

Bioinformatics informations for Constructed Mammalian expression vector using nested PCR technique

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Abstract : Present study aims to used virtual design to transfer cytomegalovirus premotor (CMV) and red fluorescence protein gene (RFP) to mammalian expression vector by nested PCR technique, free bioinformatics soft wear were used to design primers, linkers and vector maps, the results show that DII design had 6179bp which created by insertion 1463 bp after cutting with *AflII* which insert in GFP- vector at site 1618 bp. The nested PCR product were 1473 bp ,1461 bp without any addition , 1481 bp after linker and supported nucleotides addition using nested PCR .1463 bp after cutting with *AflII* which insert in GFP- vector at site 1618 bp.

Key words : virtual design, nested PCR technique, free bioinformatics soft wear.

Introduction

Nested PCR was one of important technique used in genetic researches and genetic engineering laboratories, nested PCR based on amplified specific DNA sequence using 4 primer in two steps, sometimes the PCR product in the first step using as DNA template in the second step. Nested PCR used for more fidelity amplification and for amplified target DNA sequence with additive nucleotides that added to primers with every steps^(1,2).

The bioinformatics conceders as a main part of genetic engineering and genetic study programmers were Asttaau programmers that carry out specialized programs to deal with the DNA data, some of these programs were closed with company produced genetic materials and others are free online, these software have been updating to overcome problems associated with genetic studies^(3,4).

Mammalian expression vectors were constructed into different patterns and the target gene was inserted in different sites in back bone vector and this insertion must be based on genetic engineering role in gene cloning. Constructed mammalian vector begins theoretically by bioinformatics software's to design primers, linkers and vectors. Different strategies have been used DNA sequence manipulation for primers, linkers, probes and vectors for more specify and new designs which have new features suitable with target gene transcription, translation, regulation and post translation processing in transform cells, thus new sequences are added to vectors for these aims like Internal ribosome entrysite IRES, inverted repeats IR, regulation sequences and restriction site sequence⁽⁵⁻⁷⁾.

Subjected and software's

Data and raw DNA sequence: Turbo-GFP vector and Turbo-RFP sequence from Evrogen company <http://www.evrogen.com/products/vectors/pTurboGFP-N/pTurboGFP-N.shtml> and **Error! Hyperlink reference not valid.**, (2013)⁽⁸⁻¹¹⁾, these sequences were processed in the following software's

Table (1) online bioinformatics software's used in vector design

PCR primer design	http://primer3.ut.ee/ http://www.biomol.unb.br/sms2/pcr_primer_stats.html
Vector graphic and design	http://www.biomol.unb.br/sms2/rest_digest.html http://www.addgene.org/analyze-sequence/
PCR amplification	http://www.biomol.unb.br/sms2/pcr_products.html

Vector Design (DII)

Red fluorescence protein gene and CMV promoter inserted in blank vector (GFP) in intragenic sequence as following.

1- Primer Design

1-The raw data of RFP and GFP vector were download from evrogenewebsite , (Table 1)

2-RFP sequence transfer to primer3 software (table 1) to detected site of primers and design.

3-Primers ware choose, then it transfers to sms-primer state bioinformatics (table 1) for certification.

2- Linker Design.

1- Linker of *Af*III restriction site was added in to two patterns:

5'-**CTT AAG** -3'

3'-**GCC TTG**-5'

A- Forward sequence of linker restriction site was added to forward primer.

B- Forward sequence of linker restriction site was added to revers primer.

C- Revers sequence linker restriction site was added to revers primer.

2- Supported sequence was added to primers according to *biolabs* guide for restriction enzyme activity support (<https://www.neb.com/tools-and-resources/usage-guidelines/cleavage-close-to-the-end-of-dna-fragments>)⁽¹²⁾.

3- Complete primers (primer sequence, linker and supporting sequence) was transfer to sms-primer state bioinformatics (table 1) for certification.

3-Sequence Digest and Map Construction

1- Products transfer to Addgene site (table 1)for physical map construction .

2- Then Product was digested using restriction digest in sequence manipulation site (Table 1)by *Af*III.

3- The GFP vector sequence was digested by *Af*III, (Appendix ,3)

4- The RFP gene sequence was inserted in GFP- vector to constructed DII vector.

5- The sequence of DII vector transferred to addgene site to creation physical map and features of the new vector.

6- The DII sequence transfer to sequence manipulation suite for creation the complete map of DII vector (translation probability of proteins in vector with restriction site).

4- Confirmatory Test

Confirmatory test performed by the same software in construction vector as a following:

- 1- Re-amplified target sequence in DII vector using same primer, positive results must be had longer than in blank vector (GFP).
- 2- Digested DII vector by *AfIII*, positive result must be give same sequence which used in creation vector D1.

Results

In this designRed fluorescence protein gene with CMV promoter were inserted in the GFP- vector, this design was performed using a nested PCR technique which based on added restricted site of *AfIII* linker with addition supported bp to employ cutting by restriction enzyme into two steps as in Table (2). The results of primer design are in the Table (2)with linkers and supported sequence.

Table (2) Primers sequences Features of RFP gene with CMV premotor amplification in free bioinformatics software to created DII vector.

Single base run	Hairpin formation	Self-annealing	GC clump	GC%	TM	Primer sequences 5"→3"	Design
pass	pass	pass	pass	33.33	64.1	F- * CTT AAG TAA TAG TAA TCA ATT ACG GGG TCA	D3
pass	pass	pass	pass	54.55	69.5	R- * CTT AAG GGA GGT GTG GGA GGT T	
pass	pass	pass	pass	41.67	74.1	F- * CGG CGACTT AAG TAA TAG TAA TCA ATT ACG GGG TCA	D4
pass	pass	pass	pass	41.67	74.1	F- * CGG CGACTT AAG TAA TAG TAA TCA ATT ACG GGG TCA	
pass	pass	pass	pass	42.86	66.6 °	F- ACT CCT * CTT AAG TTT GTA TGC TCG TCA G	D5
Pass	pass	pass	pass	40.00	68.3	R- GAT ACA ** GAA TTC ACT GGA ACA ACA CTC AAC C	

*linker sequence of *AfIII*; *supported sequence; *forward linker sequence of *AfIII*;

** Revers Linker sequence of *AfIII*

The Result of bioinformatics show that DII design had 6179 bp which consist of GFP gene, Kanamycin resistant gene and RFP gene as show in Figure (4-7) ,(4-9); some features in Table (4-7) and open reading frame in Table (4-9). After using D3 primer PCR product was 1473 bp (1461 bp without any addition , 1481 bp after second round amplified using D4 primer, 1463 bp after cutting with *AfIII* which insert in GFP-vector at site 1618 bp as show in table (3) Figure (4-6) and (4-7) .Insertion of RFP-gene with CMV promoter may be with the same reading frame or in posit reading frame of GFP- vector genes as show in Figures (3 and 5).

