



## Comparative Evaluation of the Antioxidant Activity of Some commonly used Spices

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**Abstract:** The antioxidant activity, total phenolic content (TPC) and total flavonoid content (TFC) of four natural spices namely fennel fruits (*Foeniculum vulgare*), fenugreek seeds (*Trigonella foenum-graecum*), coriander seeds (*Coriandrum sativum* Linn.) and black pepper fruits (*Piper nigrum*) were investigated. In antioxidant assay, DPPH (2,2-diphenyl-1-picrylhydrazyl) scavenging activity and ferric reducing activity studies were performed using ascorbic acid and gallic acid as standard antioxidants, respectively. *In-vitro* antioxidant study of the methanolic extracts of the tested condiments showed significant activity in DPPH method with appreciable low  $IC_{50}$  values. The ferric reducing properties of these spices were also found to be appreciable. The TFC and TPC revealed that the spices had high contents of phenolics and flavonoids. These studies suggested that these spices can be used as rich sources of natural antioxidants against various oxidative stress related diseases.

**Keywords:** Spices, Flavonoid content, phenolic content, DPPH assay, Ferric reducing power.

### Introduction

Antioxidants are referred to as compounds which are able to inhibit the oxidation of different biomolecules and helps in repairing the damages caused to the body tissues due to oxidation processes. Various study has focused on use of antioxidants from natural sources so that they could be used as healthy additives and potential antioxidant in our daily food intake habits<sup>1</sup>. Antioxidants either synthetic or natural are potent scavengers of free radicals and have beneficial effects on human health and disease prevention<sup>2,20-22</sup>.

Oxidative metabolism is essential for the survival of cells. A side effect of this dependence is the production of free radicals and other reactive oxygen species that cause oxidative changes. The most widely used synthetic antioxidants in food i.e., butylated hydroxyl toluene (BHT) and butylated hydroxyl anisole (BHA) are very effective in their role as antioxidants. However, their use in food products has been failing off due to their instability or their suspected action as promoters of carcinogenesis. For this reason, there is a growing interest in the studies of natural healthy (nontoxic) additives as potential antioxidants<sup>1</sup>. It is known that constituents of plants are associated in reducing the risk of many chronic diseases, in which antioxidants play a major role in their protective effects<sup>3</sup>.

Herbs and spices are considered an important part of human diet and have been used for thousands of years in traditional medicine and also to enhance color, flavor and aroma of the food<sup>4</sup>. The presence of active components in spices has been demonstrated over the last 30 years and their therapeutic properties have been demonstrated by the presence of many substances, including some vitamins, flavonoids, terpenoids, carotenoids, phytoestrogens and minerals<sup>5</sup>.

Spices are common food additives, which are always used as flavoring, seasoning, and coloring agents, and sometimes as preservatives, but they may also be used as medicines. Many spices have been recognized to have medicinal properties and possess many beneficial effects on health such as antioxidant activity, digestive stimulant action, anti-inflammatory, antimicrobial, hypolipidemic, antimutagenic, anticarcinogenic potential etc. The presence of antioxidative and antimicrobial phenolic constituents in spices gives food-preserving properties<sup>6</sup>.

Black pepper (*Piper nigrum*) is one of the most popular spice products in oriental countries, largely used as a flavoring agent in foods. The components of the pepper's extract that contribute to its value as a food additive are the volatile oil for its aroma and the alkaloid compounds for the pungency<sup>7</sup>. Black pepper is used in skin care, muscle and joint pains, and in improving blood circulation and respiratory systems. The seeds of coriander (*Coriandrum sativum* Linn.) are mainly responsible for the medical use of coriander and have been used as a drug for indigestion, against worms, rheumatism and pain in the joints<sup>8</sup>. Fenugreek (*Trigonella foenum-graecum*) has been reported to exhibit pharmacological properties such as antitumor, antiviral, antimicrobial, anti-inflammatory, hypotensive and antioxidant activity<sup>9</sup>. Fennel fruit (*Foeniculum vulgare*), a dry seed, is traditionally used as anti-inflammatory, analgesic, carminative, diuretic, antispasmodic agent, in treatment of glaucoma and hypertension<sup>10</sup>. These ingredients also have been used in ayurvedic medicines for ailment of various disorders majorly gastrointestinal imbalances which may have been attributed due to their high antioxidant activity<sup>11</sup>.

