



International Journal of PharmTech Research

CODEN (USA): IJPRIF, ISSN: 0974-4304 Vol.9, No.2, pp 98-105, 2016

Protective role of turmeric extract (*Curcuma longa*) in the lipid profile and activity of antioxidant in the male rats treated by lithium carbonate

Afyaa S. Nasir*, Haider S. Jaffat

Science Faculty, University of Kufa, Najaf, Iraq

Abstract: The antioxidant effect of turmeric (*Curcuma longa*) extract against side effects induced by lithium carbonate (Li2CO3) were studied. The experiment was carried out on fifty male rats distributed randomly into 5 groups of 10 animals in eachgroup. The (1) group was kept as a normal control was received normal saline, rats of group (2) and (3) were given only lithium carbonate only in a dose 4 and 8 mg/kg for induction of oxidative state on rats, While other groups (4) was received lithium carbonate at dose 4 mg/kg with turmeric extract at dose 1 g for 1 kg of diet and groups (5) was received lithium carbonate at dose 8 mg/kg with turmeric extract at dose 1 g for 1 kg of diet. Results showed that oral administration of turmeric extract in rats with oxidative state by Li2CO3 decrease the lipid profile parameters and increase antioxidant enzymes. Conclusively: treatment by turmeric extract was produce a protective effect against oxidative stress by Li2CO3 in the male rats.

Keywords: lithium, lipid profile, lipid peroxidation, glutathione peroxidase, superoxide dismutase, male rat.

Introduction

Lithium is used in medicine more than 50 years ¹ especially in manic depression. However, lithium is applied in many fields as lithium therapy^{2,3,4,}neurodegenerative disease ^{5,6}. The usefulness of lithium in cure of bipolar disorder was reported in many researches ⁷. On other side, many studies were recorded adverse effects of lithium in the human ^{8,9,10}. The ROS are substances synthesized in the body during the normal metabolic processes as a series of incomplete reduction of oxygen molecule ^{11,12}. The living organisms are protect themselves from oxidative stress by producing antioxidants represent molecules having the capability to scavenge ROS^{13,14}.

Lithium have a wide medical applications and major in the balance between anti and pro-oxidative processes. In addition, lithium also effect on lipid metabolism during changes thyroid hormones levels ¹⁵. The precise mechanism to explain how lithium change the lipid metabolism is unknown¹⁶.

However, little data about the changes in lipoprotein metabolism during lithium treatment. Some authors demonstrated the relationship between the lithium side effects and lipids metabolism. These studies were suggested the changes in plasma lipid and lipoprotein levels following lithium treatment. This is very essential in psychiatric patients when taken lithium for long period¹⁷.

Materials and methods

Lithium carbonate (Li2CO3) was purchased from the Norginecompany, U.K.

Turmeric rhizomes were purchased from the local market in AL-Najaf city.

Preparation of phenolic extract of turmeric (curcuminoids): The rhizomes were crushed to powder by using a blender, take a bout 100g of powdered were added to 500ml of 80% ethanol and put the mixture in soxhelt system during 24h. After that, resulting extracts were filtered using filter paper and concentrated to dryness in rotary evaporator in the room temperature.

Then, the recipient was transferred to a separating funnel, and 2 N (HCl) were added gradually to get pH 2, then, washed with 10 ml chloroform three times. The solution was separated into two levels, the down level contain the phenols (curcuminoids) were residue, weighted and kept in a refrigerated until using it ¹⁸.

Determination of lipid profile activity

Total cholesterol kit for quantitative determination of total cholesterol in serum was supplied by Biolabo SA, France¹⁹.

Serum HDL-Cholesterol level was measured by HDL-Cholesterol phosphotungstic acid (PTA) precipitant kit (Biolabo, France)¹⁹.

Very low density lipoprotein (VLDL) were measured by using the following formula: $VLDL = TG (mg/dl) / 5^{19}$.

Low density lipoprotein (LDL) were measured by using the next formula: LDL=TC(mmol/l)-VLDL(mmol/l)-HDL(mmol/l)¹⁹.

Triglycerides Kit was supplied by Biolabo, France, for measuring triglycerides in human serum ²⁰.

Determination of antioxidant enzymes

Measurment of SOD activity by ELISA Kit (Elabscience, U.S.A.) (www.elabscience.com).

