of PGE2, TNF α , IL- 1 β and IL- 6 as shown in anti-arthritic model of Freund's adjuvant induced arthritic in rats.⁶⁹

7.5.6. *Terminalia chebula* Retz (Combretaceae)

Part used is the herb, fruit

Active constituents

T. chebula contain hydrolysable tannins 32%, flavonoids, sterols, amino acids, fructose, resin, fixed oils. hydrolysable tannins: Chebulic, chebulinic, chebulagic, gallic, were the main components which are responsible for anti - arthritic effect⁷⁰



Uses

- ✓ It has antifungal activity, used in leucorrhoea, It has antioxidant activity which is useful in protecting the neurons from the effect of free radicals
- ✓ Aqueous ethanol extract of *Terminalia chebula*, showed anti-arthritic in CFA induced arthritis activity and formaldehyde model.

Mechanism of action

- ✓ The plant perform its action as anti-arthritic activity through its modulatory effect on pro- inflammatory cytokine released in the synovium⁷¹

The hydroalcoholic extract of *T. chebula* produced a significant inhibition of joint swelling in both formaldehyde induced and CFA-induced arthritis.

- ✓ *T. chebula* could be used as a modulatory agent in treatment of rheumatoid arthritis⁷².

That acetone extract of *T. chebula* fruits showed anti-arthritic activity on CFA induced arthritis model through reduction of the inflammatory components⁷³ higher dose of 320mg/kg of *Terminalia chebula* showed promising activity as antiarthritic and anti-inflammatory activities in contrast to the traditional NSAIDs which causes ulceration of the stomach and GIT bleeding⁷⁴.

7.5.7. Aloe vera

Aloe barbadensis Mill (Alliaceae) part used: leaves

Active constituents : anthraquinones, anthraquinone C-glycosides and anthrones⁷⁵ which constitute about 30% from the constituents of the latex⁷⁶-, polysaccharides which is a linear polymer of glucose and mannose , amino acids, lipids, sterols and vitamins and enzymes⁷⁷

Uses

Aloe vera is used in **topical preparation** for skin ailments

It has many medicinal values, anthraquinone and anthranilic acid are quite efficacious against arthritis and articular Rheumatism⁷⁷

Mechanism of action

Aloe vera inhibits the COX pathway so decrease prostaglandin E2 production from arachidonic acid. *Aloe vera* extract (5.0% leaf homogenate) decreased inflammation by 48% in a rat adjuvant-induced arthritic inflammatory model^{78, 79}.

Science has not stopped yet, and the need for discovery of many herbal sources having the activity to be used in many disease continue.

Nature provides us with different herbs which possess the activity to be used in treating rheumatoid arthritis (Table 1)

Table 1: List of plants used in rheumatoid arthritis

No	Name of the plant	Family	Part used	References
1	<i>Aristolochia bracteata</i>	Aristolochiaceae	Whole plant	80
2	<i>Ammania bacifera</i>	Lytraceae	Whole plant	81
3	<i>Calotropis gigantea</i> (L.) R. Br.	Asclepiadaceae	Latex extract Root extract	82 83
4	<i>Capparis spinosa</i>	Capparidaceae	Fruit	84
5	<i>Cassia uniflora</i>	Caesalpiniaceous	Stem	85
6	<i>Centella asiatica</i>	Mackinlayaceae	Leaves	86



7	<i>Cleome rutidosperma</i>	Capparidaceae	Aerial parts	87
8	<i>Commiphora incisa</i>	Burseraceae	Resin Mansumbinoic acid	88
9	<i>Commiphora mukul</i>	Burseraceae	Gum-guggul	89
10	<i>Cocculus hirsutus</i>	Menispermaceae	Roots	90
11	<i>Cyperus rotundus</i>	Cyperaceae	Essential oil	91
12	<i>Cyperus esculentus</i> L.	Cyperaceae	Essential oil	91
13	<i>Dalbergia volubilis</i>	Fabaceae	Stem bark	92
14	<i>Daucus carota</i> L.	Apiaceae	Ethanol extract	93
15	<i>Delonix elata</i>	Fabaceae	Bark	94
16	<i>Elaeocarpus sphaericus</i>	Elaeocarpaceae	Fruit	95
17	<i>Euphorbia atiquorum</i>	Euphorbiaceae	Whole plant	96
18	<i>Ficus bengalensis</i>	Moraceae	Stem bark	97
19	<i>Glycirrhiza glabra</i>	Fabaceae	Rhizomes	98
20	<i>Glycosmis pentaphylla</i>	Rutaceae	Stem bark	99
21	<i>Hemidesmus indicus</i> (L)	Asclepiadaceae	Ethanol extract of root	100 101
22	<i>Justicia gendarussa</i> Burn f.	Acanthaceae	leaves	102
23	<i>Lawsonia Innermis</i>	Lythraceae	Leaves	14
24	<i>Leucas aspera</i> (Willd) Spreng.	Lamiaceae	Aerial parts	103
25	<i>Machalis macrantha</i>	Lauraceae	Bark	104
26	<i>Merremia tridentata</i> (L.) Hall	Convolvulaceae	Aerial parts , root	105
27	<i>Ncyntanthes arbortristis</i> Linn	<i>Oleaceae</i>	Leaves	106, 107
28	<i>Paederia foetida</i> L	Rubiaceae	Leaves	108, 109
29	<i>Phyllanthus amarus</i>	Euphorbiaceae	Herbs	110
30	<i>Pistiostratios</i> Araceae	Araceae	Leaves	111
31	<i>Pongammia pinnata</i>	Fabaceae	Leaves	112
32	<i>Premna serratifolia</i> L	Verbenaceae	Wood	113
33	<i>Punica grantum</i>	Punicaceae	Seeds	114
34	<i>Randiadam etorum</i>	Rubiaceae	Fruit	115
35	<i>Ricinus communis</i>	Euphorbiaceae	Leaves	116
36	<i>Saraca asoka</i> (Roxb.) wilde	Leguminosae	Flower , leaves	117
37	<i>Strychnos potatorum</i> Linn	Loganiaceae	Seeds	118
38	<i>Saussaurea lappa</i>	Asteraceae	Roots	119
39	<i>Sidarhom bifolia</i>	Malvaceae	Aerial parts	120
40	<i>Tinospora cardifolia</i>	Menispermaceae	Leaves	121
41	<i>Urtica pilulifera</i>	Urticaceae	Leaves	122
42	<i>Urgenia indica</i>	Liliaceae	Bulb	123
43	<i>Vernonia anthelmintica</i>	Asteraceae	Seeds	124
44	<i>Wedeli acalendulaceae</i>	Asteraceae	Leaves	125
45	<i>Withania somnifera</i> (L) Dunal		Root	126, 127