Considering the beneficial effects of spices in treating diseases, methanolic extracts of these commonly used spices i.e. coriander seeds, fenugreek seeds, black pepper fruits and fennel fruit were evaluated for their comparative antioxidant activity evaluation by various *in-vitro* antioxidant models.

## Experimental

### Chemicals

Aluminum chloride (AlCl<sub>3</sub>), Sodium hydroxide (NaOH), Sodium nitrite (NaNO<sub>2</sub>), DPPH (1,1-Diphenyl-2-picrylhydrazyl), Folin-ciocateau reagent, Sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>), Trichloroacetic acid (TCA), Potassium ferricyanide, Sodium phosphate buffer, Ferric chloride, Ascorbic acid, Gallic acid and Quercetin.

### Collection and authentication

The spices were collected from the local market in Bhubaneswar, India and authenticated by a local botanist.

### Extraction

The extraction procedure was done by the process of maceration in which the spices were soaked in methanol solvent for three days with occasional shaking for proper extraction and followed by filtration. The filtrates were evaporated under vacuum and dried to a constant weight using a freeze-drier.

### In vitro antioxidant activity evaluation

#### DPPH radical scavenging activity method

Stock solutions (1 mg/ml) for each ingredient extract was prepared in methanol and diluted subsequently to various concentrations with methanol in time of assay. For the assay, 1 ml of different concentration of spice extracts were mixed with 500 μl of DPPH solution and 1.5 ml of methanol in sequence and then the samples' were scanned against blank at 517 nm. Similarly readings were taken for standard ascorbic acid<sup>12</sup>. IC<sub>50</sub> values of DPPH radical scavenging activity were determined by the following formula: % radical scavenging activity =  $(Abs_{control} - Abs_{sample}) / Abs_{control} \times 100$

#### Reducing power assay

2.5 ml of the spices extracts (1 mg/ml) were taken and mixed with 2.5 ml sodium phosphate buffer (0.2M, pH 6.6) and 2.5 ml of 1 % potassium ferricyanide solution. The mixtures were incubated at 50°C for 20

minutes. Then 2.5 ml of trichloroacetic acid (10%) were added to them and the solutions were centrifuged at 650 rpm for 10 mins. Then 5ml of the supernatants were mixed with 5 ml of distilled water and 1 ml of ferric chloride solution (0.1%) and the absorbances were measured at 700 nm. Higher absorbances of the reaction mixtures indicate a higher reducing power<sup>2</sup>.

### Total flavonoid content (TFC)

In a 10 ml test tube, 0.3 ml of extracts (1 mg/ml), 3.4 ml of 30% aqueous methanol, 0.15 ml of sodium nitrite (0.5 M) and 0.15 ml of aluminum chloride (0.3 M) were mixed. After five minutes 1 ml of sodium hydroxide (1M) was added, mixed well and absorbance was taken at 506 nm. The standard curve was constructed using quercetin concentration range of 50-250  $\mu\text{g/ml}$ . Total flavonoids were expressed as quercetin equivalents(mcg) per gram of extract<sup>13</sup>.

### Total phenolic content (TPC)

As per the standard protocol, 40  $\mu\text{l}$  of a methanolic extract (1mg/ml) of individual spice, 3.16 ml of distilled water and 200  $\mu\text{l}$  of Folin – Ciocalteu reagent were mixed. After incubation of 8minutes, 600  $\mu\text{l}$  of sodium carbonate solution was added and mixed. The mixture was incubated at 40<sup>0</sup>C for 30 min before taking its absorbance at 765 nm. The calibration curve was prepared using Gallic acid as standard from 100 to 200  $\mu\text{g/ml}$ . TPC were expressed as gallic acid equivalent(mcg) per gram of extracts<sup>13</sup>.



