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Conventional and molecular approaches in bacterial contamination detection for meat samples

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Abstract: Since meat and its derivatives are the main sources of human deals, it must be free of contamination and hazard .This study was conducted to investigate the probability presents of bacterial contamination of several types of imported meat. All type of meat specimens show a rate of bacterial contamination. Conventional culture methods reveals that *Salmonella spp*. form the higher rate of isolated bacteria followed by Staphylococcus and Bacillus , while *Stapylococcus and Pseudomonas* form the predominant detected isolates by molecular assay using PCR techniques . Susceptibility of isolated bacteria to antibiotics reveals that Impenem and Nalidic acid are the more effective antibiotics against all types of bacteria . Detection of MIC against *Salmonella* isolates reveals also that Impenem is the most is the most effective with low concentration reach to 1.4 ug / ml.

Key words : Meat, Bacterial contamination, molecular assay, MIC. Impenem.

Introduction

Meatand is an essential source of good quality protein that provide us with the major amino acids for our daily requirement. Microbial source tracking (MST) methods allow the identification of the types of microbial contaminants, the extent of contamination, and the possible source of contamination (1).Bacteria have accounted for more than 70% of deaths associated with foodborne transmission (,2).Microbial contamination include broad range of foods not only meat, Coliform group and *Escherichia coli* as fecal indicator contamination were implicated in 18 and 7% of Domiati cheese samples, and were not found in any Feta cheese and sterilized milk samples. *Staphylococcus aureus*, aerobic spore formers, yeasts and molds were detected in 4.5%, 40%, 22.5% of Domiati cheese samples respectively (3).

Molecular approaches are useful tools for the detection of fungi or bacteria even of their low quantity depending on nucleic acid sequence in addition to rapid molecular detection (4,5). Bacteria are one of the most agents for food poising .Occurrence of bacteria on food don't effect the taste or texture of food but it cause a big health problem through the causes of several disease according to bacterial types(6).

A number of studies have pointed outbreak of infections due to consumption of food contaminated and poor hygiene, however, few of these reports provide evidence of several outbreak caused by *Salmonella*, *Shigella*, *E. coli* and *Listeria* spps in different parts of the world (7). Many types of bacteria produced substances that had an effective virulanicity value. Exopoly saccharides (EPS) can be extracted from different type of bacteria and their content and antibacterial value varies with bacterial types, it was found that extract was higher in *Bacillus subtilis* in comparison with Pseudomonas aueroginosa(8).

Meat contamination occurs through poor handling and storage practices. Many types of bacteria found on meat some of them are pathogenic and form a major cause of food poising and other diseases such as Campylobacter ,Salmonella, E.coli , Bacillus and other types (4).

Infectious bacteria produce miserable toxins and enzymes like *Escherichia coli* O157:H7. It is azoonotic foodborne pathogen of major importance that produced Shiga toxin. The diseases associated with *E. coli* O157:H7 infection are hemorrhagic colitis (HC) and hemolytic uremic syndrome (9).

The *Clostridium* genus form other bacteria types that cause meat and food contamination, *Clostridium perfringens* is an important pathogen of human gastrointestinal tract diseases such as food poisoning, antibiotic-associated diarrhea, and sporadic diarrhea as well as nosocomial diarrheal disease outbreaks and can cause morbidity if contamination of raw meat and poultry occurs, if these products are properly handled and prepared, particularly in restaurants and catering facilities.(10).

Food-producing animals, including cattle, chickens and turkeys are recognised as reservoirs of enteric bacteria, such as *Salmonella* spp., *Campylobacter* spp., *Escherichia coli*, *Clostridium perfringens*, *Listeria* spp., and *Yersinia* spp. (2,11).

Microbial studies on imported meat available in Iraqi markets were limited, so the present study was carried out to searching for the presence of bacterial contaminants on different type of imported meat using conventional and molecular methods.

Materials and Methods

Samples collection

Fifty specimens of different types of meat (AlMurad, Al Kaffel ,Al Huda ,Al Hasnawi, Jeckor, fresh sheep meat and fresh beef meat) were taken in this study. Twosmall pieces of each specimens were added to separated plain tube contain brain heart infusion broth in addition to swab samples that taken from meat surface then brought to laboratory .

