

Anti hyperlipidemic and Antioxidant Activity of Extract *Cinnamomum zeylanicum* in male Rats Fed a High Fat Diet

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Abstract : The study was divided into four groups (7 rats in each group) for 8 weeks in 3 days in each week. Include gives high cholesterol diet (Libitum). Group 1: control animals give 0.5 ml/kg of normal saline only. Group 2: administrated with 100g/kg of volume dose from libitum only. Group 3: was administrated with 0.5 ml/kg of volume dose from ECE at concentration 250 mg/kg plus 100g/kg of libitum. Group 4: administrated with 0.5 ml/kg of volume dose from ECE at concentration 500 mg/kg plus 100 g/kg of libitum. At the end of treatment period (8 weeks), rats were scarified, serum sample obtained for assessment of antioxidant parameters and lipid profile, the result indicated that ECE 250 and 500 mg/kg show a significant decline ($p < 0.05$) in activities of MDA and a significant raise ($p < 0.05$) in GSH levels. This results showed the concentration 500 mg/kg of ECE have high effect of antioxidant. In line of high cholesterol diet, the result shown, that libitum have significantly increased ($p < 0.05$) in total cholesterol, TG, LDL, HDL and VLDL and body weight but when gives libitum with extract, the results shown that the parameters near to normal.

Key words: Hyperlipidemia, ECE, Lipid profile.

Introduction

Hyperlipidemia refers to hypercholesterolemia, hypertriglyceridemia and hyperlipoproteinemia, are an important risk factor for develop cardiovascular diseases. Hyperlipidemia is increase level of total cholesterol, triglycerides and low-density lipoprotein cholesterol among decrease in high-density lipoprotein cholesterol. The predictor of coronary artery disease, fatty liver disease and carcinogenesis which is associated with the formation of reactive oxygen species^{1,2}.

Antioxidant means "against oxidation", antioxidants work to guard lipids, proteins and nucleic acids by radicals from peroxidation. They hinder or interval the oxidation of other particles by stopping the beginning or spread of oxidizing chain reactions. "Antioxidants are effective because they are ready to give up their possess electrons to free radicals. After a free radical gains the electron from an antioxidant it no lengthier needs to attack the cell and the chain reaction of oxidation is destroyed^{3,4}.

'Cinnamon', *Cinnamomum zeylanicum* (Lauracea), locally known as Qerfah or Darsin is an older and essential spice with wide-ranging requests in flavoring, perfumery, drinks and drugs⁵. In some previous studies, an essential oil of cinnamon is identified to have antibacterial⁶. Previously, some studies also suggested that cinnamon owns strong free radical scavenging capability⁵, antioxidant and antimutagenic³ activities and properties of LDL⁶.

Materials and Methods

Preparation of Plant Ethanol Extract

To formation ethanolic cinnamon extract (ECE), 20 g of cinnamon powder were kept in thimble was extracted with 200 ml 90% ethanol in a soxhlet extractor for 24 hour. The extract was concentrated in a vacuum at 60 C° using rotary evaporator, to evaporate the remaining solvent. The extract was kept in a freeze dryer for 24 hour yielding semisolid residues of extract⁷.

Preparation of doses (Libitum)

High cholesterol diet (HCD) mixture was made by addition cholesterol (100g), cholic acid (50g) in 1000 ml of coconut oil added with egg. The coconut oil contain many of fatty acid such as; Lauric acid 45-50%, Myristic acid 13-20% , Palmitic acid 7-10% , and Caprylic acid 5-10%⁸.

Biochemical analysis

Determination of Serum Glutathione Activity

The test is intended for quantitative of glutathione concentration in serum through the enzyme linked immunosorbant assay (ELISA) using bio Elisa reader EL x800 (biokit, U.S.A.).The assay Max Glutathione ELISA kit was achieved according to the manufacturing company (CUSABIO, U.S.A.)¹⁹.

Determination of Lipid Peroxidation Activity (MDA)

Cell Bio labs TM MDA Adduct ELISA Kit (USA) is an enzyme immunoassay¹⁰.