Fig.1: A-Fruits of black pepper, B-Fennel fruit, C-Fenugreek seeds, D-Coriander seeds

## Results

### Total phenolic and flavonoid contents

Total flavonoids and total phenolic contents were determined as quercetin equivalents in micrograms per gram of extract (QE/gm of extract), while total phenolic contents were calculated as gallic acid equivalents in micrograms per gram of extract (GAE/gm of extract). Results from the quantitative determination of total phenolics and flavonoids of the methanolic extracts of the different spices were summarized in Table 1 and Table 2. All the calculations were done using standard equation obtained from standard calibration curves of quercetin and gallic acid as shown in Figure 2 and Figure 3 respectively. All the spices showed high total flavonoid content and total phenolic content. TFC of *Coriandrum sativum* Linn. was  $23.87 \pm 0.81$ , *Foeniculum vulgare* was  $23.41 \pm 0.656$ , *Piper nigrum* was  $20.83 \pm 1.649$  and *Trigonella foenum-graecum* was  $10.58 \pm 0.46$  mcg QE/gm of extract. Again, TPC of *Coriandrum sativum* Linn. was  $76.376 \pm$

0.99, *Foeneculumvulgare* was  $51.56 \pm 3.22$ , *Piper nigrum* was  $119.39 \pm 0.99$  and *Trigonella foenum- graecum* was  $59.18 \pm 5.79$  mcg GAE/gm of extract. These results indicated that the phenolic and flavonoid compounds had a major contribution to the antioxidant capacity of herbal spices.

**Table 1: TFC (Total flavonoids content) of the Spices**

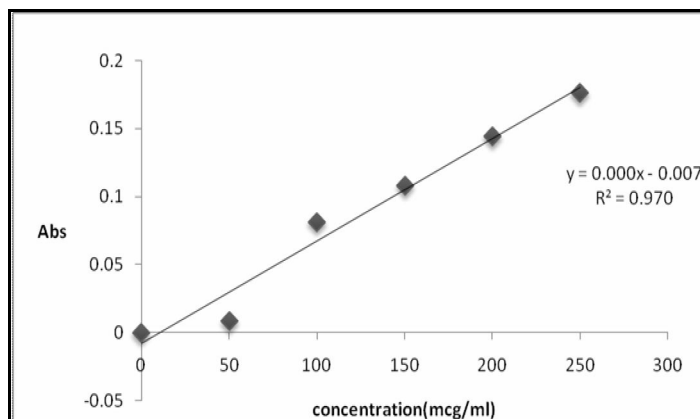
| SI No. | Spices                           | Total Flavonoid Content(QE/g of extract)* |
|--------|----------------------------------|---|
| 1      | <i>Coriandrum sativum L.</i>     | $23.87 \pm 0.81$                          |
| 2      | <i>Foeneculum vulgare</i>        | $23.41 \pm 0.656$                         |
| 3      | <i>Piper nigrum</i>              | $20.83 \pm 1.649$                         |
| 4      | <i>Trigonella foenum-graecum</i> | $10.58 \pm 0.46$                          |

\*(Values are expressed as Mean  $\pm$  S.D.)

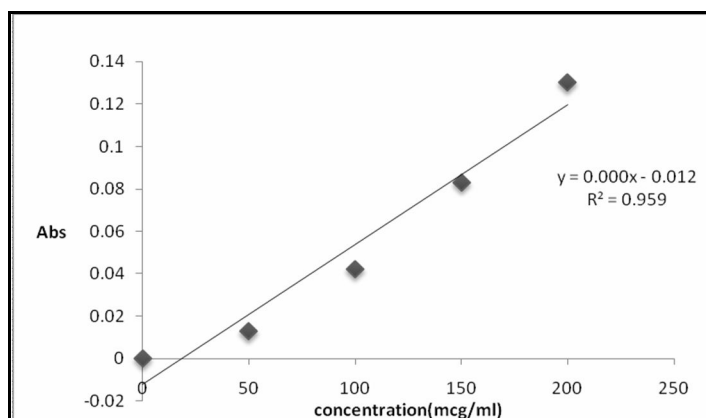
**Table 2: TPC (Total phenolic content) of the Spices**

| SI No. | Spices                          | Total Phenolic Content(mcg/ml of GA)* |
|--------|---------------------------------|---------------------------------------|
| 1      | <i>Coriandrum sativum Linn.</i> | $76.376 \pm 0.99$                     |
| 2      | <i>Foeneculum vulgare</i>       | $51.56 \pm 3.22$                      |
| 3      | <i>Piper nigrum</i>             | $119.39 \pm 0.99$                     |
| 4      | <i>Trigonella foenumgraecum</i> | $59.18 \pm 5.79$                      |