7.5.8.Plant constituents and RA

- **Gallic:** 128, 129 **Ferulic:** 130, 131 **Apigenins:** 132, 133 , 134 , 135, **Kaempferol:** 128, 136
- **Quercetin :**137, 138, 139 **Resveratrol :**140,141, 142_143 **Genistein:** 144, 145
Arctigenin: 146, 147

8.Some important relation in RA

- ✓ **8.1.Vitamin D** is an essential supplement for the immune system, it is produced normally by the body when the skin is exposed to sunlight. Deficiency of vitamin D may be one of the leading cause of autoimmune diseases especially RA 148 Vitamin D and its derivatives have been shown to suppress T-cell proliferation and inhibit the expression of pro-inflammatory cytokines involved in RA pathogenesis
- ✓ **8.2.Omega-3 polyunsaturated** fatty acids, found mainly in cold water fish e.g., salmon and the oil produced, can be useful for inflammatory arthritis like rheumatoid arthritis



BUT.

- ✓ Omega-3 fatty contraindicated with some medication of high blood pressure. Also patient taking aspirin must take care of omega-3 fatty as it can causes bleeding



8.3.Calcium

Calcium is very important for healthy bones and it is needed for the prevention of osteoporosis (brittle bones). Low-fat, milk, cheese, yogurt and the types of fish that are eaten with the bones (canned salmon or sardines) were the best sources of calcium

8.4.Protein

It is important for repairing cells of muscles and healthy immune system, select lean protein sources(chicken), seafood, beans, peas, nuts and seeds.

8.5.Folic acid

Folic acid is needed to prevent medication related side effects (liver abnormalities and gastrointestinal intolerances) especially for people taking methotrexate.

8.6.Glucosamine sulphate and chondroitin

Glucosamine sulphate and chondroitin supplements were used as supplement, they may help in improving the health of damaged cartilage, these compound were found to decrease inflammation

9.What to avoid in RA

9.1.Weight and Arthritis

Excess weight puts an extra pressure on your load-bearing joints: feet, ankles, knees, hips and back. When you walk the pressure in your knee joints increase, losing weight will help you and will make a difference than any food or supplement.

There are two measuring parameters of weight which is helpful in health problems average age taken was 18-65 but pregnant and lactating women were not included

- ✓ BMI : body mass index
- ✓ WC : waist circumference adults age 18 to 65 years with the exception of pregnant and lactating women.

9.2. Trans or saturated fats

Trans or saturated fats, increase the risk of getting CVD. These two harmful fats were found in processed and fried foods, they also come from animal sources of food. It is preferable to use Polyunsaturated and monounsaturated as the main source of fat it can be found in naturally in olive and canola oil, avocados and nuts.

9.3. Gout and diet

Gout is a type of inflammatory arthritis caused by the deposition of uric acid crystal in the joints, causing inflammation and severe pain. Uric acid is produced from purines based food, increase in the concentration of uric acid may be due to the kidney problems(excreting too little uric acid) or the body consume too many dietary purines. Decrease the amount of intake of food high in purines such as meat and seafood also organ meats (liver, kidney and sweetbreads). Vegetables, legumes, cereal, dairy products, soybean products and eggs are low in purine include.

Finally

Before the discovery of synthetic drugs man was completely dependent on the medicinal plants for the treatment of many disease. The medicinal value of plants has been recognized by every person of this society. There are many synthetic drugs that are being used as standard treatment for rheumatoid arthritis but they have adverse effect that can compromise the therapeutic treatment so these adverse effects increase the chances for the use of herbal plants for the treatment rheumatoid arthritis.

Rheumatoid arthritis is an inflammatory disease affecting the joint and can lead to improper movement or loss of function. RA is an autoimmune disease that can occur at any age but women are more likely to develop disease than men(3:1), it is more common in persons over the age of 30 years. RA is a systemic disease this means that it can affect the whole body, including internal organs : lungs, heart and blood. RA is a disease that affect vital part of the body which is the musculo-skeletal system and any neglect or delay in treatment may lead to dangerous health problems.