Determination of lipid profile activity

Total cholesterol kit for quantitative determination of total cholesterol in human serum was supplied by Biolabo SA, France,

Serum HDL-Cholesterol level was measured by HDL-Cholesterol phosphotungstic acid (PTA) precipitant kit (Biolabo SA, France) Triglycerides Kit was supplied by Biolabo SA, France. for measureable of triglycerides in human serum. Very Low Density Lipoprotein (VLDL) were measured by the next principle: $VLDL = TG (mmol/l) / 5$ and Low Density Lipoprotein (LDL) were measured by the next formula: $LDL = TC(mmol/l) - VLDL(mmol/l) - HDL(mmol/l)$ ¹¹.

Biostatistical Analysis

The results were expressed as (mean \pm standard deviation). Pooled t- test was used for the comparison between control and other groups in the measured parameters. One way analysis of variance (ANOVA) followed by least significant difference (L.S.D.) analyses at 0.05% probability of levels. All statistical analysis were performed using Excel program (2010) from Microsoft Company. USA. and MegaStat. The difference will be significant when $P < 0.05$ value.

Results

No significant difference was show in initial weight in animals gives a libitum when compared all groups with control. Significant increase in final weight in animals dives a libitum when compared groups of libitum plus ECE 250 mg/kg with control, also show significant raise in liver weight and spleen weight in rats fed a libitum when compared all groups with control.

Table1: Activity of *Cinnamomum zeylanicum* on weights of Animals and their organs in rats fed high cholesterol diet (libitum)

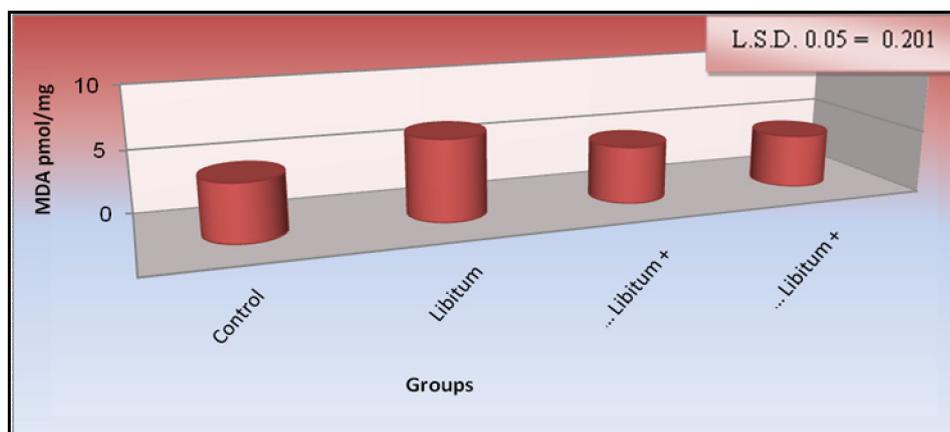
Groups	Initial weight	Final weight	Liver weight	Kidney weight	Spleen weight
Control	178.00 ± 8.367	200.00 ± 7.071	5.82 ± 1.089	2.28 ± 0.259	1.88 ± 0.217
Libitum	182.00 ± 8.367	205.80 ± 9.121	*8.44 ± 0.251	*3.08 ± 0.239	*3.80 ± 0.274
Libitum + ECE 250	180.68 ± 8.245	*182.44 ± 10.003	*6.32 ± 0.164	2.42 ± 0.084	*2.82 ± 0.130
Libitum + ECE 500	180.00 ± 10.000	181.00 ± 10.05*	*6.04 ± 0.089	2.32 ± 0.192	2.74 ± 0.305*
L.S.D. 0.05	7.934	8.698	0.15	0.153	0.211

The effect of ECE on lipid profile, show nearby raised in total cholesterol in group of rat fed a libitum as compared with control. Cinnamon extract significantly decline in total cholesterol in group treated with libitum plus ECE 250 mg/kg and in group gives a libitum plus ECE 500 mg/kg body weight in contrast with rat group fed on libitum only.