All samples were incubated at 37c for 24-48 hours, swabs of each samples were cultured on different types of solid media including nutrient agar, MaConkey agar, EMB agar, Manitol salt agar and blood agar. Colonies were identified morphologically and biochemically(12).

Antibiotic susceptibility test

Different types of antibiotic disk were used to investigate the sensitivity and resistance of isolated bacteria to Cephalixin, Impenem, Ampicilin, Nadlxic acid and Cefotaxim using Kirrby and Bauer disk diffusion method, the CLSI, (13) were dependent in results interpretation.

Molecular assay

Small pieces of each meat specimens type were cultured for 24 hrs. at 37c in brain Swabs of selected drugs were used for bacterial DNA extraction using promega DNA EXTRACTION KIT, the concentration of DNA were estimated by nanodropspectronic.

DNA for each samples were amplification by polymerase chain reaction (PCR) using set of specific primers listed I table 1, the DNA amplification was done by using mono and multiplex amplification process with a final product of 20 and 25 ul respectively. Agarose gel electrophoreses for the products were carried out and the bands of bacterial genes were detected by E-graph gel documentation(14).

Reference	Size	Sequence (5-3)	Primer	Bacteria
15	370bp	GGC CGT GTT GAA CGT GGT CAA ATC A	TstaG422	Staphylococcus
	_	TIA CCA TTT CAG TAC CTT CTG GTA A	Tstag765	spp.
16	884bp	CCGATACGCTGCCAATCAGT	Ec1	Escherechia
	_	ACGCAGACCGTAAGGGCCAGAT	Ec2	coli
17	480bp	TATCCTCTCTATATGCACAG	LT3	
	_	CTGTAGTGGAAGCTGTTATA	LT4	
18	100bp	GTG AAA TTA TCG CCA CGT TCG GGC AA	InvA	Salmonella
		TCATCG CAC CGT CAAAGG AAC C		spp.

Table 1 : Primers used in bacterial diagnosis

Results

Cultural investigations on all samples of imported meat show occurrence of bacterial contamination including different types of bacteria .*Salmonella Spp.* (11 isolates) Form the most predominant followed by *Staphylococcus and Bacillus spp.* (10 isolates , while *,proteus spp.* represent the lower isolates (table,2)

%	No. of				Bacterial types			
	Isolates	Mumtaz	Jekoor	Al- Hasnawi	Al- Kafeal	Al- Murad	Al-Huda	
20.83%	10	1	2	2	2	2	1	Staphylococcus spp.
6.25%	3		1		1		1	Proteus spp.
20.83%	10	2	3	1	1	2	1	Bacillus spp.
16.66%	8	1	1	2	1	1	2	Pseudomonas spp
22.91%	11	1	3	2	1	2	2	Salmonella spp.
12.5%	6	2	1	-	-	1	2	Escherechia coli
	48	7	11	7	6	8	9	Total

Table 2. Distribution of bacterial isolates on meat specimens

Susceptibility of bacteria isolated from different meat sources to five types of antibiotics reveals that resistance or sensitivity of isolates were differs with the differences of bacteria and antibiotics types. Impenem represented the most effected antibiotics against the tested bacteria. Furthermore, *Escherichia coli and Proteus spp.* Show more sensitivity to most antibiotics (table 3).

Cefotaxime 5ug	Nadlixic acid 30 ug	Ampicillin 30 ug	Impenem 10 ug	Cephalexin 30ug	Type of Bacteria
Ι	Ι	R	S	Ι	Staphylococcus spp.
S	S	R	S	R	Proteus spp.
R	Ι	R	Ι	R	Bacillus spp.
R	S	R	S	Ι	Pseudomonas spp
Ι	Ι	R	S	S	Salmonella spp.
S	S	R	S	Ι	Escherechia coli

The eleven isolates of Salmonella, the predominant bacterial isolates, were selected for detection of the minimum inhibitory concentration (MIC) towards three types of antibiotics. Results confirmed that the lowest concentrations of MIC appeared with impenem when compared with other types of antibiotics (table 4).