The quantitativedetermination of GPX concentration in serum through the enzyme linked immunosorbant assay using ELISA kit (Elabscience, U.S.A.) (www.elabscience.com).

Measurment of MDA activity by ELISA Kit (Elabscience, U.S.A.) is an enzyme immunoassay (www.elabscience.com).

Experimental Design:

Fifty male albino rats strain (*Rattusn orvegicus*) weighting (225-250g) obtained from the animal house in the science faculty/Kufa university. The rats kept under observation for one week before starting the experiment for acclimatization. fed on standard diet and water *ad libitum*. Then animals were divided into five groups of six rats in each. The first group was fed on the basal diet, normal saline and served as control. The second and third groups were received lithium carbonate at doses 4, 8 mg/kg respectively. The fourth and fifth groups were administration lithium carbonate at dose 4, 8 mg/kg plus turmeric extract (curcuminoids) at dose 1 g/1 kg respectively for 6, 8 weeks.Half number of rats from each groupafter 6 weeks of experiment were anaesthetized by Ketamine and xylazine and blood samples have collected by heart puncture and put into serum tubes in the room temperature for several minutes and were centrifuged for 20 minutes at 3000 rpm. At the end of experiment (8 weeks) the remainder of rats also anaesthetized by the same method and the blood samples were saved.

Statistical Analysis: Data were expressed as mean±S.E. and Statistical Analysis was carried using computerized SPSS program version (21) with one way ANOVA²¹.

Results and Discussion:

The results in table (1) and (2) show significant increase in the total cholesterol, triglycerides, low density lipoprotein and very low density lipoprotein in the serum of rats administration of lithium carbonate for six, eight weeks. The present results demonstrated that lithium, even at therapeutic doses, disturbs lipid metabolism. This disturbance may be started by the changes in the activity of lipoprotein lipase, a initial enzyme that plays an important role in the metabolism, transport and tissue uptake of lipid fractions. Lithium is shown to reduce the activity of this enzyme. The inhibitory effect of lithium was potentiated in the presence of citrate. It had already been recorded that citrate makes lithium very soluble and a lot of works were undertaken to make citrate salt of lithium for therapeutic purposes ²². The precise mechanism by which lithium inhibits lipoprotein lipase activity was not known closely, however the activity of this enzyme depends on the presence of free-SH groups ²³. It is possible that lithium by interacting with some essential-SH groups in the active site of the enzyme reduces enzyme activity.

In addition, in the present study used turmeric extract (curcuminoids). The main component of curcuminoids was curcumin antioxidant activity of curcumin has been reported as early as 1975. It acts as a scavenger of oxygen free radicals²⁴. *In vitro*, curcumin can significantly inhibit the generation of reactive oxygen species (ROS) such as superoxide anions, H_2O_2 , and nitrite radical generation by activated macrophages, which play an important role in inflammation²⁵. curcumin reduces serum and liver cholesterol levels in mice²⁶ and also reported to have anti-inflammatory activity in standard animal models²⁷. It has been reported by Ruby²⁸ studying the antitumor and antioxidant activity of natural curcuminoids that curcumin inhibits the generation of superoxide radicals. curcumin also reduced lipid peroxidation in rat liver microsomes, erythrocyte membranes, and brain homogenates^{29,30}. Because ROS had been implicated in the development of potential to control these diseases though it was antioxidant activity. Several studies have reported the antioxidant property of curcumin, increment endogenous antioxidant levels³¹. curcumin also inhibits the induction of nitric oxide synthase in activated macrophages and down regulates nitric oxide formation^{32,33}.

The results in the table (3) and (4) reported significant decrease (p<0.05) in the levels of SOD and GPX and significant increase (p<0.05) in the level of MDA. In the group of rats were demonstrated lithium carbonate only for the 6, 8 weeks. Several studies were recorded lithium toxicity can be connected with oxidative stress ^{34,35,36,37} but contradicting outcomes were also reported^{38,39}. Furthermore, oxidative stress was also found to be involved into the pathophysiology of bipolar disorder⁴⁰. As long term lithium therapy was used in the cure of this disease, the question of the influence of lithium on oxidative stress was an issue of great importance. The organisms developed a complex system of defense against oxidative stress which includes numerous substances, among other things antioxidant enzymes, namely superoxide dismutase, glutathione peroxidase and catalase ^{41,42}.