Table (3) Nested PCR steps to created RFP –gene with linker of *AfIII*

1	CGG CGA CTT AAG TAATAGTAATCAATTACGGGGTCATTAGTTCATAGCCCATAATGGA GTTCCCGCTTACATAACTACGGTAAATGGCCCGCCTGGCTGACCGCC CAACGACCCCCGCCATTGACGTCAATAATGACGTAGTTCCATAGT AACGCCAATAGGGACTTCCATTGACGTCAATGGGTGGAGTATTTAC GGTAAACGCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTA CGCCCCCTATTGACGTCAATGACGGAAATGGCCCGCCTGGCATTATG CCCAGTACATGACCTTATGGGACTTCCCTACTTGGCAGTACATCTAC	1461 bp product from linear template RFP-CMV base 8 to base 1481(D2 F - D2 R). (1473) bp for
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	GTATTAGTCATCGCTATTACCATGGTGATGCGGTTTGGCAGTACAT CAATGGCGTGGATAGCGGTTGACTCACGGGATTCCAAGTCTCC ACCCCATTGACGTCAATGGAGTTGTTGGCACCAAAATCAACGG GACTTCCAAAATGTCGTAACAACACTCCGCCATTGACGCAAATGGG CGGTAGGCGTGTACGGTGGGAGGTCTATAAAGCAGAGCTGGTTA GTGAACCGTCAGATCCGCTAGCGCTACCGGTCGCCACCATGGTGAGC GAGCTGATTAAGGAGAACATGCCATGAAGCTGTACATGGAGGGCA CCGTGAACAACCACCACTCAAGTGCACATCCGAGGGCGAAGGCAA GCCCTACGAGGGCACCCAGACCATGAGAACATCAAGGTCGTCAGGGC GGCCCTCTCCCCCTCGCCCTCGACATCCTGGCTACCAGCTTCATGTAC GGCAGCAGAACCTCATCAAGCACCCCTCCGGCATCCCCGACTTCTT TAAGCAGTCCTCCCTGAGGGCTTCACATGGGAGAGAGTCACCACAT ACGAAGACGGGGCGTGCTGACCGCTACCCAGGACACCAGCCTCCA GGACGGCTGCCTCATACAACGTCAAGGTTAGAGGGTGAACCTCC CAGCCAACGGCCCTGTGATGCAGAACAAACACTCGGCTGGGAGGC CTCCACCGAGACGATGTACCCCGCTGACGGCGGCCACCTGATCTGCAA TGTGACATGGCCCTGAAGCTCGTGGCGGGGCCACCTGATCTGCAA CCTTGAGACCACATACAGATCCAAGAACCCCGCTACGAACCTCAAG ATGCCCGCGTCTACAACGTGGACACAGACTGGAAAGAACATCAAGG AGGCCGACGATGAGACCTACGTCGAGCAGCACGAGGTGGCTGTGGC CAGATACTACTGGTGGCGCTGGTATGGAGGTAAAGGTGGAGGA GGTCCGGACTCAGATCTGAGCTAAGCTCGAATTCTGCAGTCGA CGGTACCGCGGGCCGGATCCACCGGATCTAGATAACTGATCATA ATCAGCCATACCACATTGTAGAGGTTTACTGCTTAAAAAACCT CCCACACCTCCCTTAAGACATAG	D3 amplification and 1485 bp for D4 amplification
2	TTAAGTAATAGTAATCAATTACGGGTCATTAGTCATAGCCCATA ATGGAGTCCCGCTTACATAACTACGGTAAATGGCCCGCCTGGCTGA CCGCCAACGACCCCCGCCATTGACGTCAATAATGACGTAGTCCC ATAGTAACGCCAATAGGGACTTCCATTGACGTCAATGGGTGGAGTA TTTACGGTAAACGCCACTTGGCAGTACATCAAGTGTATCATATGCC AAAGTACGCCCTATTGACGTCAATGACGGAAATGGCCCGCCTGGCA TTATGCCAGTACATGACCTTATGGACTTCTACTTGGCAGTACAT CTACGTATTAGTCATCGCTATTACCATGGTATGGCTGGGTTTGGCAGT ACATCAATGGCGTGGATAGCGGTTGACTCACGGGATTCCAAGT CTCCACCCATTGACGTCAATGGAGTTGGCACCAAAATCA ACGGGACTTCCAAAATGCGTAACAACACTCCGCCATTGACGCAA TGGCGGTAGCGTGTACGGTGGGAGGTCTATAAAGCAGAGCTGG TTAGTGAACCGTCAGATCGCTAGCGTACCGGCCACCATGGT GAGCGAGCTGATTAAGGAGAACATGCCATGAAGCTGTACATGGAG GGCACCGTGAACAACCACCACTCAAGTGCACATCCGAGGGCGAAG GCAAGCCTACGAGGGCACCCAGACCATGAGAACATCAAGGTCGTCGA GGCGGCCCTCTCCCCCTCGCCCTCGACATCCTGGCTACCAGCTTCAT GTACGGCAGCAGAACCTCATCAAGCACCCCTCCGGCATCCCCGACT TCTTAAGCAGTCCTCCCTGAGGGCTTCACATGGGAGAGAGTCACC ACATACGAAGACGGGGCGTGCTGACCGCTACCCAGGACACCAGCC TCCAGGACGGCTGCTCATACAACGTCAAGGTTAGAGGGTGAACT TCCCAGCCAACGGCCCTGTGATGCAGAACAAACACTCGGCTGGGA GGCCTCCACCGAGACGATGTACCCCGCTGACGGCGGCCCTGGAGGC GCATGTGACATGGCCCTGAAGCTCGTGGCGGGGCCACCTGATCTG CAACCTTGAGACCACATACAGATCCAAGAACCCCGCTACGAACCTC AAGATGCCCGCGTCTACAACGTGGACACAGACTGGAAAGAACATCA AGGAGGCCACGATGAGACCTACGTCGAGCAGCACGAGGTGGCTGT GGCCAGATACTACTGGTGGCGCTGGTATGGAGGTAAAGGTGGA GGAGGTTCCGGACTCAGATCTGAGCTAAGCTCGAATTCTGCAGT CGACGGTACCGCGGGCCGGATCCACCGGATCTAGATAACTGATC	1463 bp linear fragment from linear parent Untitled, base 8 to base 1470 (AflIIc ttaag - AflIIc ttaag).