AMP(≥32ug/ml)	CTX(≥	IMP(≥4ug/ml	Salmonella	
	64ug/ml)		isolates No.	
3.6	3.4	1	Isolate 1	
8.4	2.2	0.5	1 solate 2	
5.2	6.4	1.2	Isolate 3	
4.6	4.8	2.6	Isolate 4	
10.6	3.2	1.2	Isolate 5	
6.8	1.4	0.5	Isolate 6	
8.2	6.4	4.3	Isolate7	
12.8	8.2	1.6	Isolate8	
4.4	0.8	0.4	Isolate9	
12.8	3.8	0.8	Isolate10	
8.6	2.6	2.2	Isolate11	
		1.4 ±	Mean $\pm SD$	

Table 4. Minimum inhibitory concentration (MIC) of *Salmonella* isolates to β Lactam antibiotics.

Molecular tools were used for further detection of bacterial contamination in meat specimens .Monoplex and multiplex polymerase chain reaction assay proved appearance of different type of bacteria with variation in numbers of isolates. Staphylococcus spp. and Salmonella spp. form the predominant bacterial occurrence in all type of meat that reach to 18 isolate for each, whereas, Proteus spp. appearance was rare (table 5, figure 1 and 2).

Table 5. genetic diagnosis of bacterial contamination of different type of meat

	No of contaminated specimens	No. of specimens	Types of meat					
Escherechia coli	Salmonella	Pseudomonas	Bacillus	Proteus	Staphylococcus			
	spp.	spp	spp.	spp.	spp.			
2	1	4	1	-	4	12	16	Al-Huda
3	2	4	2	1	3	14	20	Al-Murad
4	1	3	3	-	4	15	20	Al-Kafeal
3	2	3	2	-	2	10	14	Al-Hasnawi
2	2	2	2	-	2	9	10	Jekoor
2	1	2	2	-	3	9	10	Mumtaz
16	9	18	12	1	18	69	90	total

Detection of bacterial contamination of meat specimens by molecular assay give areal proof for meat contamination, using of universal bacterial primers pointed out of various bands represent the appearance of different bacterial isolates (Figure 1, 2).

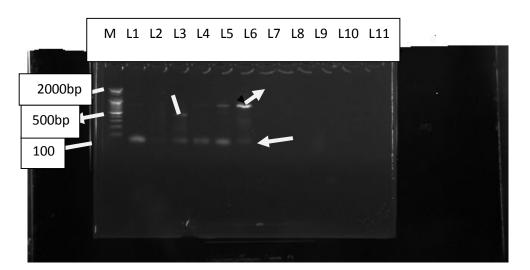


Figure 1.Gel electrophoresis of extracted DNA using multiplex PCR assay, different bands refer to different type of bacterial contaminants (100,370 and 884bp).

Discussion

All type of human uptake foods must avoided of microbial contamination, meat is one of the most food that uptake worldwide because of their essential nutrientmaterials. Contamination of food with Pathogenic microorganisms cause illness, toxic and complications that leads some time to death (19).

Results of the present study pointed that all types of studied meats were contaminating with one or more bacterial type, although some of them are not harmful. The occurrence of bacteria on meat due to their compositions that make it as a good media for growth and multiplying of all type of microorganisms, meat is not only sensitive to microbes but it involved in disease spreading (20).

Different type of bacteria were isolated and identified in this study, *Staphylococcus and Salmonella* form the predominant isolated bacteria. The rate of isolated bacteria from meat differs with the type of meat, place and storage method. In many countries, meat regarded as a great source of pathogenic bacteria that cause many diseases. Many types of bacteria, some of them are pathogenic such as *E. coli, Campylobacter and Salmonella, proved* as an contaminant agent for meat (21).

The present study revealed that some of bacterial isolated from imported meat are pathogenic for human such as most enteric bacteria and *Stapylococcus*. Ali *etal*, (22) found that 84% of their meat samples were found to be contaminated with bacterial species, including *Klebsiella, Enterobacter, Staphylococcus aureus and Bacillus subtilis*. Contamination of food by microbes can be inhibited by other microorganisms, Contamination by mould in bread can be inhibited by several microorganism, Lactic Acid Bacteria (LAB) has no degrading effect over the nature, taste and texture of bread, makes it suitable for the bio preservation of bread(23).