References

1. Lee, D.M and Weinblatt, M.E: Rheumatoid arthritis. *Lancet*, 358: (2001)903–911.
2. Bang, J.S.; O.h, D.H.; Choi, H.M.; Sur, B.J.; Lim, S.J.; Kim, J.Y.; Yang, H.I.; Yoo, M.C.; Hahm, D.H. and Kim, K.S.: Anti-inflammatory and anti-arthritic effects of piperine in human interleukin1- α stimulated fibroblast like synoviocytes and in rat arthritis models. *Arthritis Res. Ther.* (2009), 11: 1-9.
3. Hegen, M.; Keith, J.C and Collins, M. :Utility of Animal models for identification of potential therapeutics for rheumatoid arthritis. *Ann Rheum. Dis.* (2008), 67: 1505-1515.
4. Reddy, J.S.V; Deval Rao, G. and Rajya, L.G. : A review on anti-arthritic activity of some medicinal plants: *Journal of Global Trends in Pharmaceutical Sciences JGTPS.* (2014), 5(4) 2061-2073.
5. Arend, W. : The pathophysiology and treatment of rheumatoid arthritis. *Arthritis Rheum* (1997), 40(4): 595-7.
6. Beck, G. and Habicht, G. S.: The Evolution of the Immune System (1996),60-71.
7. More, P. and Pai, K.: Immunomodulatory effects of *Tinospora cordifolia* (Guduchi) on macrophage activation. *Biology and Medicine.* (2011),3:134-140.
8. Lu Yin; Fan Jie; Zhao Yunpeng; Chen Shaoyuan; Zheng Xiaodong; Yin Yuanming and Fu Chengxin: Immunomodulatory activity of aqueous extract of *Actinidia macrosperma*, *Asia Pac J Clin. Nutr* (2007),16: 261-265.
9. Agrawal, S. S.; Khadase C. S. and Gokul, S.: Studies on Immunomodulatory Activity of *Capparis zeylanica* Leaf Extracts. *International Journal of pharmaceutical Sciences and Nanotechnology* (2010), 3:887-892.
10. Scott, D.L; Wolfe, F and Huizinga, T.W: Rheumatoid arthritis, *Lancet* (2010), 376:1094-108.

11. Tedesco, A; D'Agostino, D; Soriente, I; Amato, P; Piccoli, R. and Sabatini, P. : A new strategy for the early diagnosis of rheumatoid arthritis: a combined approach. *Autoimmunity Reviews*.(2009), 8:233–7.
12. Roberts, L.J; Cleland, L.G; Thomas, R. and Proudman, S.M; Early combination disease modifying anti-rheumatic drug treatment for rheumatoid arthritis. *Medical Journal of Australia* (2006),184:122–5.
13. Firestein, G.S: Etiology and pathogenesis of rheumatoid arthritis, In: Harris, E.D; Budd, R.C; Genovese, M.C; Firestein, G.S; Sargent, J.S, and Sledge, C.B.: *Kelley's Textbook of Rheumatology*, Saunders Elsevier, Philadelphia, Pa, USA, (2005),7, 996–1042
14. Kore, K.J and Shete, R.V. Anti-Arthritic activity of Hydro alcoholic extract of *Lawsonia innermis* against adjuvant arthritis. *Int. j. drug dev & res* (2011), 3(4): 217-224
15. Shin, H.Y, Jeong – tang inhibits the stem cell factor-induced migration and inflammatory cytokines secretion in mast cells. *J.Ethanopharmacology* (2003), 85: 157-161.
16. Bevaart, L. and Vervoordeldonk, M.J; Evaluation of Therapeutic Targets in Animal Models of Arthritis How Does It Relate to Rheumatoid Arthritis? *Arthritis and Rheumatism*. (2010), 62(8):2192–2205.
17. Ewa, M, P.: Angiogenesis in Rheumatoid Arthritis. *Arthritis Res*, (2002), 4(3): S81-S90.
18. Bansod, M.S: Therapeutic Effect of A Poly Herbal Preparation On Adjuvant Induced Arthritis In Wistar Rats. *Int. Journal of Pharm Sciences*(2011), 3 (2): 186-192
19. Buadonpri, W: Synthetic Curcumin inhibits Carragenan-induced Paw edema in rats. *J Health Res*. (2009), 23(1):11-16.
20. Pandey,S.: Arthritis an autoimmune disorder: Demonstration of In-vivo anti-arthritic activity. *Inter J Pharm Life Sci*. (2010), 1(1):38-43.
21. Telang, R.S.: Studies on analgesic and anti- inflammatory activities of Vitex negundo. *Indian Journal of Pharmacology* (1999), 31(5): 363-366.
22. Lee, J.H; Zhou, H.Y; Cho, S.Y; Kim, Y.S; Lee, Y.S and Jeong, C.S. : Anti-inflammatory mechanisms of apigenin: inhibition of cyclooxygenase-2 expression, adhesion of monocytes to human umbilical vein endothelial cells, and expression of cellular adhesion molecules. *Arch Pharm Res* (2007), 30(10): 1318-27.
23. Wilder, R.L: Genetic factors regulating experimental arthritis in mice and rats. *Curr Dir Autoimmun* (1999), 21–65.
24. Kleinau, S and Erlandsson, H : Adjuvant oils induce arthritis in the DA rat I. Characterization of the disease and evidence for an immunological involvement. *J. Autoimmune* (1991), 4: 871–80.
25. Wilder, R.L: Streptococcal cell-wall-induced arthritis in rats: an overview; *Int J Tissue React*(1988), 10:1–5.
26. Richard A. Harvey. Lippincott's illustrated reviews, pharmacology 5 th edition (2011),538-543.
27. Shivanand, P.: *International Journal of Pharmacy & Life Sciences*(2010), 1: 38-43.
28. Agrawal, S; Misra, R. and Aggarwal, A.: Autoantibodies in rheumatoid arthritis: association with severity of disease in established RA. *Clin Rheumatol* (2007), 26: 201– 4.
29. White, B. Ginger: an overview . *The American Family Physician*, (2007),75: (11)1689–1691.
30. Al-Nahain, A.; Jahan , R. and Rahmatullah, M.: *Zingiber officinale*: A Potential Plant against Rheumatoid Arthritis. *Arthritis* (2014), ID 159089, 8 pages. <http://dx.doi.org/10.1155/2014/159089>
31. Mishra, R.K.; Kumar, A. and Kumar, A. : Pharmacological activity of *Zingiber officinale*. *International Journal of Pharmaceutical and Chemical Sciences*.(2012),1(3): 1073–1078.
32. Yoshikawa, M ; Hatakeyama, S; Chatani, N; Nishino, Y and Yamahara, J. : Qualitative and quantitative analysis of bioactive principles in *Zingiberis rhizoma* by means of high performance liquid chromatography and gas liquid chromatography. On the evaluation of *Zingiberis rhizoma* and chemical change of constituents during *Zingiberis rhizoma* processing. *Yakugaku Zasshi*. (1993),113:307–315.
33. Yang, Z.; Yang, W.; Q. and Peng et al., Volatile phytochemical composition of rhizome of Ginger after extraction by headspace solid-phase micro extraction, petrol ether extraction and steam distillation extraction. *Bangladesh Journal of Pharmacology*. (2009), 4(2):136–143.
34. Hsu, Y.L.; Chen, C.Y.; Hou M.F. et al. : 6-dehydrogingerdione, an active constituent of dietary Ginger, induces cell cycle arrest and apoptosis through reactive oxygen species/c-Jun N-terminal kinase pathways in human breast cancer cells. *Molecular Nutrition and Food Research*. (2010), 54(9):1307–1317.
35. Grzanna, R.; Lindmark, L. and Frondoza, C. G. Ginger a herbal medicinal product with broad anti-inflammatory actions. *Journal of Medicinal Food* (2005), 8(2):125–132.