	ATAATCAGCCATACCACTTGTAGAGGTTTACTTGCTTAAAAAA CCTCCCACACCTCCC	
3	TTAAGACATAG	11 bp linear fragment from linear parent Untitled, base 1471 to base 1481 (AflIIc ttaag - sequence end). TTAAGACATAG
4	CGGCGAC	7 bp linear fragment from linear parent Untitled, base 1 to base 7 (sequence start - AflIIc ttaag)

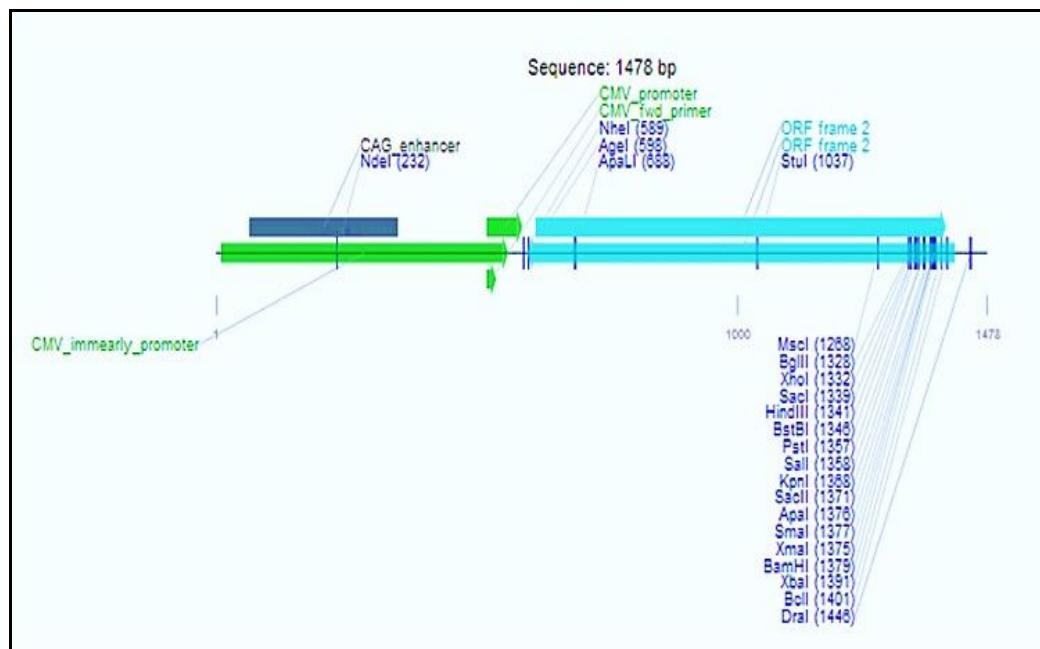


Figure (1) Physical map of RFP gene with CMV promoterin analysis sequence –addgene

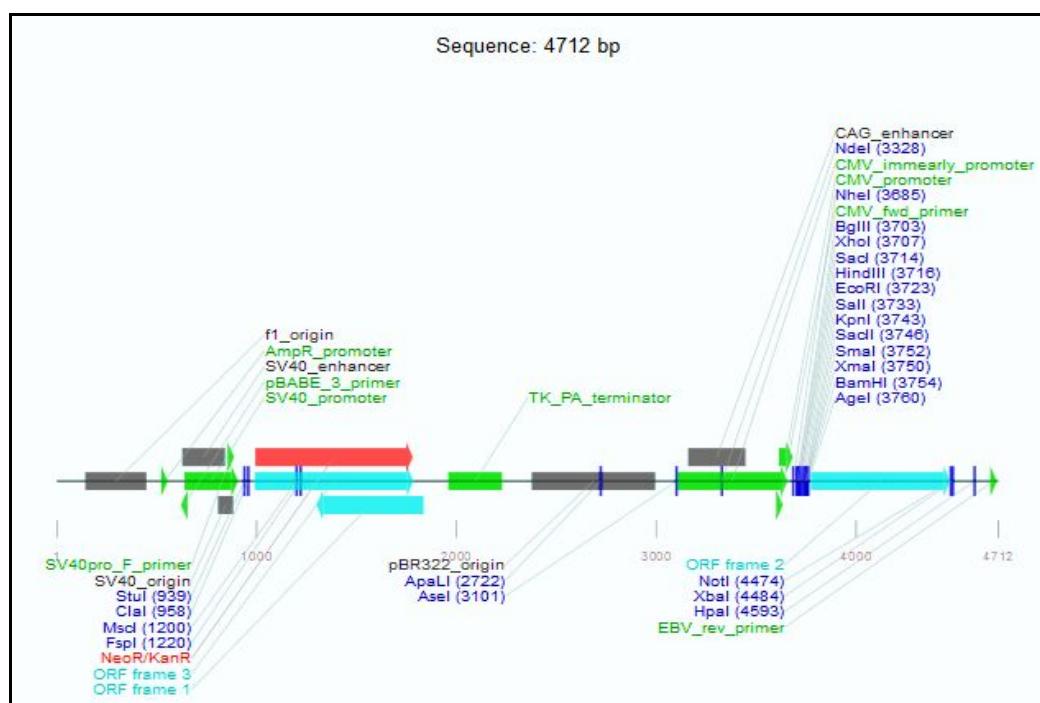


Figure (2) physical map of GFP- vector linearized using *AflIII* restriction enzymein analysis sequence –addgene