Lithium given for a longer period (2 months) caused a significant decrease in SOD and GPX in rats. Naziroglu⁴³ found decreased SOD and GPX in healthy subjects undergoing lithium treatment. These outcomes seem to confirm our assumption regarding possible adjuvant application of any antioxidant in lithium treatment.

Intraperitoneal lithium treatment for a period of 7 days changed neither SOD nor GPX in rat livers⁴⁴. Lithium carbonate provided to rats in drinking water for a period of 4 weeks markedly influenced neither GPX nor SOD in liver. Lipid peroxidation (MDA) was also unaffected⁴⁵. In other study one month administration in diet resulted in the decrease of hepatic lipid peroxidation in rats under different dietary regimens⁴⁶. Chinese scientists lithium and Long reported that lithium exerted the divergent effect on lipid peroxidation in rat livers. Lower doses resulted in inhibition, whereas the higher concentrations showed a stimulating influence⁴⁷. An increased GPX activity in the liver of diabetic rats was observed, whereas SOD remained unchanged as a consequence of lithium treatment ⁴⁸. However, it has already been mentioned that differences of the lithium's action in physiologic and pathologic states in rats were observed. Concerning human beings, it has also been reported that the lithium effect on cognitive functions differed in healthy subjects from that found in psychiatric patients⁴⁹.

In addition, the results show Dietary turmeric lowered lipid peroxidation by enhancing the activities of antioxidant enzymes its conform with previous studies⁵⁰.

It exerted beneficial effect in preventing oxidative stress in rats ⁵¹. Dietary antioxidants have preventative effects on oxidative stress. The antioxidant mechanism of turmeric was attributed to its major component in curcuminoid compoundes called curcumin. curcumin was conjugated structure which includes two methoxylated phenols and an enol form of β -diketone. The structure showed a typical radical trapping ability as a chain breaking antioxidant ⁵². Curcumin exhibit a differential antioxidant activity in several *in vitro* and *in vivo* models, for example, preventing lipid peroxidation in a variety of cells such as erythrocytes and rat liver microsomes, where peroxidation is induced by Fenton's reagent, as well as for metals and hydrogen peroxide (H₂O₂). Furthermore, it has been reported that curcumin is a bi-functional antioxidant ⁵³, because of its ability to react directly with reactive species and to induce an up-regulation of various cytoprotective and antioxidant proteins. Curcumin is able to scavenge superoxide anion (O2-.)^{54,55}, hydroxyl radicals (.OH)⁵⁶, singlet oxygen⁵⁷, nitric oxide⁵⁸, peroxynitriteand peroxyl radicals (ROO.)⁵⁹.

Together, these mechanisms might explain, at least in part, some of the cytoprotective effects of this compound. Features as the presence of phenolic groups in the structure of curcumin explains its ability to react with reactive oxygen species (ROS) and reactive nitrogen species (RNS) and might probably be one of the mechanisms through which curcumin treatment protects erythrocytes from oxidative damage.

In conclusion

Oral administration of turmeric extract (*curcumalonga*) to toxicity male rats for 6, 8 weeks decrease the lipid profile parameters also increase antioxidant enzyme levels in serum. Therefore, this study recommended that intake of turmeric in food may be useful for patients who suffer from manic depression to reduce the side effects of lithium when taken for long period.

Table (1) Effect of the interference between the extract	s and dose in	the lipid	profile levels	in the rats
treated with lithium carbonate for six weeks.				

Treatment	Daga	ТС	TG	HDL	VLDL	LDL
1 reatment	Dose	(mg/dl)	(mg/dl)	(mg/dl)	(mg/dl)	(mg/dl)
1.32003	4 mg/kg	63.67±2.88	53.67±16.86	26.33±5.03	10.73 ± 3.37	26.60±8.87
LIZCOS	8 mg/kg	81.33±1.15	60.33±19.55	41.33±2.08	12.07±3.91	27.93±2.91
Li2CO3&	4 mg/kg	59.33±1.15	34.00±5.29	34.33±4.50	6.80±1.05	15.53±0.81
С	8 mg/kg	63.33±4.16	36.33±3.21	45.00±6.55	7.27±0.64	11.07 ± 7.10
Con	trol	49.33±15.04	25.33±12.85	37.00±8.54	6.67±1.52	7.00 ± 3.00
L.S.D	. 0.05	11.592	13.454	8.565	2.694	6.581

Number of animals = 5 for each group Each value represents mean \pm S.E. Li2CO3 : Lithium carbonate C : Turmeric extract (Curcuminoids)

Table (2) Effect of the interference between the extract	ts and dose in the lipid profile levels in the rats
treated with lithium carbonate for eight weeks.	

