



Antimicrobial Potential of Hemolymph of A Fresh Water Crab *Oziotelphusa Senex Senex* ((Fabricius 1798))

*¹D.Sumalatha, ²Jayanthi J and ³Ragunathan MG

¹Department of Biotechnology, Valliammal College For Women

²G.S.Gill Research Institute, Guru Nanak College, Chennai-India

³Department of Advanced Zoology and Biotechnology, Guru Nanak College, Chennai-India

Abstract: Crustaceans, have an immense immunological defence against pathogenic microorganisms. In the present study, effort has been made to find the antimicrobial activity of haemolymph collected from a freshwater crab *Oziotelphusa senex senex*. The hemolymph collected was subjected to antimicrobial assay by well diffusion method against clinical pathogens. Six bacterial species namely *Escherichia coli*, *Klebsiella pneumonia*, *Streptococcus pyrogenes* *Pseudomonas aeruginosa*, *Staphylococcus aureus* and *Bacillus sp*. The result shows a strong response of haemolymph against the clinical pathogens which confirms the immune mechanism of the freshwater crab.

Keywords: *O senex senex*, AMP, Hemolymph, antibacterial activity, pathogens.

Introduction

The emergence of new infectious diseases and resistance to antibiotics by the existing ones led to the development of new drug discovery(1).Antimicrobial peptides are important in the first line of the host defense system of many animal species. Their value in innate immunity lies in their ability to function without either high specificity or memory. Moreover their small size makes easy to synthesize without dedicated cells or tissues and they rapidly diffuse to point of infection(2). Recent experimental data from invertebrates suggests the past exposure to pathogens in individual animals can lead to enhanced immunity and some are also known to have considerable specificity by recognizing non self pathogen associated receptors that are highly conserved in evolution(3). Some of the known innate responses in invertebrates include phagocytosis, nodulation and encapsulation, synthesis of AMP and activation of proteolytic cascades that lead to melanization, blood coagulation, release of stress responsive proteins and molecules believed to function in opsonization and iron sequestration(4). The crabs are in intimate contact with aquatic environment rich in pathogenic microbes and are prone to infection by those microbes at various stages of growth, and losses due to disease can be enormous (5). An. Over the past several years, many antimicrobial peptides have been found and characterized in crab species. The first antimicrobial peptide characterised was a proline peptide of 6.5KDa from the hemocytes of the shore crab *Carinus maenas* (6). The antimicrobial peptide Callinecin is a cationic antimicrobial peptide of 3.7 KDa isolated from the blue crab, *Callinectes sapidus*. (7).Recently, scygonadin, an anionic antimicrobial peptide isolated from seminal plasma of the mud crab *Scylla serrata*(8).

Antimicrobial activity has been detected in several decapod crustaceans, including lobsters, crabs, shrimps and freshwater crayfish (9,10). Fresh water crabs are an important of the fauna of limnic environments (11).About 1300 species of fresh water crabs are distributed throughout the tropics and subtropics(12).Some of the Brachyuran crabs have shown pronounced activities and may be useful in the Biomedical area .The

potential of Freshwater crabs as a source of biologically active products is largely unexplored. The circulating hemolymph in crustaceans contain biologically active substances such as complement, lectins, clotting factors and antimicrobial peptides [13]. Hence a broad screening of Freshwater crabs for bioactive compounds is necessary. The present study was aimed to analyze antibacterial activity of hemolymph from *Ozotetelphusa senex senex*.

Sample collection

Freshwater Crab (*Ozotetelphusa senex senex*) was collected from the paddy field in Kundratur, Thiruvallur district, Tamilnadu, India. Healthy Male and female crabs of uniform size and free from disease were used for experimental purpose and each crab was subjected to single bleed collection. The weight of the collected crabs was ranging between 50-100g. The crabs were acclimatized for a week in the laboratory.

Collection of hemolymph

Haemolymph of *O. senex senex* was collected aseptically from the base of one of the second walking legs using a sterile syringe. To avoid haemocyte degranulation and coagulation, the hemolymph was collected along with ice-cold citrate EDTA buffer (510m M NaCl ; 0.1M glucose; 30mM trisodium citrate; 20mM citric acid; 10mM EDTA, pH 4.6)(14) as anticoagulant. Haemolymph was centrifuged at 2000rpm for 15min at 4°C. Supernatant was collected by aspiration and stored at 4°C until use.