Table(3) The sequence of DII vector

TTAAGGCGTAAATTGTAAGCGTTAATATTGTTAAAATTGCGCTTAAATTGTTAAATCA GCTCATTTTAACCAATAGGCCAAATCGGCAAACCCCTATAAATCAAAAGAATAGAC CGAGATAGGGTTGAGTGTGTCAGTTGGAACAAGAGTCCACTATTAAAGAACGTGGAC TCCAACGTCAAAGGGCAGAAAACCCTATCAGGGCGATGGCCCCTACGTGAACCAC CCTAATCAAGTTTGCGAGGTGCCGAAAGCACTAAATCGAACCTAAAGGGAG CCCCGATTAGAGCTTGACGGGAAAGCCGGCAACGTGGCGAGAAAGGAAGGGAGAA AGCGAAAGGAGCGGGCGTAGGGCGCTGGCAAGTGTAGCGGTACGCTGCCGTAACCAC CACACCCGCCGCCTTAATGCGCCGCTACAGGGCGCTCAGGTGGACTTTGGGAAAT GTGCGCGGAACCCCTATTGTTATTCTAAATACATTCAAATATGTATCCGCTCATGAGA CAATAACCCCTGATAAATGCTCAATAATATTGAAAAAGGAAGAGTCCCTGAGGCGGAAAGAA CCAGCTGTGGAATGTGTGTCAGTTAGGGTGTGAAAGTCCCCAGGCTCCCCAGCAGGCAGA AGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCAGGTGTGAAAGTCCCCAGGCTCCC CAGCAGGCAGAAGTATGCAAAGCATGCATCTCAATTAGTCAGCAACCATACTCCGCCCC AACTCCGCCATCCGCCCTAACTCCGCCAGTCCGCCATTCTCCGCCATGGCTGAC TAATTGTTATTATGAGAGGCCAGGGCCCTGGCCCTGAGCTATTCCAGAAGTAG TGAGGAGGCTTTTGAGGCCTAGGCTTGCAAAGATCGATCAAGAGACAGGATGAGGA TCGTTCGCATGATTGAAACAAGATGGATTGCACGCAGGTTCTCCGCCGCTGGTGGAGA GGCTATTGGCTATGACTGGCACAACAGACAATCGGCTGCTGTGATGCCGCCGTGTCGG CTGTCAGCGCAGGGCGCCGGTTCTTGTCAAGACCGACCTGTCGGTCCCTGCGCAGCT ACTGCAAGACGAGGCAGCGCGCTATCGTGGCTGCCACGACGGCGTTCCCTGCGCAGCT GTGCTGACGTTGCACTGAAGCGGGAAAGGGACTGGCTGCTATTGGCGAAGTGCCGGGGC AGGATCTCCTGTCATCTCACCTGCTCCTGCCAGAAAAGTATCCATCATGGCTGATGCAATG CGGCCGGCTGCATACGCTTGATCCGGCTACCTGCCATTGACCCATTGACCCAGCAGAACATCGCA TCGAGCGAGCACGTACTCGGATGGAAGCCGGCTTGCGATCAGGATGATCTGGACGAAGA GCATCAGGGGCTCGGCCAGCGAACTGTCGCCAGGCTCAAGGCAGCATGCCGACGGC GAGGATCTCGTCGTGACCCATGGCGATGCCCTGCTTGCGAATATCATGGTGGAAATGGCC GCTTTCTGGATTATCGACTGTGGCCGGCTGGGTGTGGCGGACCGCTATCAGGACATAGCG TTGGCTACCGTGATATTGCTGAAGAGCTGGCGCGAATGGGCTGACCGCTTCCTGCT TTACGGTATGCCGCTCCGATTGCAAGCGCATGCCCTCTATGCCCTCTTGACGAGTTCT CTGAGCGGGACTCTGGGTTGAAATGACCGACCAAGCGACGCCAACCTGCCATACGAG ATTTCGATTCCACCGCCGCCCTCTATGAAAGGTTGGCTTGGCGAATCGTTCCGGACGCC GGCTGGATGATCTCCAGCGCGGGATCTCATGCTGGAGTTCTCGCCCACCCCTAGGGGA GGCTAACTGAAACACGGAAGGAGACAATACCGGAAGGAACCCCGCCTATGACGGCAATAA AAAGACAGAATAAAACGCACGGTGTGGCTGTTGTCATAAACCGGGGTTGGCTCCCA GGGCTGGCACTCTGCGATAACCCACCGAGACCCATTGGGCAATACGCCGCTTCTT CCTTTCCCACCCACCCCAAGTCGGGTGAAGGCCAGGGCTCGCAGCCAACGTCGG GGCGCAGGCCCTGCCATAGCCTCAGGTTACTCATATACTTAGATTGATTAAACTTC ATTTTAATTAAAAGGATCTAGGTGAAGATCCTTTGATAATCTCATGACCAAAATCCCT TAACGTGAGTTTCGTTCACTGAGCGTCAGACCCGTAGAAAAGATCAAAGGATCTTCTG AGATCCTTTCTGCGCGTAATCTGCTGCTGCAAACAAAAAACCCCGCTACCAGCGG TGGTTGTTGCCGGATCAAGAGCTACCAACTCTTCCGAAGGTAACTGGCTCAGCAGA GCGCAGATACCAAATACTGCTCTTAGTGTAGCCGTAGTTAGGCCACCACTCAAGAAC TGTAGCACCCTACATACCTCGCTCTGCTAATCCTGTTACCAGTGGCTGCTGCCAGTGGCG ATAAGTCGTGTTACCGGGTTGGACTCAAGACGATAGTTACCGGATAAGGCCAGCG GGGCTGAACGGGGGGTTCGTCACACAGCCCAGCTGGAGCGAACGACCTACACCGAACTG AGATACCTACAGCGTGAGCTATGAGAAAGCGCCACGCTCCGAAGGGAGAAAGCGG AGGTATCCGGTAAGCGGCAGGGTGGAAACAGGAGAGCGCACGAGGGAGCTCCAGGGGG AACGCCCTGGTATCTTATAGTCCTGTCGGTTGCCACCTCTGACTTGAGCGTCGATT TGATGCTCGTCAGGGGGCGGAGCCTATGGAAAAACGCCAGCAACGCCCTTTACGGT TCCTGGCCTTTGCTGGCCTTTGCTCACATGTTCTGCGTTATCCCTGATTCTGTTGGA TAACCGTATTACCGCCATGCAATTAGTTATAAGTAATCAATTACGGTAATGGCCCG CAACGACCCCCGCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAGGG
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ACTTTCCATTGACGTCAATGGGTGGAGTATTACGGTAAACTGCCACTTGGCAGTACATCA
AGTGTATCATATGCCAAGTACGCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGC
ATTATGCCAGTACATGACCTTATGGGACTTCCTACTTGGCAGTACATCTACGTATTA
GTCATCGCTATTACCATGGTATGCAGGTTTGGCAGTACATCAATGGCGTGGATAGCGGTT
TGACTCACGGGGATTCCAAGTCTCCACCCATTGACGTCAATGGGAGTTGTTGGCACC
AAAATCAACGGGACTTCCAAAATGTCGAACAACCTCCGCCATTGACGCAAATGGCGG
TAGGCAGTGTACGGTGGGAGGTCTATATAAAGCAGAGCTGGTTAGTGAACCGTCAGATCCGC
TAGCGCTACCGGACTCAGATCTCGAGCTCAAGCTCGAATTCTGCAGTCACGGTACCGCG
GGCCCGGGATCCACCGGTGCCACCATGGAGAGCGACGAGAGCGGCCCTGCCGCATGGA
GATCGAGTGCCGCATCACCAGCACCCCTGAACGGCGTGGAGTTCGAGCTGGTGGCGCGGA
GAGGGCACCCCCGAGCAGGGCCCATGACCAACAAGATGAAGAGCACCAAGGCACCC
ACCTTCAGCCCACCTGCTGAGCCACGTGATGGCTACGGCTTCTACCAACTCGGCACCTA
CCCCAGCGGCTACGAGAACCCCTCCTGCACGCCATCAACAACGGCGCTACACCAACACC
CGCATCGAGAAGTACGAGGACGGCGCGTGCACGTGAGCTTCAGCTACCGCTACGAGG
CCGGCCCGTGTACGGCAGCTCAAGGTGATGGGCACCGGCTCCCGAGGACAGCGTGT
CTTCACCGACAAGATCATCCGAGCAACGCCACCGTGGAGCACCTGCACCCATGGCGAT
AACGATCTGGATGGCAGCTTCACCGCACCTCAGCCTGCGCACGGCGTACTACAGCT
CCGTGGTGGACAGCCACATGCACTCAAGAGCGCATCCACCCAGCATCTGCAGAACGG
GGGCCCCATGTTGCCCTCCGCCGTGGAGGAGGATCACAGCAACACCGAGCTGGCATC
GTGGAGTACCAAGCACGCCCTCAAGACCCGGATGCAAGATGCCGTGAAGAATAAGCGGCC
GCGACTCTAGATCATAATCAGCCATACCACTTGTAGAGGTTTACTTGCTTAAAAAAC
TCCCACACCTCCCCCTGAACCTGAAACATAAAATGAATGCAATTGTTGTTAACTGTT
ATTGCAGCTTATAATGGTTACAAATAAGCAATAGCATCACAAATTTCACAAATAAGCAT
TTTTTCACTGCATTCTAGTTGTGGTTGTCACACTCATCAATGTATCTTAAGTAATAGTAA
TCAATTACGGGGTCATTAGTCATAGCCATATATGGAGTCCCGTGTACATAACTACGGT
AAATGGCCCGCCTGGCTGACCGCCCAACGACCCCCGCCATTGACGTCAATAATGACGTAT
GTTCCCATAGTAACGCCAATAGGGACTTCCATTGACGTCAATGGTGGAGTATTACGGTA
AACTGCCACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCTATTGACGTCA
ATGACGGTAAATGGCCCGCTGGCATTATGCCAGTACATGACCTATGGACTTCCACT
TGGCAGTACATCTACGTATTAGTCATCGCTATTACCATGGTATGCCGTGGCAGTACAT
CAATGGCGTGGATAGCGGTTGACTCACGGGATTCCAAGTCTCCACCCATTGACGTCA
ATGGGAGTTGTTGGCACCAAAATCAACGGACTTCCAAAATGCGTAACAACCTCGCC
CCATTGAGCAAATGGCGGTAGCGTGTACGGTGGAGGCTATATAAGCAGAGCTGGTT
AGTGAACCGTCAGATCCCTAGCGTACCGCTACCGGTGCCACCATGGTGGAGCAGTGA
AGAACATGCCATGAAGCTGTACATGGAGGGCACCGTGAACAACCACCACTCAAGTGCAC
ATCCGAGGGCGAAGGCAAGCCCTACGAGGGCACCCAGACCATGAGAATCAAGGTCGTCA
GGCGGCCCTCTCCCTCGCCTCGACATCCTGGCTACAGCTTGTACGGCAGCAGAA
CCTCATCAAGCACCCCTCGGCATCCCCACTTCAAGCAGTCTTCCAGCTGAGGGCTTC
ACATGGGAGAGAGTCACACATCGAAGACGGGGCGTGTGACCGCTACCCAGGACACC
AGCCTCCAGGACGGCTGCCTCATCTACACGTCAAGGTTAGAGGGGTGAACCTCCAGCCA
ACGGCCCTGTGTACGAGAAAACACTCGGCTGGAGGGCTCCACCGAGACGATGTACCC
CGCTGACGGCGGCCCTGGAAAGGCGCATGTGACATGGCCCTGAAGCTCGTGGCGGGGCCAC
CTGATCTGCAACCTTGAGACCACATACAGATCCAAGAAACCCGCTACGAACCTCAAGATGC
CCGGCGTCTACAACGTGGACCACAGACTGGAAAGAATCAAGGAGGCCACGGATGAGACCT
ACGTCGAGCAGCACGAGGGTGGCTGGCCAGATACTCTACTGGTGGCGCTGGTGTGGAGG
TAAAGGTGGAGGAGGTTCCGGACTCAGATCTCGAGCTCAAGCTCGAATTCTGCAGTCAC
GGTACCGCGGGCCGGATCCACCGATAGATAACTGATCATAATCAGCCATACCAACAT
TTGTAGAGGTTTACTTGCTTAAAAACCTCCACACCTCCGAATTGATACA