Treatment	Dose	TC (mg/dl)	TG (mg/dl)	HDL (mg/dl)	VLDL (mg/dl)	LDL (mg/dl)
1.32003	4 mg/kg	84.00±3.60	62.67±2.51	23.67±3.21	12.53±0.50	47.80±6.23
LIZCOS	8 mg/kg	102.00±7.21	71.67±6.65	19.33±1.15	14.33±1.33	68.33±6.99
Li2CO3&	4 mg/kg	51.67±2.88	36.00±1.73	41.67±2.88	7.20±0.34	2.80±0.34
С	8 mg/kg	57.67±3.06	46.67±4.04	46.33±3.21	9.33±0.81	2.00±1.05
Con	trol	46.67±15.27	25.33±11.01	39.33±9.01	8.00±2.00	9.00±3.60
L.S.D	. 0.05	11.592	13.454	8.565	2.694	6.581

Number of animals = 5 for each group Each value represents mean \pm S.E.

Li2CO3 : Lithium carbonate C : Turmeric extract (Curcuminoids).

Treatment	Dose	MDA (ng/ml)	SOD (ng/ml)	GPX (pg/ml)
Li2CO3	4 mg/kg	27.33±1.36	0.35 ± 0.03	91.99±28.36
	8 mg/kg	33.11±8.95	0.24±0.05	53.57±15.39
Li2CO3& C	4 mg/kg	23.94±4.39	0.16±0.06	44.00±15.85
	8 mg/kg	20.92±2.68	0.14±0.01	47.12±17.26
C	Control	24.73±2.31	0.19±0.04	82.77±27.26
L.S	.D. 0.05	14.128	0.185	36.340

Table (3) Effect of the interference between the extracts and dose in the antioxidant levels in the rats treated with lithium carbonate for six weeks.

Number of animals = 5 for each group Each value represents mean \pm S.E. Li2CO3 : Lithium carbonate C : Turmeric extract (Curcuminoids)

Table (4) Effect of the interference between the extracts and dose in the antioxidant levels in the rats treated with lithium carbonate for eight weeks.

Treatment	Dose	MDA (ng/ml)	SOD (ng/ml)	GPX (pg/ml)
Li2CO3	4 mg/kg	32.29±14.20	0.10±0.06	64.47±25.63
	8 mg/kg	31.47±12.31	0.11±0.02	56.23±45.20
Li2CO3& C	4 mg/kg	32.44±15.20	0.19±0.02	75.63±20.69
	8 mg/kg	32.69±12.10	0.18±0.03	55.28±31.52
C	Control	23.68±10.10	0.20±0.04	78.56±19.19
L.S	D. 0.05	14.128	0.185	36.340

Number of animals = 5 for each group Each value represents mean \pm S.E.

Li2CO3 : Lithium carbonate C : Turmeric extract (Curcuminoids)

References :