Protein Estimation Assay

The protein content of crude haemolymph samples were estimated by Lowry's method using Bovine serum albumin as standard [15] using UV spectrophotometer.

Microbial strains used

Antibacterial activity of crab was determined against 6 different bacterial strains viz, *Klebsiella pneumonia*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Streptococcus pyogenes*, *Bacillus subtilis*, *Staphylococcus aureus*

Preparation of inoculum

Strains of 6 bacterial cultures were used in this study. Identical colonies were isolated and sub cultured in 2 ml of nutrient broth for 4 hrs at 37°C.

Antimicrobial activity

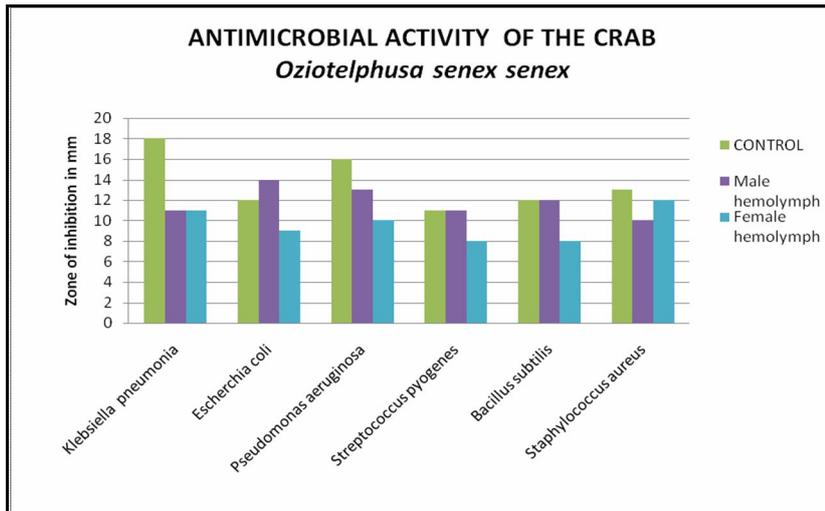
In vitro antibacterial assay was carried out by using standard well diffusion method (16,17]. 100 µL of diluted bacterial suspension (5×10^6 cfu /ml) of test bacterial strains were swabbed on the surface of Hinton agar. After 5 min, 100µl of hemolymph was added and the plates were incubated at 37°C for 24 hrs. The diameters of the zone of inhibition of growth was measured with the help of a scale. More than 12 mm in the measurement is sensitive zones, between 4 to 12 mm is moderately sensitive and zones less than 4 mm are resistant.

Results

Antimicrobial Assay

Antibacterial Assay

Antibacterial activity of the male and female hemolymph sample of *O. senex senex* was used for the present study. The zone of inhibition in different bacterial strains against *O. senex senex* hemolymph is shown in (Fig 1). In antibacterial activity the highest zone of inhibition was observed in the hemolymph of male crab, *O. senex senex* against *E. coli* (14 ± 1 mm) and the minimum activity was observed against *Staphylococcus aureus* (10 ± 1 mm). In the hemolymph of female crab, *O. senex senex* the highest zone of inhibition was observed against *Klebsiella pneumonia* (11 ± 1 mm) and the minimum activity observed against *Streptococcus pyogenes* and *B acillus subtilis* (8 ± 1 mm). The antibacterial agent of tetracycline showed activity against all the bacterial strains tested. The highest zone of inhibition was observed against *Klebsiella pneumonia* (18 ± 1 mm) mm) the minimum activity observed against *Streptococcus pyogenes* (11 ± 1 mm).