Table2: Activity of *Cinnamomum zeylanicum* on lipid profile in rats fed high cholesterol diet (libitum)

Groups	TC	TG	HDL	LDL	VLDL
Control	90.8 ± 1.10	74.6 ± 0.55	40.4 ± 1.14	42.8 ± 2.59	14.8 ± 1.48
Libitum	*223.0 ± 10.95	*199.8 ± 7.09	*32.6 ± 1.14	*77.4 ± 5.59	*48.8 ± 6.69
Libitum + ECE 250	*151.2 ± 1.30	*157.0 ± 2.74	*34.6 ± 0.55	*55.8 ± 7.79	*34.8 ± 2.05
Libitum + ECE 500	120.8 ± 0.84*	76.0 ± 5.48	42.6 ± 1.95*	46.8 ± 1.92	15.6 ± 1.14
L.S.D. 0.05	3.904	4.561	1.08	4.564	2.944

A result show effect of ECE on malondialdehyde (MDA) and GSH, there were significant increase in MDA and GSH among rats fed on libitum as compared to control. Cinnamon extract significantly decrease in MDA and GSH in group treated with libitum plus concentrations cinnamon extract 250 mg/kg and in groups that gives a libitum plus concentrations cinnamon extract 500 mg/kg in contrast with groups of rats fed on libitum only.

**Figure1: Effect of *Cinnamomum zeylanicum* extract on levels of MDA in animals fed a libitum**

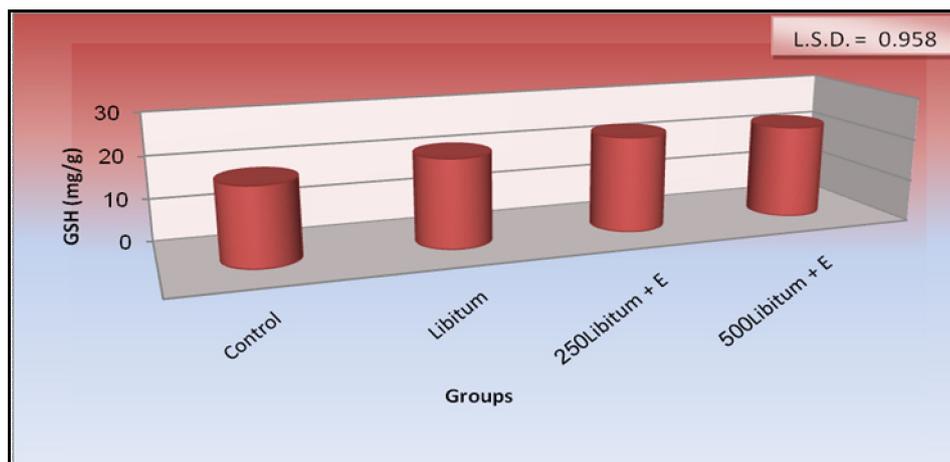


Figure2: Activity of *Cinnamomum zeylanicum* extract on level of GSH in animals fed a libitum

Discussion

Ethanollic cinnamon extract significantly increase ($p < 0.05$) in GSH. "This observation is consistent with administration of high cholesterol in experimental rats, this results are in agreement with those of others, who studied the effect of high fat diet^{12,13}, that study "Effects of *Lyciumbarbarum* aqueous and ethanol extracts on high fat-diet induced oxidative stress in rat liver tissue".

"It has been reported that lower GPx activity is generally accompanied with an increase of MDA concentration^{14,15}. "The ability of the extract to protect the heart and aorta against hypercholesterolemia-induced MDA lipid peroxidation and oxidative stress may explain its folklore use in the management of cardiovascular diseases^{16,17}.

The effect of ethanollic cinnamon extract on lipid profile, from obtained result it was observed that significant rise in total cholesterol in group of rat fed a libitum, in contrast to control rats. This results similar with⁸ "The mechanism of action of cholic acid is two folds: an increase in cholesterol absorption and a concomitant suppression of cholesterol 7 α -hydroxlyase activity that results in decreased cholesterol excretion"¹⁸⁻²⁰. Cholic acid increases absorption of cholesterol by its emulsifying.