- 1. Subhash, M. N., Vinod, K.Y., and Srinivas, B.N. Differential effect of lithium on 5-HT1 receptorlinked system in regions of rat brain. Neurochem. Int. 35,1999, 337.
- Koong, S.S., Reynolds, J.C., Movius, E.G., Keenan, A.M., Ain, K.B., Lakshmana, M.C., and Robbin, S. J. Lithium as a potential adjuvant to 1311therapy of metastatic, well differentiated thyroid carcinoma. J. Clin. Endocrinol. Metab. 84, 1999,912.
- 3. Bogazzi, F., Bartalena, L., Brogioni, S., Scarcello, G., Burelli, A., Campomori, A., Manetti, L., Rossi, G., Pinchera, A., and Martino, E. Comparison of radioiodine with radioiodine plus lithium in the treatment of Graves' hyperthyroidism. J. Clin. Endocrinol. Metab. 84, 1999, 499.
- 4. Bogazzi, F., Bartalena, L., Campomori, A., Brogioni, S., Traion, C., DE-martino, F., Rossi, G., Lippi, F., Pinchera, A., and Martino, E. Treatment with lithium prevents serum thyroid hormone increase after thionamide withdrawal and radioiodine therapy in patients with Graves' disease. J. Clin. Endocrinol. Metab. 87, 2002, 4490.
- 5. Alvarez, G., Munoz-montanoj, R., Satrustegui, J., Avila, J., Bogonez, E., Diaz-nido, J. Lithium protects cultured neurons against β-amyloid-induced neurodegeneration. FEBS Lett. 453, 1999, 260.
- 6. MANJIH.K., MOORE G.J., CHENG. Lithium at 50: have the neuroprotective effects of this unique cation been overlooked? Biol. Psychiatry. 46, 1999, 929.
- 7. Carneys, M. and Goodwing, M. Lithium a continuing story in the treatment of bipolar disorder. ActaPsychiatr. Scand. Suppl. 426, 2005, 7.
- 8. Dichtl, B., Stevens, A., Toliervey, D. Lithium toxicity in yeast is due to the inhibition of RNA processing enzymes. EMBOJ. 16, 1997, 7184.
- 9. 9.Thakur, S.C., Thakur, S.S., Chaube, S.K. and Singh, S.P. Subchronic supplementation of lithium carbonate induces reproductive system toxicity in male rat. Reprod. Toxicol. 17, 2003, 683.
- 10. KOZMA C. Neonatal toxicity and transient neurodevelopmental deficits following prenatal exposure to lithium: Another clinical report and a review of the literature. Am. J. Med. Genet. 132, 2005, 441.
- 11. Drewa, G., Krzyzynska-malionwska, E., Wozniaka, A., Protas-drozd, F., Mila-kierzen, C., Rozwodowska, M., Kowaliszyn, B., and Czajkowski, R. Activity of superoxide dismutase and catalase

and the level of lipid peroxidation products reactive with TBA in patients with psoriasis. Med. Sci. Monit. 8, 2002, BR338.