Isolated bacteria were tested for their sensitivity to set of antibiotics, results expressed variation in susceptibility of isolates to different antibiotics. Variation in bacterial susceptibility correlated with type of

Salmonella isolates were chose for detection of MIC against three antibiotics type, Impeneme expressed the lowest MIC for all isolates in compared with other types of antibiotics. Many types of antibiotics had a broad spectrum action to different types of microbes, although there is some specificity of antibiotics action according to their target sites on microbes (25). The antibiotic sensitivity test and MIC had a good value in detection of inhibitory effects, itwas observed that MIC Value of Azithromycin against Proteus vulgaris, *Enterococcus faecalis, and Enterobacter aerogenes* was 4µg/ml, 1.2µg/ml and 0.12µg/ml respectively(26).

Using of polymerase chain reaction assay in bacterial contaminant detection give a good tools in more accurate detection, the three gene primers used in this study reveals surly appearance *Salmonella*, *Staphylococcus and E.coli*. Several studies on microbial meat contamination pointed out variation in percentage of contaminants according to tolls that used in bacterial isolation and identification, most of these studies performed that PCR assay is the more accurate method in microbial detection (11, 22).

Conclusion:

Cultural and molecular assay reveals that Salmonella and Staphylococcus are the most bacterial contaminants for meat specimens, furthermore ,the susceptibility investigation showed that Impenem an Nalidic acid antibiotics are the more effective against these types of bacteria .

References

- 1. Montiel-Sosa, J.F; Ruiz-Pesini, E.,Montoya, J..Rocales, P., Lopez-Perez, M.J. and Perez-Martos, A. (2000). Direct and highly species-specific detection of pork meat and fat in meat product by PCR amplification of mitochondrial DNA. J.Agric..Food Chem.48:2829-2832.
- 2. Hughes C., Gillespie I.A., O'Brien S.J. and The Breakdowns in Food Safety Group,(2007).Foodborne transmission of infectious intestinal disease in England and Wales, 1992-2003 Food Control. 18, 766-772.
- 3. Sharif ,O.M.S; Ibrahim, G.A.;Tawfek, N.F.;Effat, B.A.;ElShafei, K.;El Din,H.M and Salem M.M.A. (2014). Prevalence of some pathogenic microorganisms in factories Domiati, Feta cheeses and UHT milk in relation to public health sold under market conditions in Cairo. Int.J.Chem.Tech.Res..6(5): 2807-2814.
- 4. Pillai, S. D., and E. Vega. (2007). Molecular detection and characterization tools, p. 65–91. In J. W. Santo Domingo and M. J. Sadowsky (ed.), Microbial source tracking. ASM Press, Washington, DC.
- 5. Hathout, A S., Abo-Sereih, N.A.Sabri, B.A.; Sabah, A.F. and Aly, S. (2015). Molecular identification and control of some pathogenic Fusarium species isolated from maize in Egypt. Int. J.Chem.Tech.Res. 7(1):44-54.
- 6. Brooks G.F, J.S. Butel and S.A. Morse. (2007). Jawetz. Melnick and Adelberg's Medical Microbiology. 24th ed, McGraw-Hill.
- 7. 7.Zweifer C, Fischer R, Stephan R (2008). Microbiological contamination of pig and cattle carcasses in different small-scale Swiss abattoir, Meat. Sci., 78: 225-231.8.Anima, N.;and Raghavan, C.M. (2014).
- 8. Production and characterization of exopolysacharides (EPS) from the bacteria isolated from Pharma lab sinks. Int.J. PharmTech Res.6(4):1301-1305.
- 9. Cray, W. C. J. and Moon H. W. (1995). Experimental infection of calves and adult cattle with *E. coli* O157:H7. Appl. Environ. Microbiol. 61: 1568-1590.
- 10. Grass, J.E., Gould, L.H. and Mahon, B.E. (2013). Epidemiology of foodborne disease outbreaks caused by *Clostridium perfringens*, United States, 1998-2010. Foodborne 11. Vasut, R.G., Rubeci, M.D. (2009).
- 11. Food contamination with psychrophilic bacteria. Lubrari Scientific MedicinaVeterinara XLII(2) :325-329.
- 12. MacFaddin, J.F. (2000). Biochemical test for identification of medical bacteria. 3 thed. Williams and Wilkins- Baltimor. USA.
- 13. Clinical and Laboratory Standards Institute (CLSI). (2012). Performance Standards for Antimicrobial Susceptibility Testing; 22ed. Informational Supplement. 32(3).PA, USA.