S.NO	ORGANISM	Zone of inhibition in mm		
		Control	Male Haemolymph	Female Hemolymph
1	<i>Klebsiella pneumonia</i>	18	11	11
2	<i>Escherichia. Coli</i>	12	14	9
3	<i>Pseudomonas aeruginosa</i>	16	13	10
4	<i>Streptococcus pyogenes</i>	11	11	8
5	<i>Bacillus subtilis</i>	12	12	8
6	<i>Staphylococcus aureus</i>	13	10	12

Discussion

In recent years, great attention has been paid to study the bioactivity of natural products due to their potential pharmacological utilization. The present research investigation is made on the basis of in search of antimicrobial peptides from the hemolymph of *O. senex senex*, a fresh water crab collected from the rice field environment. In the present study the crab hemolymph shows antimicrobial activity against different range of bacterial strains of both gram positive and gram negative bacteria. In decapods crustaceans, it is known that environmental changes may affect the immune ability to susceptibility against pathogen infection. Previous works show that decapod crustaceans contain factors with antibacterial activity, particularly in the hemolymph or in the hemocytes. Antibacterial activity was reported in different body-parts of *Pagurus bernhardus* (Hermit crab), *Pandalus borealis* (Northern shrimp), *Hyas araneus* (Spider crab) and *Paralithodes camtschatica* (King crab) [18]. A similar result was observed with the hemolymph of some brachyuran crabs against clinical pathogens [19]. Antibacterial activity has been previously described in a wide range of crustacean species. Antimicrobial peptides have been established as key players in animal defense systems. An antimicrobial peptide, which is isolated from a decapod crustacean (a crab, named *Thalamita crenata*), possess an immense antibacterial activity (20). One of such antimicrobial protein from the crab hemolymph (*Charybdis lucifera*) has been extensively studied against *E. coli* and *P. aeruginosa* (21). The influence of crab hemolymph against wide range of clinical pathogens proves that crustaceans are very good source of antimicrobial potency (22). Antibacterial peptides can also be induced in epidermal cells in response to wounding or infection in the cuticles [23]. The whole process of synthesizing antibacterial proteins may take few minutes or hours after the changes, and these are secreted into the haemolymph of which some are lysozyme [24] and andropin [25]. These proteins show strong resistance to the microbial Growth.. The present study indicates that haemolymph of *O. senex senex* may contain potential antibiotics. The antimicrobial assay done so far will serve as a baseline data for further studies that may confirm the hypothesis that brachyuran crabs haemolymph are indeed potential sources of novel compounds with biological potential. The revealing and development of the antimicrobial compounds in the haemolymph will provide an opportunity for the production of new compounds with natural activities as alternatives to antibiotics. Further purification of the active compounds is necessary in order to identify their chemical nature and to evaluate their potency as novel drug.

Conclusion

The biological significance of the presence of the AMP in the crab hemolymph is still unclear. Considerable effort is being put into investigating the therapeutic potential of these peptides. The present study indicates that the haemolymph of crab would be a good source of antimicrobial agents and would replace, the existing inadequate and cost effective antibiotics. Following these in our present study, the crab haemolymph showed strong activity against the growth of selected microbes. The result suggests that the crab can produce antimicrobial substances instantly to combat microbial infection.