- 12. Sontakke, A.N. and Tare, R.S. A duality in the roles of reactive oxygen species with respect to bone metabolism. Clin. Chim. Acta 318, 2002, 145.
- 13. Hu, H.L., Forsey, R.J., Blades, T.J, Barratt, E.J., PARMARP., POWELL J.R. Antioxidants may contribute in the fight against ageing: an in vitro model. Mech. Age. Dev. 121, 2000, 217.
- 14. Cirak, B., Inci, S., Palaoglu, S., Bertan, V. Lipid peroxidation in cerebral tumors. Clin. Chim. Acta 327, 2003, 103.
- 15. Zetin, M. Psychopharmacohazardology: major hazards of the new generation of psychotherapeutic drugs. International J. Clin. Practice. 58 (1): 2004 ,58-79.
- 16. Weirzbicki, A.S., Hardman, T.C., Cheung, J., Patel, M., Smallberger, S., Lumb, P.J. and Lant, A.F. Relation between sodium-lithium counter transporter and hypertriglyceridemia in type V hyper lipidemia. Am. J. Hypertension (14): 2001, 32-37.
- 17. Soares, J.C., Mallinger, A.G., Dippold, C.S., Forster Wells, K., Frank, E. and Kupter, D.J. Effect of lithium on platelet membrane phosphoinositides in bipolar disorder patients. Psychopharmacology (Berl) 149 (1): 2000, 12-16.
- 18. Harborn, J.B. Phytochemical methods. 2nded (Ed.).Chapman and Hall. 1984, pp. 288.
- 19. Tietz, N. Text book of clinical chemistry, 3rd Ed., C.A. Burtis, E.R. Ashwood, W.B. Saunders . pp. 1999, 703-1699.
- 20. Tietz, N. Clinical guide to laboratory test 4th Ed., 2006, pp. 1074-1077.
- 21. Al-Rawi, K. Entrance to the Statistics. Second edition. Faculty of Agriculture and Forestry, University of Mosul. 2000.
- 22. Kerry, R. The management of patients receiving lithium treatment. In: Lithium research and therapy. (Johnson, F.N. ed.), Academic press, New York, London. 1975, pp. 143-163.
- 23. Tornqvist, H. and Belfrage, P. Purification and some properties of monoacylglycerol- hydrolysins enzyme of rat adipose tissue. J. Biol. Chem. 251: 1976, pp. 813-819.
- 24. Suramanian, M.; Sreejayan, N.; Rao, N.; Devasagayam, T. and Singh, B.Diminnution of singlet oxygen DNA damage by curcumin and related antioxidants, Mutat Res. 1994, pp. 249-311.
- 25. Joe, B. and Lokesh, B. Role of capsaicin, curcumin and dietary fatty acids in lowering the generation of reactive oxygen species in rat peritoneal macrophages, Biochim. Biophys. Acta, 1994, pp. 224-255.
- 26. Suadicani, P.; Hein, H. and Gyntelberg, F. Serum selenium concentration and risk of ischaemic heart disease in a prospective cohort study of 3000 males. Atherosclerosis, 96: 1992, pp. 33-42.
- 27. Ammon, H. and Wahl, M. Pharmacology of Curcuma longa. Planta Med. Feb; 57(1): 1991, pp. 1-7.
- 28. Ruby, A.; Kuttan, G.; Dinesh-Babu, K.; Rajeshkharan, K. and Kuttan, R. Antitumor and antioxidant activity of natural curcuminnoids, Cancer Lett. 1995, pp. 79-94.
- 29. Reddy, A. and Lokesh, B. Studies on spice principles as antioxidant in the the inhibition lipid peroxidation of rat liver microsomes, Mol. Cell Biochem. 1992, pp. 111-117.
- 30. Reddy, A. and Lokesh, B. Effect of dietary turmeric *(curcuma longa)* on iron induced lipid peroxidation in the rats liver, Food Chem. Toxicol. 1994, pp. 32-279.
- 31. Venkatesan, N. Curcumin attenuation of acute adriamycin myocardial toxicity in rats, Br. J Pharmacol. 1998, pp. 124-425.
- 32. Brouet, I. and Obshima, H.Curcumin, an antitumor promoter, and anti-inflammatory agent inhibits induction of nitric oxide synthase in activated macrophages, Biochem. Biophys. Res commun.1995, pp. 206-533.
- 33. Pan, M.; Lin-Shiau, S. and Lin, J. Comparative studies on the suppression of nitric oxide synthase by curcumin and its hydrogenated metabolites through down regulation of I kappa B kinase and NF kappa B activation in macrophages, Biochem. Pharmacol. 2000, pp. 60-65.
- Malhotra, A. and Dhawan, D. Zinc improves antioxidative enzymes in red blood cells and hematology in lithium-treated rats. Nutr. Res 28: 2008, pp. 43-50.
- 35. Ahmad, M.; Elnakady, Y.; Farooq, M. and Wadaan, M. Lithiuminduced toxicity in rats: blood serum chemistry, antioxidative enzymes in red blood cells and histopathological studies. Biol. Pharm.Bull 34: 2011, pp. 272-277.
- Nciri, R.; Allagui, M. and Bourogaa, E. Lipid peroxidation, antioxidant activities and stress protein (HSP72/73, GRP94) expression in kidney and liver of rats under lithium treatment. J. Physiol. Biochem. 68: 2012, pp. 11-18.