36. Haghghi, M.; Khalvat, A. Toliat, T. and Jallaei, S. Comparing the Effects of Ginger (*Zingiber Officinale*) Extract and Ibuprofen on Patients with Osteoarthritis. *Archives of Iranian medicine.* (2005), 8(4): 267-271.
37. Xinangcai, X. (1998). Complete External Therapies of Chinese Drugs, Foreign Languages Press, Beijing, Peoples Republic of China.
38. Minghetti, P.; Sosa, S.; Cilurzo, F.; Casiraghi, A.; Alberti, E.; Tubaro, A.; Loggia, R. and Montanari, L. Evaluation of the Topical Anti-Inflammatory Activity of Ginger Dry Extracts from Solutions and Plasters. *Planta medica* (2007), 73(15):1525-1530.
39. Fingado, M. (2012) Compresses and therapeutic applications, translator Tessa Therkleson and Sarah Therkleson, Floris Books, Edinburgh, United Kingdom.
40. Therkleson, T. Ginger Compress Therapy for Adults with Osteoarthritis. *Journal Advanced Nursing*, (2010). 66(10): 2225 – 2233.
41. Ammon, H.P and Wahl, M.A. Pharmacology of *Curcuma longa*. *Planta Med* (1991),57:1-7.
42. Louay, L.: Medicinal and pharmacological properties of Turmeric (*Curcuma longa*): A review. *Int J Pharm Biomed Sci.* (2014), 5(1):17-23.
43. Basnet, P. and Skalko-Basnet, N. Curcumin: an anti-Inflammatory Molecule from a Curry Spice on the Path to Cancer Treatment. *Molecules* (2011), 16(6): 4567-98.
44. Funk, J.L; Oyarzo, J.N and Frye, J.B et al. Turmeric extracts containing curcuminoids prevent experimental rheumatoid arthritis. *J. Nat. Prod.* (2006), 69(3): 351.
45. Goel, A; Kunnumakkara, A.B and Aggarwal, B.B. : Curcumin as curecumin: from kitchen to clinic. *Biochem Pharmacol* (2008),75(4): 787-809.
46. Huang, M.T; Lysz, T and Ferraro, T. et al. Inhibitory effects of curcumin on in vitro lipoxygenase and cyclooxygenase activities in mouse epidermis. *Cancer Res* (1991),51:813-819.
47. Mahe, A.; Khan, H.; Samiullah, L. and Siddique, K.M. : A review on Phytochemical and Pharmacological studies of Kundur (*Boswellia serrata* Roxb ex Colebr.) A Unani drug. *Journal of Applied Pharmaceutical Science* (2012), 2 (3):148-156.
48. Anderson, J; Davis, M; Joly, F. and Puteaux, S.,: Boswellic Acids: Potent Active Ingredients from a Traditional Remedy. *Cosmetic Science Technology.* (2007), 100-105.
49. Abdel-Tawab, M; Werz, O. and Schubert-Zsilavec, M. : *Boswellia serrata*: an overall assessment of in vitro, preclinical, pharmacokinetic and clinical data. *Clin Pharmacokinet.* (2011), 50(6):349-369.
50. Gupta, P.K; Samarakoon, S.M; Chandola, H.M and Ravishankar, B. Clinical evaluation of *Boswellia serrata* (Shallaki) resin in the management of Sandhivata (osteoarthritis). *Ayu.* (2011),32(4): 478-482.
51. Ammon, M. T.; Safayhi, H.; Mack, T. and Sabieraj, J. Mechanism of anti-inflammatory actions of curcumine and boswellic acids. *Journal of Ethnopharmacology.* (1993), 38(2, 3): 113-119.
52. Singh, G. B.; Surjeet, S.; Bani, S. and Kaul A. Boswellic acids - a new class of anti-inflammatory drugs with a novel mode of action. International Seminar Traditional Medicine, *Calcutta.* (1992), 81-82
53. Safayhi, H., 5-lipoxygenase inhibition by acetyl-11-keto-boswellic acid (AKBA) by a novel mechanism, *Phytomedicine*, (1996), 71-2.
54. Sailer, E.R., et al., Structure activity relationships of the nonredox-type noncompetitive leukotriene biosynthesis inhibitor acetyl-11-keto-boswellic acid, *Phytomedicine*, (1996), 73-74.
55. Safyhni, H., et al., Concentration-dependent potentiating and inhibitory effects of Boswellia extracts on 5-lipoxygenase product formation in stimulated PMNL, *Planta Medica*, (2000), 110-3.
56. Safayhi, H., et al., Inhibition by boswellic acids of human leukocyte elastase, *Journal of Pharmacology and Experimental Therapeutics*, (1997), 460-3.
57. Faixova, Z and Faix,S.:Biological effects of rosemary *Rosmarinas officinalis* L essential oil (A Review): *FOLIA VETERINARIA* (2008), 52, 3-4: 135-139.
58. AL- Sereitia, M.R.; Abu-Amerb, K.M. and Sena, P. Pharmacology of *Rosemarinus officinalis* Linn. and its Therapeutic Potentials. *Indian Journal of Experimental Biology.* (1999), 37: 124-131.
59. Beninca, J.P.; Dalmarco, J.B; Pizzolatti, M.G and Frode, T.S Analysis of the anti-inflammatory properties of *Rosmarinus officinalis* L. in mice. *Food Chem.* (2011), 124:468–475.
60. Altinier, G.; Sosa, S.; Aquino, R.P; Mencherini, T; Delia Loggia, R. and Tubaro, A.: Characterization of topical antiinflammatory compounds in *Rosmarinus offieinalis* L. *J. Agrie Food Chem.*(2007), 55(5): ni S-n 23.
61. Miühlbauer, R.C.; Lozano, A.; Palacio, S.; Reinli, A.and Felix, R. Common herbs, essential oils and monoterpenes potently modulate bone metabolism. *Bone* (2003),32: 372-380