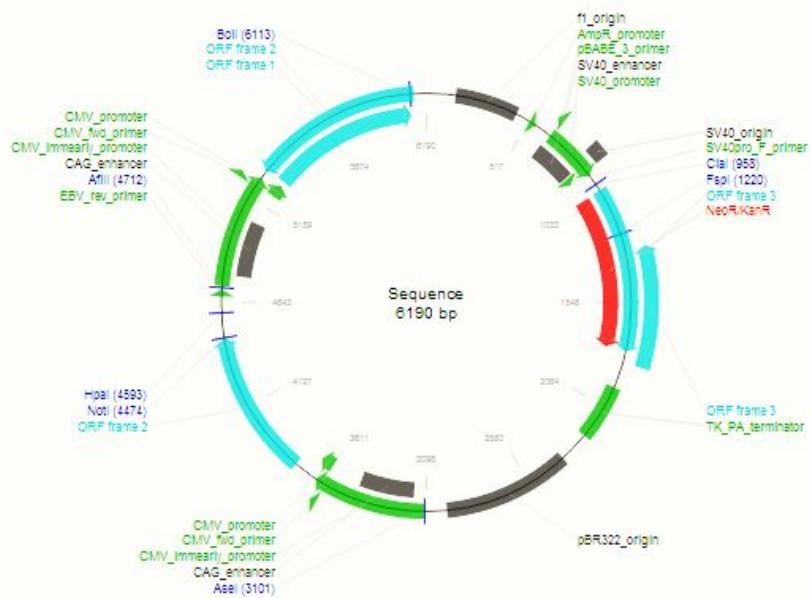


Figure (3) Physical map of DII vector show RFP- gene with the same direction of vector genesin analysis sequence –addgene.

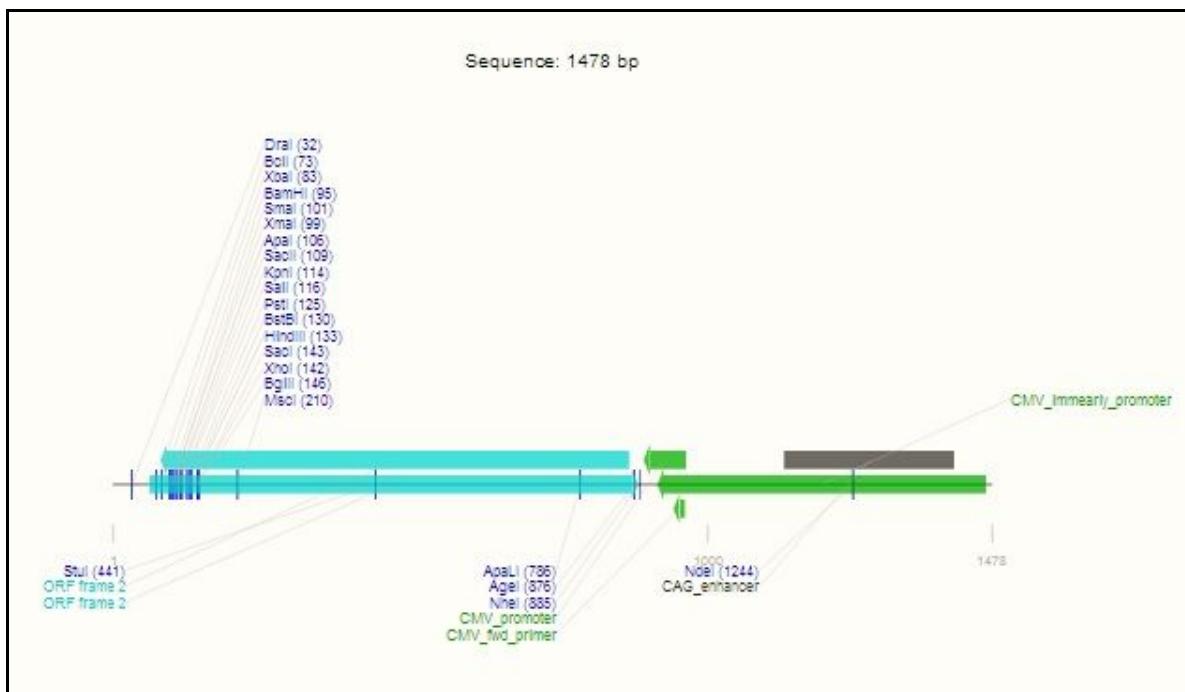


Figure (4) Physical map of DII vector in opposite direction of RFP-gene with CMV promoter in analysis sequence –addgene software.