\*(Values are expressed as Mean  $\pm$  S.D.)



**Fig .2: Calibration curve of quercetin standard**



**Fig .3: Calibration curve of Gallic acid standard**

### DPPH radical scavenging activity

The parameter  $IC_{50}$  is used for the interpretation of the results from the DPPH method and is defined as the concentration of substrate that causes 50% loss of the DPPH activity. Spice extracts examined in relation to their  $IC_{50}$  value and  $IC_{50}$  values of *Coriandrum sativum* Linn. was  $751 \pm 1.414$ , *Foeneculum vulgare* was  $746.9 \pm 2.81$ , *Piper nigrum* was  $444.62 \pm 3.38$  and *Trigonella foenum-graecum* was  $772 \pm 2.45$  (mcg /ml). The methanolic extract of *Piper nigrum* has the highest scavenging activity as compared to the other natural spices used in this study due to its less  $IC_{50}$  value as shown in Table 3.

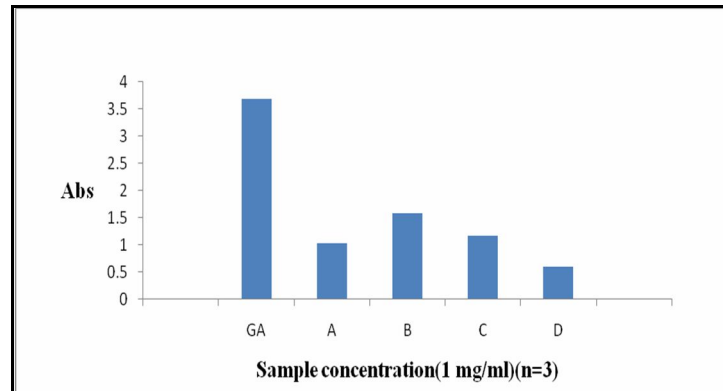
**Table 3:  $IC_{50}$  values of the spices and standard ascorbic acid in DPPH radical scavenging assay**

| SI No. | Ingredients and Standard         | $IC_{50}$ Value(mcg/ml)* |
|--------|----------------------------------|--------------------------|
| 1      | <i>Coriandrum sativum</i> Linn.  | $751 \pm 1.414$          |
| 2      | <i>Foeneculum vulgare</i>        | $746.9 \pm 2.81$         |
| 3      | <i>Piper nigrum</i>              | $444.62 \pm 3.38$        |
| 4      | <i>Trigonella foenum graecum</i> | $772 \pm 2.45$           |
| 5      | Ascorbic acid                    | $19.83 \pm 0.11$         |