Cinnamomum zeylanicum extracts have capability to dawn normalize free radicals raise, enhance liver, kidney and cholestatic biomarkers, perfect hepatic marker enzymes, decrease fibrosis severity **Cinnamomum zeylanicum** extracts have ability to dawn regulate lipid profile elevation²¹⁻²³.

Free radicals produced from CCl₄ made peroxidation of fat cell membrane which can be whole harm because it leads to modification, in the biological functions of membrane, such as amount of flexibility, and can lead to inactivation of membrane linked receptors or enzymes, which impaired normal cellular function. Lipid peroxidation product, the MDA commonly used as biomarker of oxidative stress²⁴⁻²⁹. The results in this study showed raised in lipid peroxidation in group treated with CCl₄ for 8-weeks compared to control.

References

1. Roberts, C. K.; Barnard, R. J.; Sindhu, R. K.; Jurczak, M.; Ehdaie, A. and Vaziri, N. D. (2006). Oxidative stress and dysregulation of NAD(P)H oxidase and antioxidant enzymes in diet induced metabolic syndrome. *Metabolism*, 55: 928–934.
2. Zainab Sajid , Abdul AL-Hadi Salil , Haider Salih (2016) ; Histological and Physiological study of the effect of prazosin hydrochloride on liver and kidney of rats (*Rattus norvegicus*); *International Journal of PharmTech Research* ; Vol.8, No.10, pp 72-80 .
3. Dekkers, J.; Doornen, L. and Kemper, H. (1996).The role of antioxidant vitamins and enzymes in the prevention of exercise-induced muscle damage. *Sports Medicines*, 21(3): 213-238.
4. Jayaprakasha, G. K.;Negi, P. S.; Jena, B. S.; Rao, J. L. (2007). Antioxidant and antimutajenic activities of *Cinnamomum zeylanicum*fruit extracts. *J. Food Compos Anal*, 20: 6-330.