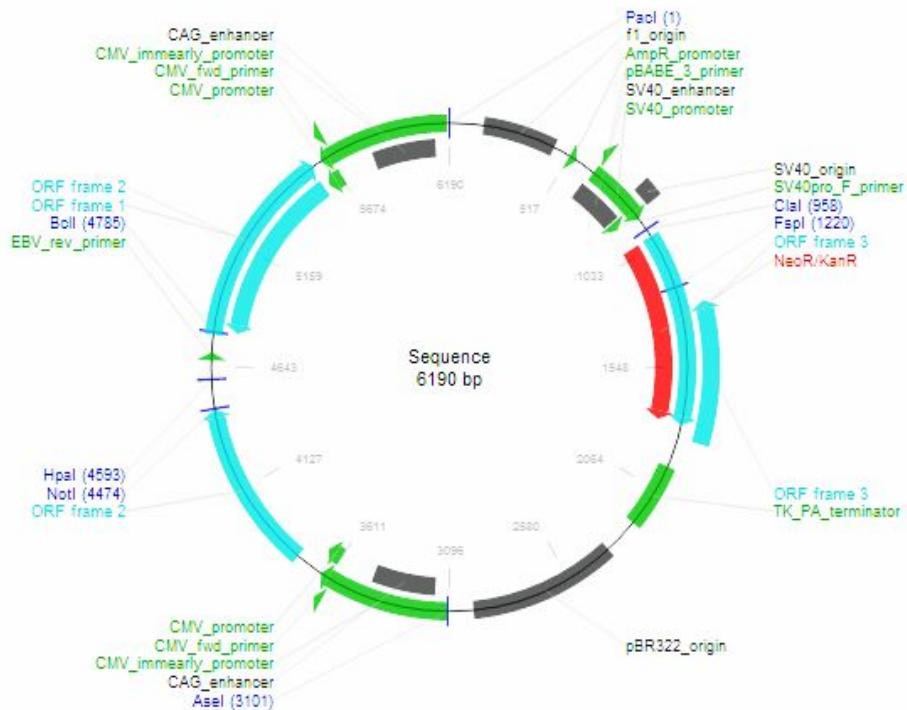


Figure (5) Physical map of DII vector when RFP- gene with CMV promoter insertion in opposite direction with the vector genes in analysis sequence – addgene software.

Table (4) Some features of DII vector with its positions

	SITE	
	From	to
<u>f1_origin</u>	448	142
<u>AmpR_promoter</u>	527	555
<u>pBABE_3_prime</u>	641	621
<u>SV40_enhancer</u>	842	627
<u>SV40_promoter</u>	639	907
<u>SV40_origin</u>	806	883
<u>SV40pro_F_primer</u>	868	887
<u>NeoR/KanR</u>	993	1781
<u>TK_PA_terminator</u>	1959	2228
<u>pBR322_origin</u>	2376	2995
<u>CMV_immearly_promoter</u>	3104	3656
<u>CAG_enhancer</u>	3159	3446
<u>CMV_fwd_primer</u>	3613	3633
<u>CMV_promoter</u>	3614	3683
<u>EBV_rev_primer</u>	4685	4704
<u>CMV_immearly_promoter</u>	4720	5271
<u>CAG_enhancer</u>	4775	5062
<u>CMV_promoter</u>	5229	5298

Table (5) Open reading frame position of DII vector

ORF frame 3	990	1784
ORF frame 2	1835	1299
ORF frame 2	3773	4471
ORF frame 2	6128	5307
ORF frame 3	5322	6110

Table (6) Confirmatory test sequence of DII using primer 1.

	TAATAGTAATCAATTACGGGGTCAATTAGTTCATAGCCCATAATATGGA GTTCCCGCGTTACATAACTTACGGTAAATGGCCCGCCTGGCTGACCGC CCAACGACCCCCGCCATTGACGTCAATAATGACGTATGTCCTCATA GTAACGCCAATAGGGACTTCCATTGACGTCAATGGGTGGAGTATT ACGGTAAACTGCCACTTGGCAGTACATCAAGTGTATCATATGCCAA GTACGCCCTATTGACGTCAATGACGGTAAATGGCCCGCCTGGCAT TATGCCAGTACATGACCTTATGGACTTCCTACTTGGCAGTACAT CTACGTATTAGTCATCGCTATTACCATGGTGATGCGGTTTGGCAGT ACATCAATGGCGTGGATAGCGGTTGACTCACGGGGATTCCAAGT CTCCACCCCATTGACGTCAATGGGAGTTGTTGGCACCAAATCA ACGGGACTTCCAAAATGTCGTAACAACACTCCGCCATTGACGCAA TGGGCGGTAGGCCTGTACGGTGGGAGGTCTATATAAGCAGAGCTGG TTTAGTGAACCGTCAGATCCGCTAGCGCTACCGGACTCAGATCTCGA GCTCAAGCTTGAATTCTGCAGTCAGCGTACCGGCTGGGAGCTGG CACCGGTCGCCACCATTGGAGAGCGACGAGAGCGGGCCTGCCGCCAT GGAGATCGAGTGCCGCATCACCAGCACCTGAACGGCGTGGAGTTC GAGCTGGTGGCGGGAGAGGGCACCCCGAGCAGGGCGCATG ACCAACAAGATGAAGAGCACCAAAGGCCTGACCTTCAGCCCCT ACCTGCTGAGCCACGTGATGGCTACGGCTTCTACCACTCGGCACC TACCCAGCGGCTACGAGAACCCCTTCCTGCACGCCATCAACAACG CGGGCTACACCAACACCGCATCGAGAACAGTACGAGGAGGGCG GCTGCACGTGAGCTTCAGCTACCGCTACGAGGCCGGCGGTGATC GGCGACTTCAAGGTGATGGGACCGGCTCCCGAGGACAGCGTGA TCTTCACCGACAAGATCATCGCAGCAACGCCACCGTGGAGCACCT GCACCCATGGCGATAACGATCTGGATGGCAGCTTCACCCGCACCT TCAGCCTGCGCACGGCGCTACTACAGCTCCGTGGAGCAGCCA CATGCACTTCAAGAGCGCCATCCACCCAGCATCTGCAGAACGGG GGCCCATGTTGCCCTTCGCCGTGGAGGAGGATCACAGCAACA CCGAGCTGGGATCGTGGAGTACCAAGCACGCCCTCAAGACCCGGA TGCAGATGCCGGTGAAGAATAAAAGCGGCCGCGACTCTAGATCATAA TCAGCCATACCACATTGTAGAGGTTTACTGCTTAAAAAACCTC CCACACCTCCCCCTGAACCTGAAACATAAAATGAATGCAATTGTTGT TGTAACTTGTATTGAGCTTATAATGGTTACAAATAAGCAATA GCATCACAAATTCAAAATAAGCATTTCCTACTGCATTCTAGTT GTGGTTGTCAAACACTCATCAATGTATCTTAAGTAATAGTAATCAAT TACGGGGTCAATTAGTCATAGCCATATATGGAGTCCCGCGTACAT AACTTACGGTAAATGGCCCGCCTGGCTGACCGCCAAACGACCCCG CCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGCCAATAG GGACTTCCATTGACGTCAATGGGTGGAGTATTACGGTAAACTGCC CACTTGGCAGTACATCAAGTGTATCATATGCCAAGTACGCCCTAT TGACGTCAATGACGGTAAATGGCCCGCCTGGCATTATGCCAGTACA TGACCTTATGGACTTCCTACTTGGCAGTACATCTACGTATTAGTC	3077 bp product from linear template Untitled, base 3102 to base 6178 (2 - 21).