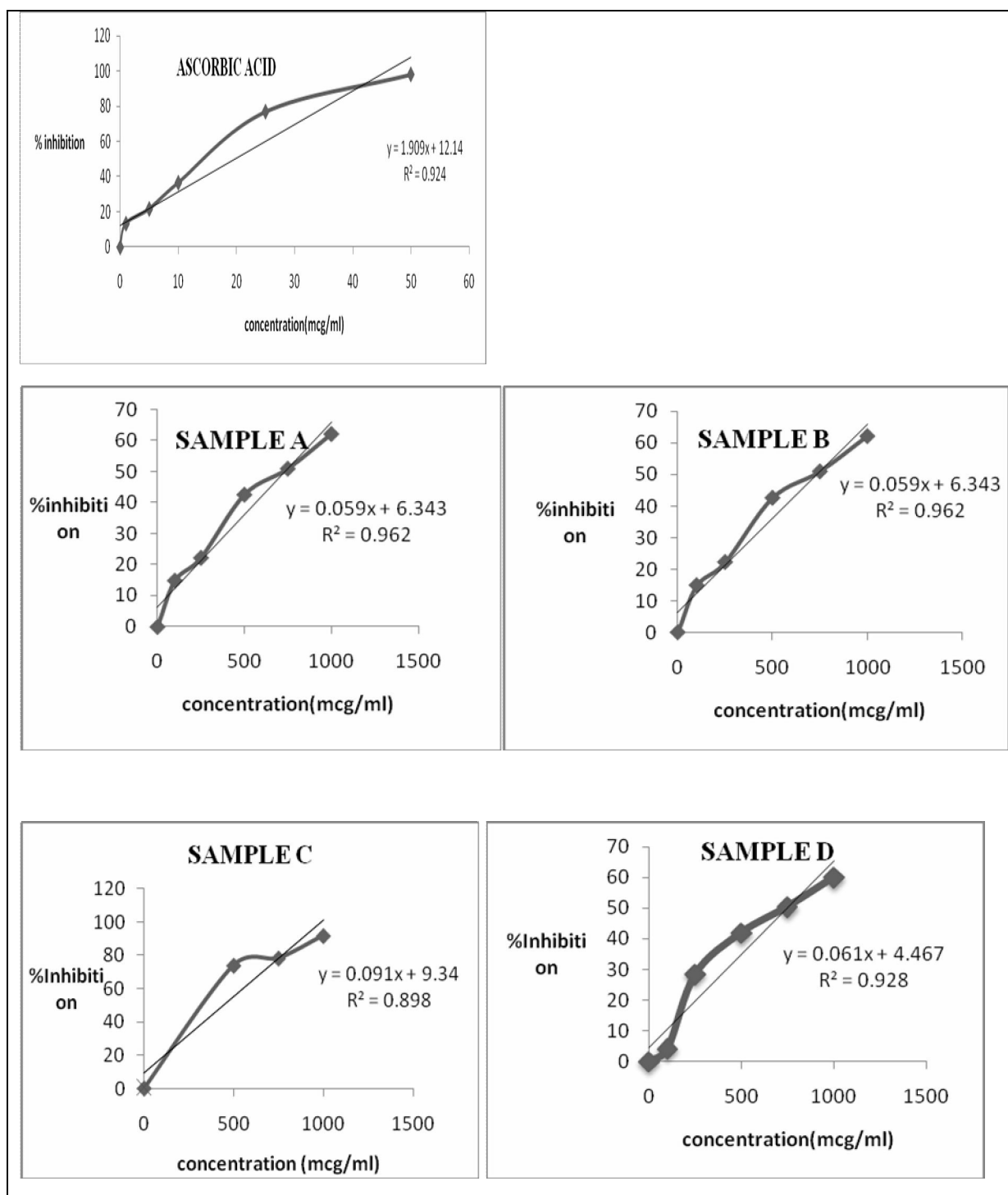
\*(Values are expressed as Mean  $\pm$  S.D.)

### Reducing power assay

In this assay, the yellow color of the test solution changes to green depending on the reducing power of test specimen. The presence of reductants in the solution causes the reduction of the  $Fe^{3+}$ /ferricyanide complex to the ferrous form. Therefore,  $Fe^{2+}$  can be monitored by the measurement of the absorbance at  $700 \text{ nm}^{14}$ . The order of ferric reducing property of tested spice samples were found to be *Foeneculum vulgare* > *Piper nigrum* > *Coriandrum sativum* L. > *Trigonella foenum-graecum* (Figure 4).