References

1. G Rameshkumar, S Ravichandran, G Kaliyavarathan, TT Ajithkumar Antimicrobial peptide from the crab, *Thalassidroma crenata* (Latreille, 1829) World journal of Fish and Marine sciences 1(2):74-79,(2009).
2. Bonman.H.G. Peptide antibiotics and their role in innate immunity Annual Review Immunology,13:61-92(1995)
3. Janeway.Jr CA, Medzhitov R Innate Immune Recognition Annual Review Immunology,20:197-216(2002)
4. Lee SY,Soderhall K .Early events in crustaceans innate immunity. Fish Shellfish Immunology 12:421-437. (2002)
5. D. A. Hudson and R. J. G. Lester, "Parasites and symbionts of wild mud crabs, *Scylla serrata* (Forsk.) of potential significance in aquaculture," *Aquaculture*, vol. 120, no. 3-4, pp. 183–199, (1994).
6. Ai HS,Liao JX ,HuangX D,Yin ZX, WengSP, Zhao ZY . A novel phenoloxidase 2 exists in shrimp hemocytes.Dev Comp Immunology:33: 59-68. (2009)
7. Ma Z,Zhao J, Wu ShiC, Zhang C.The quantitative research of composite immune indicator for crustaceans. Fish Shellfish Immunology 28 :187-192.
8. Spersad SV,Haug T, Vasskogt, stensvag K,Hyastatin a glycine –rich multi-domain antimicrobial peptide isolated from the spider crab, *Hyas araneus* hemocytes.mol Immunol: 46:2604-2612.(2009)
9. Stewart, J.E. and B.M. Zwicker,. Natural and induced bactericidal activities in the hemolymph of the lobster; *Homarus americanus*: products of hemocyte-plasma interactions. Canadian J. Microbiol., 18: 1499-1509.9(1972)
10. Noga, E.J., Arroll, T.A. Fan and Zhigin, .Specificity and some physiochemical characteristics of the antibacterial activity from the blue crabs *Callinectes sapidus*. Fish-Shell fish-Immunology, 6(6): 403-12(1996)
11. Wehrmann,I.S Celio Magalhaes;Patricio Hernaez and Mantelatto ,F.L. .Maga offspring production in three freshwater crab species (Brachyura Pseudothelphusidae) from the Amazon region and central America.Zoologia 27 (6) 965-972.(2010)
12. Yeo,D.C.J., Ng.P.K.L., Neil Cumberlidge,Celio Magalhaes, Daniels.S.R and Campos,M.,Global diversity of crabs (Crustacea : Decapoda Brachyura) in fresh water.In E.V.Balian C.Levveque,H.Segers and K.Martens."Freshwater animal Diversity Assessment. Hydrobiologia, developments in hydrobiology, 198-595(1) 275-286.(2008)
13. M. I. Hoq, M. U. Seraj, and S. Chowdhury, "Isolation and characterization of antimicrobial peptides from the mud crab, *Scylla serrata*," *Pakistan Journal of Biological Sciences*, vol. 6, pp.1345–1353, (2003).
14. Soderhall .K.I. and Smith V.J. Separation of the hemocyte populations of *Carcinus maenas* and other marine decapods and phenoloxidase distribution.Devel.comp.Immunol.7.229-239. (1983)
15. Lowry, O.H., J.N. Rosenbrough, A.L. Farr and R.J. Randall .Protein measurement with folin-phenol reagent. J. Biol. Chem.vol 193, pp 265–75.(1951).
16. Mayer AM, Rodriguez AD, Berlinck RG, Fusetani N. Marine compounds with antibacterial, anticoagulant, antifungal, anti-inflammatory, antimalarial, antiprotozoal, antituberculosis, and antiviral activities; affecting the immune and nervous system, and other miscellaneous mechanisms of action. Comp Biochem Physiol;153C:191-222. (2011).
17. Mitta G, Hubert F, Noel T, Roch P. Myticin, a novel cysteine-rich antimicrobial peptide isolated from hemocytes and plasma of the mussel, *Mytilus galloprovincialis*. Eur J Biochem;265:71–8 (1999).
18. Haug, T., A.K. Kjuul, E. Sandsdalen and O.B. Styrvold,. Antibacterial activity in four marine crustacean decapods. Fish Shellfish Immunol. May;12(5):371-85.(2002).
19. Bauer, A.W., W.M.M. Kirby, M.J.C. Sherris and M. Turck. .Antibiotic susceptibility testing by a standardized single disc method.American J.of Clinical Pathol. vol 45, pp 493-496.(1996).