62. Liang, Y.; Lu, J.; Zhang, L.; Wu, S.; and Wu, Y.; Estimation of black tea quality by analysis of chemical composition and colour difference of tea infusions. *Food Chem.*, (2003), 80: 283-290.
63. Kaur, A.; Nain, P. and Nain, A. Herbal Plants used in treatment of rheumatoid arthritis: a review. *Int. J. Pharm. Pharmaceut. Sci.*,(2012). 4 : (Supple 4) 44-57.
64. Sharangi, A.B. Medicinal and therapeutic potentialities of tea (*Camellia sinensis* L.) A review. *Food Res. Int.*, (2009), 42: 529-535.
65. Ahmed, S. Green tea polyphenol epigallocatechin 3-gallate in arthritis: progress and promise. *Arthritis Res. Ther.*, (2010), 12: 1-9.
66. Nanjo, F; Mori, M; Goto, K and Hara, Y.: Radical Scavenging Activity of Tea Catechins and Their Related Compounds. *Biosci Biotechnol Biochem* (1999), 63(9): 1621-3.
67. Kurien, B.T; Hensley, K; Bachmann, M. and Scofield, R.H. : Oxidatively modified autoantigens in autoimmune diseases. *Free Radic Biol Med* (2006), 41(4): 549-56
68. Singh, R; Akhtar, N and Haqqi, T.M. Green tea polyphenol epigallocatechin-3-gallate: inflammation and arthritis. *Life Sci* (2010), 86(25-26): 907-18.
69. Datta, P.; Sarkar, A.; Biswas, A.K. and Gomes, A. Anti-arthritic activity of aqueous extract of Indian black tea in experimental and clinical study. *Orient Pharm. Exp. Med.*, (2012),12: 265–271.
70. Sang, I.K.L.; Pung, M.H.; Seung, H.K.; Suppression of the onset and progression of collagen-induced arthritis by chebulagic acid screened from a natural product library. *Arthritis Rheum* (2005), 52: 345-53.
71. Raju, D.; et al : Evaluation of anti-ulcer activity of methanolic extracts of *T. chebula* fruits in experimental rats, *J. Pharmaceutical sciences & research*,(2009), 1(3): 101-107
72. Nair, V; Singh, S and Gupta, Y.K. Anti-arthritic and disease modifying activity of *Terminalia chebula* Retz. In experimental models. *J Pharm Pharmacol.* (2010),62(12): 1801-06.
73. Ramani, Y.R and Pradhan, S. Anti-arthritic Activity of Acetone Extract of *Terminalia Chebula*. *Web med Central Pharmacology* (2012),3(2):
74. Ray, P.G. and Majumdar, S.K.: Antimicrobial activity of some Indian plants. *Econ. Bot*, (1976),120-131.
75. Park, M.K; Park, J.H; Kim, N.Y; Shin, Y.G; Choi, Y.S; Lee, J.G; et al. : Analysis of 13 phenolic compounds in Aloe species by high performance liquid chromatography *Phytochem Anal*, (1998), 9(4):186–91.
76. Groom, Q.J and Reynolds, T.: Barbaloin in aloe species. *Planta Med*, (1987). 53(4):345–8.
77. Pankaj, K. S.; Deen D. G.; Ritu S. ; Priyanka P.; Sharmistha G.; Atul K. S.; Ajay K. and Kapil D. P.: Therapeutic and Medicinal Uses of *Aloe vera*: A Review *Pharmacology & Pharmacy* (2013), 4: 599-610
78. Davis, R.H.; Parker, W.L.; Samson, R.T. and Murdoch, D. P.: Isolation of a Stimulatory System in an Aloe Extract: *Journal of the American Podiatric Medical Association* (1991), 81: 473-478.
79. Hanley, D.C.; Solomon, W.A.; Saffran, B. and Davis, R. H. : The Evaluation of Natural Substances in the Treatment of Adjuvant Arthritis,” *Journal of the American Podiatric Medical Association*.(1982) 72: 275-284
80. Chitme, H.R. and Patel, N.P.: *The Open Natural Products Journal* (2009), 2: 6-15.
81. Tripathy, S; Pradhan, D and Anjana, M. Anti-inflammatory and anti-arthritic potential of *Ammania baccifera* linn. *International Journal of Pharma and Bio Sciences*.(2010),1(3): 17.
82. Kumar, V.L. and Roy, S. *Calotropis procera* latex effect affords protection against inflammation and oxidative stress in Freund's complete adjuvant induced monoarthritis in rats. *Mediators of Inflammation*, (2007), ID: 47523: 1-7.
83. Babu, S.A.R. and Karki, S.S. Anti-inflammatory activity of various extracts of roots of *Calotropis procera* against different inflammation models. *Int. J. Pharm. Pharmaceut. Sci.*, (2011), 3: 191-194.
84. Feng , X. Anti-arthritic active fraction of *Capparis spinosal*. Fruits and its chemical constituents, *Yakagakuzasshi*, (2011), 131(3): 423-429.
85. Sheetal, S.C.: Analgesic, anti-inflammatory and anti-arthritic activity of *Cassia uniflora* Mill., *Asian Pacific Journal of Tropical Biomedicine*, (2012), S181-S186.
86. Chippada, S.C.: .Antioxidant, anti-inflammatory and anti-arthritic activity of *Centella asiatica* extracts, *J. Chem Bio Phy Sci*.(2011),1(2):260–269.
87. Chakraborty, A.K.: Evaluation of anti-arthritic activity of ethanolic extract of *Cleome rutidosperma*. *Journal of Pharmaceutical Science and Technology* (2010), 2(10): 330-332