- 14. Bartlett, J. S. and Stirling D.(1998). PCR Protocols: Methods in Molecular and Biology. 2th. Humana Press Inc. Totowa. NJ.
- 15. Martineau, F., Picard, F. J., Ke, D., Paradis, S., Roy, P. H., Ouellette, M.and Bergeron, M. G. (2001).Development of a PCR assay for identification of staphylococci at genus and species levels. J. Clin. Microbiol. 39, 2541–2547.
- 16. Chen J, Griffiths N.W(1998). PCR differentiation of Escherichia coli from other gram-negative bacteria using primers derived from the nucleotide sequences flanking the gene encoding the universal stress protein. LettApplMicrobiol 27:369–376.
- 17. Leong J, VinalA.C.and Dallas W.S. (1985). Nucleotide sequence comparison between heat-labile toxin B-subunit cistrons from *Escherichia coli* of human and porcine origin. Infect Immun 48: 73–77.
- Rahn, K., S. A. De Grandis, R. C. Clarke, R. Curtiss and C. L,(1992). Amplification of an InvA gene sequence of *Salmonella typhimurium* by polymerase chain reaction as specific method of detection of Salmonella ., Mol. Cell ., Probes .6:271-279.
- 19. Fratamico PM, Bhunia AK, Smith JL (2005). Foodborne pathogens in Microbiology and Molecular Biology, Caister Academic press, Wymondham Norfolk, UK. Pp. 270-275.
- 20. Iroha, I.R.; Ugbo, E.C.; Ilang, D.C.; Oji, A.E and Ayogu, T.E (2001). Bacteria contamination of raw meat sold in Abakaliki, Ebonyi State Nigeria. J. Pub. Health and Epidemiol. 3(2):49-53.
- Kinsella K.J., Prendergast D.M., McCann M.S., Blair I.S., McDowell D.A.and Sheridan J.J (2008). The survival of *Salmonella enteric* serovars*typhimurium* DT 104 and total viable counts on beef surfaces at different relative humidities and temperatures. J. Appl. Microbiol. 106: 171 – 180.
- 22. Ali,N.A., Farooqui,A., KhanA, Khan,A.YandKazmi,S.U.(2010). Microbial contamination of raw meat and its environment in retail shops in Karachi, Pakistan J.Infect. Dev. Ctries. 4(6):382-388.
- 23. Sivasankaran, C.; Arockiaswamy, W.J.; Ramaunjam. P.; Chellamuthu, S.; Muruganantham, K. and Shanmuga m, L. (2014). Prevention of Bread Spoilage and to Enhance the Quality of Bread by using Lactic Acid Bacteria. Int. J Chem. Tech. Res. 6(9):4161-4165.
- 24. Chao,G.,Zhou,X,Jiao,X.,Qian,X.,Xu,L.,(2007).Prevalence and antimicrobial resistance of foodborne pathogens isolated from food products in China.FoodbornePathog.Dis 4(3):277-284.
- 25. OsailiTM, Al-Nabulsi AA, Shaker RR, Jaradat ZW, Taha M, Al-Kherasha M, Meherat M, HolleyR. (2014). Prevalence of *Salmonella serovars, Listeria monocytogenes, and Escherichia coli* O157:H7 in Mediterranean ready-to-eat meat products in Jordan. J FoodProt. ;77(1):106-111.
- 26. Pinky Kaur, NishantRai(2015). Bacteriological Analysis of Fresh Vegetables from Main Market of Dehradun. Int. J. PharmTech Res .8(3):415-425.