20. Noga, E.J., T.A. Arroll Fan and Zhigin, Specificity and some physiochemical characteristics of the antibacterial activity from the blue crab *Callinectes sapidus*. Fish&Shellfish- Immunology,, Volume 6, Issue 6, August 1996, Pages 403–412(1996).
21. Rameshkumar G, Ravichandran S, Kaliyavarathan G, and Ajithkumar TT Antimicrobial peptide from the crab, *Thalamita crenata* (Latreille, 1829). Wor. J. Fish Mar. Sci., 1(2): 74-79. (2009).
22. Anbuchezhian RM, Ravichandran S, Rameshkumar G, and Ajithkumar TT. Influence of crab haemolymph on Clinical Pathogens. Adv. Biol. Res., 3(3-4): 104-109. (2009).
23. M. I. Hoq, M. U. Seraj, and S. Chowdhury, "Isolation and characterization of antimicrobial peptides form the mud crab, *Scylla serrata*," *Pakistan Journal of Biological Sciences*, vol. 6, pp. 1345–1353, (2003).
24. W. J. Lee and P. T. Brey, "Isolation and characterization of the lysozyme-encoding gene from the silkworm *Bombyx mori*," *Gene*, vol. 161, no. 2, pp. 199–203, (1995).
25. C. P. Samakovilis, D. A. Kylsten, A. Kimbrell, A. Engstrom, and D. Hultmark, "The adropoin gene and its product, a male specific antibacterial peptide in *Drosophilla melanogaster*," *The EMBO Journal*, vol. 10, pp. 163–169, (1991).
26. D. Saravanan, M. Radhakrishnan(2016), Antimicrobial activity of mangrove leaves against drug resistant pathogens, International Journal of PharmTech Research, (2016), Vol.9, No.1, pp 141-146.
27. Benjamin E. Ezema, Emmanuel I. Odoemelam, Mathias O Agbo(2016), Phytochemical and Antibiotic Evaluation of the Methanol Extract of *Loranthus micranthus* Linn Parasitic on *Kola Accuminata*, International Journal of PharmTech Research, (2016), Vol.9, No.2, pp 176-181.
28. Rubila. S, Ranganathan T.V.(2016), Effect of *Allium sativum* paste against Antimicrobial, Antioxidant and Cytotoxicity activity, International Journal of PharmTech Research, (2016), Vol.9, No.3, pp 328-332.
29. Mona Al-Terehi, Ali H. Al-Saadi, HaiderK.Zaidan, Russul Hikmat Behjet, Zahraa Haleem(2015), Some plants extracts Synergism effects in Pathogenic bacteria, International Journal of PharmTech Research, (2015), Vol.8, No.10, pp 158-164.
30. Helmina Br. Sembiring, Tonel Barus, Lamek Marpaung, and Partomuan Simanjuntak(2015) , Antioxidant and Antibacterial Activity of Some Leaves Extracts (Methanol, Ethyl Acetate and N-Hexane) of *Scurrula fusca* G.Don, International Journal of PharmTech Research, 2016, Vol.8, No.9, pp 24-30.
31. K. Kayalvizhi, L. Cathrine, K. Sahira Banu(2015), Phytochemical and antibacterial studies on the leaf extracts of female *Carica papaya*.linn, International Journal of PharmTech Research, (2015), Vol.8, No.7, pp 166-170.
32. R. Selvaraj, S. Jansi Rani, N. Mahesh Kumar and A. Natarajan; Antimicrobial screening and phytochemical analysis of *Aegle marmelos* against enteric pathogens; International Journal of PharmTech Research, 2015, Vol.8, No.2, pp 244-249.
33. T. G. Nithya, Isah Mansur Aminu; Antibacterial activity of *Murraya koeniigi* leaves against Urinary Tract Infection causative pathogens; International Journal of PharmTech Research; 2015, Vol.8, No.8, pp 112-117.
34. Helmina Br. Sembiring, Tonel Barus, Lamek Marpaung, and Partomuan Simanjuntak; Antioxidant and Antibacterial Activity of Some Leaves Extracts (Methanol, Ethyl Acetate and N-Hexane) of *Scurrula fusca* G.Don; International Journal of PharmTech Research; 2015, Vol.8, No.9, pp 24-30.
35. N. Anupama, G. Madhumitha; Pharmacognostic Standardization and Anti-microbial studies of dried *Carissa carandas* fruits International Journal of PharmTech Research; 2015, Vol.8, No.8, pp 206-210.
36. Ayyappadasan Ganesan, MadhuPreetha Thangapandian, Ponmurugan Ponnusamy, Jeya Prakash Sundararaj, Sanjeeva Nayaka; Antioxidant and antibacterial activity of parmelioid lichens from Shevaroy hills of Eastern Ghats, India; International Journal of PharmTech Research; 2015 Vol.8, No.9, pp 13-23.
37. Mansureh Kamali, Susan Khosroyar, Ameneh Mohammadi; Antibacterial activity of various extracts from *Dracocephalum kotschy* against food pathogenic microorganisms; International Journal of PharmTech Research; 2015, Vol.8, No.9, pp 158-163.
38. Deepali Mor, Sahas Bansal, M. Ramachandran, Pramod Raichurkar; Review on Antibacterial, Antiviral, and Antifungal Properties of Natural Diapers and its Effect on Dermatitis; International Journal of PharmTech Research; 2015, Vol.8, No.10, pp 40-46.