88. Duwiejua, M.; Zeitlin, I.J.; Waterman, P.G.; Chapman, J.; Mhango, G.J. and Provan, G.I. Anti-inflammatory activity of resins from some species of the plant family Burseraceae. *Planta Med.* (1993), 59:12-15.
89. Mishra, L.C. :Rheumatoid arthritis, osteoarthritis and gout. In: Scientific Basis of Ayurvedic Therapy (ed). Mishra, L.C., C.R.C Press, USA, pp (2003), 167-201.
90. Bothara, S.B.: Anti-arthritis activity of root extracts of *Cocculus hirsutus*. *International Journal Pharmacy and Pharmaceutical Sciences.* (2011), 3: 175-177.
91. Biradar, S.; Kangra, V.A.; Mandavkar, Y.; Thakur, M. and Chougule, N. Anti-inflammatory, anti-arthritis, analgesic and anticonvulsant activity of *Cyperus* essential oils. *Int. J. Pharm. Pharmaceut. Sci.* (2010), 2: 112-115
92. Hye, H.K. and Gafur, M.A. Anti-inflammatory and anti-arthritis activity of a substance isolated from *Dalbergia volubilis*. *Ind. J. Med. Res.*, (1975),163: 93-100.
93. Vasudevan, M.; Gunnam, K.K. and Parle, M. (2006). Anti-nociceptive and anti-inflammatory property of *Daucus carota* seed extract. *J. Health Sci.*, 52: 598-606.
94. Murugananthan, G: Anti-inflammatory and anti- arthritis activities of *Delonix elata* bark extract. *International Journal of Research in Ayurveda & Pharmacy.* (2011), 2(6): 1819-1821.
95. Ramasamy, S. et al: *Indo-Global Research Journal of Pharmaceutical Sciences* (2012), (2): 378-382.
96. Harpalani , A.N.: Anti-inflammatory and antiarthritis potential of aqueous and alcoholic ext racts of *Euphorbia antiquorum*linn. *Pharmacologyn line.* (2011), (2): 287-298.
97. Manocha, N.; Chandra, S.K.; Sharma, V.; Sangameswaran, B. and Saluja, M. *Research Journal of Chemical Sciences* (2011), 1: 2-8.
98. Mishra, N.K., Anti-arthritis activity of *Glycyrrhiza glabra*, *Boswellia serrata* and their synergistic activity in combined formulation studied in Freund's adjuvant induced arthritis rats, *J. Pharm Educ Res*, (2011), 2(2):92-98.
99. Ramesh, P.R.: *International Journal of Pharma and Bio Sciences* (2012), (3): 328-336.
100. Mehta, A.; Sethiya, N.K.; Mehta, C. and Shah, G.B. Antiarthritis activity of roots of *Hemidesmus indicus* R.Br. (Anantmul) in rats. *Asian Pac. J. Trop. Med.* (2012), 5: 130-135.
101. Alam, M.L. and Gomes, A. Viper venom-induced inflammation and inhibition of free radical formation by pure compound (2-hydroxy-4-methoxy benzoic acid) isolated and purified from anantamul (*Hemidesmus indicus* R. Br.) root extract. *Toxicon*, (1998), 36: 207-215.
102. Paval, J.; Kaitheri, S.K.; Potu, V.K.; Govindan, S. and Kumar, R.S. Antiarthritis potential of the plant *Justicia gendarussa* Burn. *F. Clinics*, (2009), 64: 357-362
103. Kripa, K.G.; Chamundeeswari, D.; Thanka. J. and Reddy, C.U.M. Effect of hydro alcoholic extract of aerial parts of *Leucas aspera* (Willd.) link on inflammatory markers in complete Freund's adjuvant induced arthritis rats. *Int. J. Green Pharm.* (2010), 4: 281-287.
104. Tatiya, A.U. and Saluja, A.K.: *Brazilian Journal of Pharmacognosy* (2011), (21): 1052-1064
105. Kamalutheen, M.; Gopalakrishnan, S. and Syed Ismail, T. : Anti-inflammatory and antiarthritis activities of *Merremia tridentata* (L) Hall. *E-Journal of Chemistry.* (2009), 6: 943-948.
106. Bhalariao, A.R.; Desai, S.K.; Serathia, B.R.; Vartak, K.M. and Doshi, G.M. Antiarthritis studies on *Nyctanthes arbortristis* and *Maharasnadi ghan*. *Scholars Research Library.* (2011), 3: 101-110.
107. Sandhar, H.K.; Kaur, M.; Kumar, B. and Prasher, S. An update on *Nyctanthes arbortristis* Linn. *Int. Pharmaceut. Sci.*, (2011), 1: 77-86.
108. De, S.; Ravishankar, B. and Bhavasar, G.C. Investigations of the anti-inflammatory effects of *Paederia foetida*. *J. Ethnopharmacol.* (1994), 43: 31-38.
109. Singh, S.; Bani, S.; Khajuria, A.; Sharma, M.L.; Singh,G.B.; Suri, K.A. and Srivastava,T.N. Anti-inflammatory activity of *Paederia foetida*. *Fitothetrapia*, (1994), 64: 357-362.
110. Mali. : Anti-arthritis activity of standardised extract of *Phyllanthus amarus* in Freund's complete adjuvant induced arthritis. *Biomedicine & Aging Pathology*,(2011), (1):185-190.
111. Samuel K, Antiarthritis effect of aqueous and ethanolic leaf extracts of *Pistiastratiotes* in adjuvant-induced arthritis in Sprague-Dawley rats, *Journal of Experimental Pharmacology*, (2012) ; (4): 41–51.
112. Arote, S.R.: *Journal of Biomedical and Pharmaceutical Sciences* (2011), (1): 16-23.
113. Rajendran, R. and Krishnakumar, E. Antiarthritis activity of *Premna serratifolia* Linn. wood against adjuvant induced arthritis. *Avicenna J. Med. Biotech.* (2010). 2: 101-106.
114. Kothari, A.: *Journal of Pharmacy Research* (2011), (4): 4126-4128.
115. Patel, R.G.: *World Journal of Pharmaceutical Research* (2012), (1): 309-325.