**Fig.4: Shows the dose response plot for reducing powers of the spice extract samples (A- *Coriandrum sativum* L., B- *Foeneculum vulgare*, C- *Piper nigrum*, and D- *Trigonella foenum-graecum*) and standard Gallic acid (GA)**



**Fig.5: Percentage Inhibition versus concentration plot of samples (A- *Coriandrum sativum L.*, B- *Foeniculum vulgare*, C- *Piper nigrum*, D- *Trigonella foenum-graecum*) and standard ascorbic acid in DPPH radical scavenging activity study.**

## Discussion

Naturally antioxidants present in many plants, food, and beverages offer health benefits in preventing various diseases by fighting cellular damage caused by free radicals in the body<sup>15</sup>. Free radicals have been implicated in many disease conditions, the important ones being superoxide radical, hydroxy radical, peroxy radical and singlet oxygen. Herbal drugs containing radical scavengers are gaining importance in treating such diseases<sup>16</sup>. The reducing properties are generally associated with the presence of reductones, which have been shown to exert antioxidant action by breaking the free radical chain by donating a hydrogen atom and may have great relevance in the prevention and treatment of diseases associated with oxidants or free radicals.

The DPPH assay is based on the ability of the antioxidants present in the sample to decolorize DPPH free radical by virtue of their scavenging activities. Ascorbic acid was chosen as the reference antioxidant for this test. The DPPH method is widely used to test the ability of compounds to act as free radical scavengers or hydrogen donors, and to evaluate antioxidant capacity. The parameter IC<sub>50</sub> (efficient concentration value), is used for the interpretation of the results from the DPPH method and is defined as the concentration of substrate that causes 50% loss of the DPPH activity (color)<sup>17</sup>.

Phenols are very important plant constituent because of their scavenging ability due to their hydroxyl group. It has been recognized that flavonoids show antioxidant activity and their effects on human health are considerable. The mechanisms of action of flavonoids are through scavenging or chelating process. The flavonoids which contain hydroxyls are responsible for the radical scavenging effect in the plants. It has been reported that the antioxidant effect of plant products is mainly due to radical scavenging activity of phenolic compounds such as flavonoids, polyphenols, tannins, and phenoliterpenes<sup>18</sup>. The high content of flavonoids and polyphenolic compounds in the tested spices may have contributed in their antioxidant activity may be due to their redox properties, which can play an important role in adsorbing and neutralizing free radicals, quenching singlet and triplet oxygen, or decomposing peroxides<sup>19</sup>.

## Conclusion

In the present study the methanolic extracts of the selected spices have been evaluated for potential antioxidant activities. Antioxidant activities of the spices were high enough for the spices to be known as new and natural sources of antioxidant substances for use as natural additives in food. The amount of phenolic content in the extract of *piper nigrum* may be the reason for its potent antioxidant activity whereas other spices have significant phenolic and flavonoids contents which assure their antioxidant properties. It could be concluded that methanolic extracts of these spices could be used as natural means of antioxidants. Hence, inclusion of these spices in the cuisine will be helpful in preventing or slowing the progress of various oxidative stress related diseases.

## References

1. Msaada K, Jemia MB, Salem N, Bachrouch O, Sriti J, Tammar S et al. Antioxidant activity of methanolic extracts from three coriander (*Coriandrum sativum L.*) fruit varieties. *Arabian J. of Chem.*, 2013,12(011).
2. Mohamed DA, I-okbi SY. In vivo evaluation of antioxidant and anti-inflammatory activity of different extracts of date fruits in adjuvant arthritis. *Polish J. of food and nutr. Sci.*, 2004, 13/54(4): 397–402.
3. Saeed N, Khan MR, Shabbir M. Antioxidant activity, total phenolic and total flavonoid contents of whole extracts *Torilis leptophylla L.* *BMC Compl. and Alt. Med.*, 2012, 12:221.
4. Andrade KS and Ferreira SRS. Antioxidant activity of black pepper (*piper nigrum l.*) Oil obtained by supercritical CO<sub>2</sub>. III Iberoamerican Conference on Supercritical Fluids Cartagena de Indias (Colombia), 2013.
5. Shan B, Cai YZ, Sun M, Corke H. Antioxidant Capacity of 26 Spice Extracts and Characterization of Their Phenolic Constituents. *J. Agric. Food Chem.*, 2005, 53 (20): 7749–7759.
6. Zarai Z, Boujelbene E, Salem NB, Gargouri Sayari YA. Antioxidant and antimicrobial activities of various solvent extracts piperine and piperic acid from *Piper nigrum*. *LWT Food Science and Technology*, 2012.
7. Ferreira SRS, Nikolov ZL, Doraiswamy LK, Meireles MAA, Petenate AJ. Supercritical fluid extraction of black pepper (*Piper nigrum L.*) essential oil. *J. of Supercritical Fluids*, 1999, 14:235 – 245.
8. Wangenstein H, Samuelsen AB, Malterud KE. Antioxidant activity in extracts from coriander. *Food Chem.*, 2004, 88:293–297.
9. Bukhari SB, Iqbal Bhangar MI, Memon S. Antioxidative activity of extracts from fenugreek seeds (*Trigonella foenum-graecum*). *Pak. J. Analyt. and Env. Chem.*, 2008, 9(2).
10. Dua A, Aashwan M, Sanjeev G, Mahajan R. Bioreactive compounds and antioxidant properties of methanolic extract of fennel (*foeniculum vulgare* Miller). *Int. Research J. of Pharm.*, 2013, 4 (5).
11. Paul V, Tan A, Veronique B, Penlap B, Barthelemy Nyasse C, Joseph DB, Nguemo B. Anti-ulcer actions of the bark methanol extract of *Voacanga africana* in different experimental ulcer models in rats. *J. of Ethnopharmacol.*, 2000, 73:423–428.