5. Afyaa Sabah Nasir , Haider Salih Jaffat (2016); Effect of turmeric extract (*Curcuma longa*) on physiological parameters and neurotransmitters in rats treated by lithium carbonate; International Journal of PharmTech Research ; Vol.9, No.2, pp 89-97.
6. Jirovetz, L.;Buchbauer, G.; Ngassoum, M. B.; Essia, N.; Jean, J.; Tatsadjieu, L. N. and Adjoudji, O. (2002). Chemical composition and antibacterial activities of the essential oils of *Plectranthus glandulosus* and *Cinnamomum zylanicum* from Cameroon. *Sci. Pharmaceut.* 70:93-99.
7. Bafna, P. A. and Balaraman, R. (2004). Anti-ulcer and antioxidant activity of DHC-1, a herbal formulation. *J. Ethnopharmacol.*, 90:123-127.
8. Subash, B. P.;Prabuseenivasan, S. and Ignacimuthu, S. (2007). Cinnamaldehyde-a potential antidiabetic agent. *Phytomedicine*, 14:15-22
9. Hassan, F. A. (2011). Anti-Hepatotoxic Effect of the Methanolic *Anatica Hierocchuntica* Extract In CCl_4 - Treated Rats. *Eng. and Tech. Journal*, 29(20): 5-8.
10. Varsha, D.;Shinde, S.; Pawar, M. N. and Naikwade, S. (2010). Antihyperlipidemic Activity of *Cinnamomum tamala* Nees. on High Cholesterol Diet InducedHyperlipidemia. *Int. J. Pharm. Tech. Res.*,2(4): 2517-2521.
11. Brutis, C. A. and Ashwood, E. R. (1999). *Tietz textbook of Clinical Biochemistry*. 3rd ed., Saunders company, Tokyo, p. 1034-1054.
12. Armstrong, D. (1998). *In vitro* screening for antioxidant activity. *Free Radical and Antioxidant Protocols*, 108: 24-315.
13. Tietz, N.W. (1999). *Text book of Clinical Chemistry*, 3rd Ed., C.A. Burtis, E.R. Ashwood, W.B. Saunders . 703-1699.
14. Cui, B. K.; Liu, S.; Lin, X. J.; Wang, J.; Li, S. H.; Wang, Q. B.; Li, S. P. (2011). Effects of *Lyciumbarbarum* aqueous and ethanol extracts on high fat-diet induced oxidative stress in rat liver tissue. *Molecules*, 16, 9116–9128.
15. Afyaa Sabah Nasir , Haider Salih Jaffat . (2016); Protective role of turmeric extract (*Curcuma longa*) in the lipid profile and activity of antioxidant in the male rats treated by lithium carbonate; International Journal of PharmTech Research ; Vol.9, No.2, pp 98-105 .
16. Balogh, K.; Weber, M.; Erdélyi, M. and Mézes, M. (2004). Effect of excess selenium supplementation on the glutathione redox system in broiler chicken. *Acta. Vet. Hung.*, 52: 403-411.
17. Al-Dosari, M. S. (2011). Hypolipidemic and antioxidant activities of avocado fruit pulp on high cholesterol fed diet in rats. *Afr. J. Pharm.Pharmacol.*, 5: 1475–1483.
18. Abdul-Hadi Abbas Hadi , Haider Salih Jaffat (2016); Effect of Aluminum-Containing Antacid on Sperm Parameters And testicular Structure in Male Rats; International Journal of PharmTech Research ; Vol.9, No.3, pp 267-271
19. Moghadasian Mohammed H. (2002). DVM, MSc, PhD Mini review Experimental atherosclerosis A historical overview, *Life Sciences* 70 855– 865.
20. Sepideh Nourian , Ali Mohammadi Sani , Ebrahim Golmakani , Peyman Feizi , Katayoun Roghani (2016) ; Determination Antioxidant activity by High Performance Liquid Chromatography, Phenolic and Flavonoid contents of *Vincetoxicum nigrum* ; Vol.9, No.3, pp 150-157
21. Dalle-Done, I.; Rossi, R.; Colombo, R.; Giustarini, D. and Milzani, A. (2006). *Clinical Chemistry*, 52: 601- 623.
22. Nasdiwaty Daud , Rosidah , M Pandapotan Nasution (2016); Antidiabetic Activity of *Ipomoea batatas* L. Leaves Extract In Streptozotocin-Induced Diabetic Mice .International Journal of PharmTech Research ;Vol.9, No.3, pp 167-170.
23. Zainab Sajid , Abdul AL-Hadi Salil , Haider Salih (2016) ; Effects of prazosin in body weight and some hormones (TSH, T3, & T4) in rats (*Rattus norvegicus*); International Journal of PharmTech Research ; Vol.8, No.10, pp 66-71.
24. Ahmed Atia , Nadia Alrawaiq and Azman Abdullah (2016) ; Food Consumption and Body Weight in Mice Treated with Palm Oil–Derived Tocotrienol Rich Fraction (TRF) . International Journal of PharmTech Research; Vol.9, No.3, pp 262-266 .
25. Rubila. S and Ranganathan T.V. (2016) ; Effect of *Allium sativum* paste against Antimicrobial, Antioxidant and Cytotoxicity activity . International Journal of PharmTech Research; Vol.9, No.3, pp 328-332.
26. Musa Nima Mezher (2016); A Comparative Study between HBV Viral DNA Detection and Conventional Serological Methods of Diagnosis. *International Journal of PharmTech Research* ; Vol.9, No.4, pp 303-306.

27. Aprilita Rina Yanti , Maksum Radji , Abdul Mun'im , and FD Suyatna (2016) ; Antioxidant effects of Methanolic extract of *Phaleria macrocarpa* (Scheff.) Boerl in fructose 10%-induced rats. International Journal of PharmTech Research ; Vol.8, No.9, pp 41-47 .
28. Ernawati, Tri Bintarti . (2015); Acute Toxicity Study Of Ethanolic Extract Of *Solanum sanitwongsei* Craib Fruits on Mice. International Journal of PharmTech Research ; Vol.8, No.4, pp 642-647 .
29. Preethi.J and Saranya.V.T.K. (2015); Phytochemical Analysis on Leaf Extract of *Celosia argentea* Land its Efficacy of Antioxidant and Anti Bacterial Activity. International Journal of PharmTech Research ; Vol.8, No.4, pp 709-712 .