116. Kabra, M.P. and Rachhadiya, R.M.: Pharmacological investigation of hydroalcoholic extract of *Ricinus communis* leaves in arthritis induces rats, *Asian Journal of Biochemical and Pharmaceutical Research*, (2011), 4(1): 310-321.
117. Saravanan, S.; Babu, N.P.; Pandikumar, P. and Ignacimuthu, S. Therapeutic effects of *Saraca asoka* (Roxb.) Wilde on lysosomal enzymes and collagen metabolism in adjuvant induced arthritis. *Inflammo pharmacology*, (2011),19: 317-325
118. Ekambaram, S.: Evaluation of anti-arthritic activity of *Strychnos potatorum* Linn seeds in Freund's adjuvant induced arthritic rat model, *BMC Complementary and Alternative Medicine*. (2010), (10):56.
119. Uma Chandur, S.; Anti-Arthritic Activity of Root of *Saussurea lappa*; *Pharmacologia* (2011), 2 (9).
120. Gupta, S.R.: *Nat. Prod. Res* (2009) (23): 689-695.
121. Paval, J., et al., *Journal of Herbal Medicine and Toxicology* (2011), (5):11-16.
122. Abudoleh, S. and Disi, A.: Anti-arthritic activity of the methanolic leaf extract of *Urtica pilulifera* L. on albino rats. *American Journal of Pharmacology and Toxicology* (2011), 6(1): 27-32.
123. Rahman, M.M.: Anti- inflammatory, anti-arthritic and analgesic activity of the alcoholic extract of the plant *Urginea indica* kunth. *International Journal of Pharmaceutical Sciences and Research*.(2011), 2(11):2915-2919.
124. Otari, K.V.: Evaluation of Antiinflammatory and antiarthritic activities of ethanolic extract of *Vernonia anthelmintica* seeds, *Journal of Cell and Tissue Research*. (2010), 10(2): 2269-2280.
125. Panchal, A.H., *Pharmacology online* (2011), 3:175-187.
126. Anbalagan, K. and Saddique, J. Influence of an Indian medicine (ashwagandha) on acute phase reactance in Inflammation. *Indian J. Exp. Biol.* (1981), 19: 245-249.
127. Al-Hindawi, M.K.; Khafaji, S.H. and Abdul-Nabi, N.H. Anti-granuloma activity of Iraqi *Withania somnifera*. *J. Ethnopharmacol.* (1992), 37: 113-116.
128. Yoon, H.Y; Lee, E.G; Lee, H. *et al.* Kaempferol inhibits IL-1 β -induced proliferation of rheumatoid arthritis synovial fibroblasts and the production of COX-2, PGE2 and MMPs. *Int J Mol Med* (2013); 32(4): 971-7.
129. Kim S, Jun C, Suk K, et al. Gallic Acid Inhibits Histamine Release and Pro-inflammatory Cytokine Production in Mast Cells. *Toxicol Sci* 2006; 91(1): 123-31.
130. Hirata, A; Murakami, Y; Atsumi, T.: *et al.* Ferulic Acid Dimer Inhibits Lipopolysaccharide-stimulated Cyclooxygenase-2 Expression in Macrophages. *In Vivo* (2005), 19(5): 849-53.
131. Qin, J; Shang, L; Ping, A. *et al.* : TNF/TNFR signal transduction pathway-mediated anti-apoptosis and anti-inflammatory effects of sodium ferulate on IL-1 β -induced rat osteoarthritis chondrocytes *in vivo*. *Arthritis Res Ther* (2013), 15(3): 407
132. Lee, J; Kim, S and Kim, T: Anti-inflammatory Effect of Bee venom on Type II Collagen- Induced Arthritis. *The American Journal of Chinese Medicine* (2004), 32(3):361-367.
133. Sun, Q.W; Jiang, S.M; Yang, K; Zheng, J.M; Zhang, L and Xu, W.D. : Apigenin enhances the cytotoxic effects of tumor necrosis factor-related apoptosis-inducing ligand in human rheumatoid arthritis fibroblast-like synoviocytes. *Mol Biol Rep* (2012); 39(5): 5529-35.
134. Shin, G.C; Kim, C and Lee, J.M. *et al.* Apigenin-induced apoptosis is mediated by reactive oxygen species and activation of ERK1/2 in rheumatoid fibroblast-like synoviocytes. *Chem Biol Interact* (2009); 182(1): 29-36.
135. Nicholas, C; Batra, S. and Melissa, A. *et al.* : Apigenin Blocks Lipopolysaccharide-Induced Lethality *In Vivo* and Proinflammatory Cytokines Expression by Inactivating NF- κ B through the Suppression of p65 Phosphorylation. *J Immunol* (2007), 179(10): 7121-7.
136. Gong, J; Shin, D; Han, S; Kim, J. *et al.*: Kaempferol suppresses eosinophil infiltration and airway inflammation in airway epithelial cells and in mice with allergic asthma. *J Nutr* (2012); 142(1): 47-56
137. Kaussa, T; Ramberta, J; Fawazb, F. *et al.* Therapeutic and preventive properties of quercetin in experimental arthritis correlate with decreased macrophage inflammatory mediators. *Biochem Pharmacol* (2006), 72(10): 1304-10
138. Jackson, J.K; Higo, T; Hunter, W.L and Burt, H.M. : The antioxidants curcumin and quercetin inhibit inflammatory processes associated with arthritis. *Inflamm Res* (2006), 55(4): 168-75.
139. Choi, E.J; Bae, S.C; Yu, R; Youn, J and Sung, M.K. : Dietary vitamin E and quercetin modulate inflammatory responses of collagen-induced arthritis in mice. *J Med Food* (2009),12(4): 770-5.
140. Shakibaei, M; Csaki, C; Nebrich, S and Mobasheri, A. : Resveratrol suppresses interleukin-1 β -induced inflammatory signaling and apoptosis in human articular chondrocytes: potential for use as a novel nutraceutical for the treatment of osteoarthritis. *Biochem Pharmacol* (2008), 76(11): 1426-39.