- 37. Toplan, S.; Dariyerli, N.; Ozdemir, S.; Ozcelik, D.; Zengin, E. and Akyolcu, M. Lithium induced hypothyroidism: oxidative stress and osmotic fragility status in rats. Biol. Trace Elem. Res. 152: 2013, pp. 373-378.
- Machado-Vieira, R.; Andreazza, A. and Viale, C. Oxidative stress parameters in unmedicated and treated bipolar subjects during initial manic episode: a possible role for lithium antioxidant effects. Neurosci. Lett. 421: 2007, pp. 33-36.
- 39. Khairova, R.; Pawar, R. and Salvadore, G. Effects of lithium on oxidative stress parameters in healthy subjects. Mol. Med. Rep. 5: 2012, pp. 680-682.
- 40. Bruning, C.; Prigol, M.; Luchese, C.; Pinton, S. and Nogueira, C. Diphenyldiselenide ameliorates behavioral and oxidative parameters in an animal model of mania induced by ouabain. Prog. Neuropsychopharmacol Biol. Psychiatry 38: 2012, pp. 168-174.
- 41. Naziroglu, M. New molecular mechanisms on the activation of TRPM2 channels by oxidative stress and ADP-ribose. Neurochem. Res. 32: 2007, pp. 1990-2001.
- 42. De-Freitas, A. and Rocha, J. Diphenyldiselenide and analogs are substrates of cerebral rat thioredoxinreductase: a pathway for their neuroprotective effects. Neurosci. Lett. 503: 2011, pp. 1-5.
- 43. Naziroglu, M. Molecular role of catalase on oxidative stress induced Ca(2+) signaling and TRP cation channel activation in nervous system. J. Recept. Signal Transduct Res. 32: 2012, pp. 134-141.
- 44. Abdalla, D.andBechara E. The effect of chlorpromazine and Li2CO3 on the superoxide dismutase and glutathione peroxidase activities of rat brain, liver and erythrocytes. Biochem. Mol. Biol. Int. 34: 1994, 1085.
- 45. HUM, O. ; WUY, D. and WUH, S. Influence of streptozotocin-induced diabetes in rats on the lithium content of tissue and the effect of dietary lithium supplements on this diabetic condition. Metabolism 48, 1999, 558.
- 46. Tandona, E.; Dhawand, K. and Nagpaul, J. Effect of lithium on hepatic lipid peroxidation and antioxidative enzymes under different dietary protein regiments. J. Appl. Toxicol. 18: 1998, pp.187.
- 47. LIS, V. and Long, S. Effects of manganese, zircon and lithium alone on rat liver lipid peroxidation. Wei Sheng Yan Jiu. 30: 2001, pp. 142.
- 48. Srivastava, P.; Saxena, A.; Kale, R. and Baquern, Z. Insulin like effects of lithium and vanadate on the altered antioxidant status of diabetic rats. Res. Commun. Chem. Pathol. Pharmacol. 80: 1993, pp. 283.
- 49. Suwalska, A.; Lokod, Q. and Rybakowski, W. The influence of mood normalizing drugs on the cognitive functions. Psychiat. Pol. 35: 2001, pp. 245.
- 50. Arun, N. and Nalini, N. Efficacy of turmeric on blood sugar and polyol pathway in diabetic albino rats. Plant Foods Hum. Nutr. 57(1): 2002, pp. 41-52.
- 51. Suryanarayana, P.; Satyanarayana, A.; Balakrishna, N.; Zheng, H.; Gal, S.; Weiner. L.; Bar-Am, O.; Warshawsky, A.; Fridkin, M. and Youdim, M. Novel multifunctional neuroprotective iron chelator-monoamine oxidaseinhibitor drugs for neurodegenerative diseases: in vitro studies on antioxidant activity, prevention of lipid peroxide formation and monoamine oxidase inhibition. J. Neurochem., 95(1): 2007, pp. 68-78.
- 52. Masuda, T.; Maekawa, T.; Hidaka, K.; Bando, H.; Takeda, Y. and Yamaguchi, H. Chemical studies on antioxidant mechanism of curcumin: Analysis of oxidative coupling products from curcumin and linoleate. J. Agric. Food Chem. 49: 2001, pp. 2539-2547.
- 53. Dinkova-Kostova, A. and Talalay, P. Direct and indirect antioxidant properties of inducers of cytoprotective proteins. Molecular Nutr. Food Res. 52: 2008, pp. 128-138.
- 54. 54. Sreejayan, R. Nitric oxide scavenging by curcuminoids. J. Pharm. Pharmacol. 49: 1997, pp. 105-107.
- 55. Ak, T. and Gulcin, I. Antioxidant and radical scavenging properties of curcumin. Chem. Biol. Interact. 174: 2008, pp. 27-37.
- 56. Barzegar, A. and Moosavi-Movahedi, A. Intracellular ROS protection efficiency and free radical scavenging activity of curcumin. Plos. 6: 2011, pp. 12-26.
- 57. Das, K. and Das, C. Curcumin (diferuloylmethane) a singlet oxygen (1O2) quencher. Biochem. Biophys. Res. Commun. 295: 2002, pp. 62-66.
- Sumanont, Y.; Murakami, Y.; Tohda, M.; Vajragupta, O.; Matsumoto, K. and Watanabe, L. (2004). Evaluation of the nitric oxide radical scavenging activity of manganese complexes of curcumin and its derivative. Biol. Pharm. Bull. 27: 2004, pp. 170-173.

59. Kim, J.; Kim, A.; Chung, H.; Han, S.; Kim, B. and Chot, J. In vitro peroxynitrite scavenging activity of diarylheptanoids from *Curcuma longa*. Phytother. Res. 17: 2003, pp. 481-484.

```
****
```