12. Egdhami A, Sadegh F, Determination of total phenolic and flavonoids contents in methanolic and aqueous extracts of *Achillea millefolium*. Org. Chem. J., 2010, 2:81-84.
13. Ahmed D, Fatima K, Saeed R, Analysis of phenolic and flavonoid contents, and the anti oxidative potential and lipid peroxidation inhibitory activity of activity of methanolic extract of *Carissa opaca* roots and its fractions in different solvents. Antioxidants,2014 , 3: 671-683.
14. Msaada, Jmia MB, Salem N, Bachrouch O, Ji JS, Tammar S et al., Antioxidant activity of methanolic extracts from three coriander (*Coriandrum sativum* L.) fruit varieties. Arabian J..of Chem., 2013,12:011.
15. Svilaas A, Sakhi AK, Andersen LF, Svilaas T, Strom EC, Jacobs D., OseL, Blomhoff R. Intakes of antioxidants in coffee, wine and vegetables are correlated with plasma carotenoids in humans. J.Nutr., 2004, 134:562-7.
16. Bose A, MondalS, Gupta JK, Ghosh T, Debbhuti D, Si S , Antioxidant and free radical scavenging activities of *Cleome rutidosperma* Orient. Pharm. and Exp. Med., 2008, 8(2):135-145.
17. Proestos C, Lytoudi K, Mavromelanidou OK, Zoumpoulakis P, Sinanoglou VJ. Antioxidant Capacity of Selected Plant Extracts and Their Essential Oils. Antioxidants, 2013, 2:11-22.
18. Rahman MAA, Moon SS. Antioxidant polyphenol glycosides from the Plant *Drabanemorosa*. Bull Korean Chem.Soc., 2007, 28:827-31.
19. Kok CJ, Hof CHJ, Lensen JPM, van der VeldeG. The influence of pH on concentrations of protein and phenolics and resource quality of decomposing floating leaf material of *Nymphaea alba* L. (Nymphaeaceae) for the detritivore *Asellus aquaticus* (L.).Oecologia, 1992, 91:229-34.
20. Guno Sindhu Chakraborty, Antioxidant Activity of *Abutilon Indicum* Leaves, Int.J. PharmTech Res.2009,1(4), pp 1314-1316.
21. Jan R. Assa, Simon B. Widjanarko, Joni Kusnadi, Siegfried Berhimpon, Antioxidant Potential of Flesh, Seed and Mace of Nutmeg (*Myristica fragrans* Houtt), Int.J. ChemTech Res.2014,6(4),pp 2460-2468.
22. M. Boudkhili, H. Greche, S. Bouhdid, F. Zerargui, and L. Aarab, In vitro antioxidant and antibacterial properties of some Moroccan Medicinal Plants, Int.J.PharmTech Res.2012,4(2), pp 637-642.

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