141. Leonarda, H.; Xiab S.S; Jiangb, C;, B. *et al.* :Resveratrol scavenges reactive oxygen species and effects radical-induced cellular responses. *Biochem Biophys Res Commun* (2003), 309(4): 1017-26.
142. Csaki, C; Keshishzadeh, N; Fischer, K and Shakibaei, M.: Regulation of inflammation signalling by resveratrol in human chondrocytes *in vitro*. *Biochem Pharmacol* (2008), 75(3): 677-817.
143. Xuzhu, G ; Leung, B.P; Howe, H.S. *et al.* : Resveratrol modulates murine collagen-induced arthritis by inhibiting Th17 and B-cell function. *Ann Rheum Dis* (2012), 71(1): 129-35.
144. Zhang, Y; Dong, J and Zhang Q, *et al.* : Genistein inhibit cytokines or growth factor-induced proliferation and transformation phenotype in fibroblast-like synoviocytes of rheumatoid arthritis. *Inflammation* (2012), 35(1): 377-87.
145. Jia, Z; Babu, P.V; Si, H. *et al.* : Genistein inhibits TNF- α -induced endothelial inflammation through the protein kinase pathway A and improves vascular inflammation in C57BL/6 mice. *Int J Cardiol* (2013), 168(3): 2637-45.
146. Zhao, F; Wang, L and Liu, K.: *In vitro* anti-inflammatory effects of arctigenin, a lignan from *Arctium lappa* L., through inhibition on iNOS pathway. *J Ethnopharmacol* (2009), 122(3): 457-62.
147. Hyam, S.R; Lee, I.A; Gu, W. *et al.*:Arctigenin ameliorates inflammation *in vitro* and *in vivo* by inhibiting the PI3K/AKT pathway and polarizing M1 macrophages to M2-like macrophages. *Eur J Pharmacol* (2013), 708(1-3): 21-9.
148. Yassin, A; Gareeb, Hala; Mohamed, A. N and Samy, C. : The Relationship between Vitamin D and Disease Activity in Egyptian Patients with Rheumatoid Arthritis : *INTERNATIONAL TRENDS IN IMMUNITY* (2014), 2 (3):122-127.