ATCGCTATTACCATGGTGATGCGGTTTGGCAGTACATCAATGGGCG TGGATAGCGGTTGACTCACGGGGATTCCAAGTCTCCACCCCATTG ACGTCAATGGGAGTTGTTGGCACCAAAATCAACGGGACTTCCA AAATGTCGTAACAACCTCCGCCATTGACGCAAATGGCGGTAGGC GTGTACGGTGGGAGGTCTATATAAGCAGAGCTGGTTAGTGAACCGT CAGATCCGCTAGCGCTACCGGTCGCCACCAGGTGAGCGAGCTGATT AAGGAGAACATGCCATGAAGCTGTACATGGAGGGCACCGTGAACA ACCACCACTCAAGTGCACATCCGAGGGCGAAGGCAAGCCCTACGA GGGCACCCAGACCATGAGAATCAAGGTGTCGAGGGCGGCCCTCTC CCCTTCGCTTCGACATCCTGGTACCAAGCTTCATGTACGGCAGCAG AACCTTCATCAAGCACCCCTCCGGCATCCCCGACTTCTTAAGCACT CCTCCCTGAGGGCTTCACATGGGAGAGAGTCACCACATACGAAGA CGGGGGCGTGCTGACCGCTACCCAGGACACCAGCCTCCAGGACGGC TGCCTCATCTACAACGTCAAGGTTAGAGGGGTGAACCTCCCAGCCAA CGGCCCTGTATGCAGAAGAAAACACTCGGCTGGGAGGCCTCCACC GAGACGATGTACCCCGCTGACGGCGGCCCTGGAAGGCGCATGTGACA TGGCCCTGAAGCTCGTGGCGGGGCCACCTGATCTGCAACCTTGA GACCACATACAGATCCAAGAAACCCGCTACGAACCTCAAGATGCC GGCGTCTACAACGTGGACCACAGACTGGAAAGAATCAAGGAGGCCG ACGATGAGACCTACGTCGAGCAGCACGAGGTGGCTGTGGCCAGATA CTCTACTGGTGGCGCTGGTATGGAGGTAAAGGTGGAGGAGGTTCC GGACTCAGATCTCGAGCTCAAGCTCGAATTCTGCACTGACGGTAC CGCGGGCCCGGGATCCACCGGATCTAGATAACTGATCATAATCAGC CATACCACATTGTAGAGGTTTACTTGCTTAAAAAACCTCCCACA CCTCC	
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Discussion

The important part of vectors designs were primers design, (Table 4-1), some basics were dependent in it, most important was TM of annealing temperature, every primers set have approximate TM to employed accurate amplification it must be low than 68°C°. Also GC% must be low than 60%⁽⁸⁻¹⁰⁾ in present study two methods used in primer design, first software primer3 to design primer, forward primer when flanking sequences were dependent, while revers and linker that must be added to some revers primer manually as red sequence in Table (1), then every primer was pass in to software to make sure of its ability to accurate complement in amplification. Other company used special calculation of primer TM design it depended on salt concentration and pH of master mix buffer such as *biolab* stools website ⁽¹³⁾ (<http://tmcalculator.neb.com/#/>)

Results show that this technology useful to detect primer TM as show in tables (1) and to avoid nonspecific amplification, The differences of TM between different softwrer because of the database of software's were designed according to materials of master mix of commercial company which design it, thus different annealing temperature can obtain for the same sequence in different website, also it is affected by salt concentration, type of amplification (hot start, touchdown, common amplification), type of DNA polymerase have main roles in annealing temperature also (*Taqman*, long patch amplification, hot start and proofreading activity, also primer concentration was effected in annealing temperature (<https://www.neb.com/>)⁽¹⁴⁾).

In some primers; linker must be added to primer sequence in order to cutting target gene after amplification for suitable complement sequences to accurate insertion. Linker was added according to Dallas *et al.* (1998)⁽³⁾ and <https://www.addgene.org/plasmid-protocols/subcloning/> as in table(1) with modification, linker consist of 6 nucleotides followed by other 6 nucleotides for restriction enzyme activity according to *biolabs* (<https://www.neb.com/tools-and-resources/usage-guidelines/cleavageclose-to-the-end-of-dna-fragments>)⁽¹⁵⁻¹⁹⁾ some restriction enzyme need at least 3 nucleotides for high activity, the addition of these nucleotide dependent on GC percentage and TM calculation.

Other site can be used also but in present study reporter gene choses as a model for evaluate its expression in cells thus any site can be inactivated it or cause knock out gene must be discarded. The target

gene can be inserted in other site than multiple cloning site; from the lank vector (GFP) map free sequences were used for insertion it between genes in intragenic sequences that have restriction site, In present study DII design , *AfIII*site choose to insert CMV promoter with RFP gene as in figure (1 and 2)The DII were created to introduced new features in expression vector and for processing some problems with a new molecular biology productions, Molecular cloning technology development a new vectors with different properties for cloning applications, commercial company of molecular production produce vectors for variety researches applications like prokaryotic vectors, eukaryotic vectors, vector have specific gene, and basic vectors (blank) , Al-Terehi et al., and another researchers⁽²⁰⁻²⁴⁾ used MCS to inserted RFP gene using tow restriction enzyme.

All vectors have mutual elements such as origin of replication, promoters, reporter gene and marker gene, These vectors specific for expression in mammalian or prokaryotic cells, many researchers used it as blank vectors uses fluorescence protein as reporter gene or insert other genes for different applications^(9, 25-28).

References

1. Al-Terehi, M. ; Al-Saadi, A.H. K. Zaidan, H.;Al-shariafe. A.(2015) Theoretically Construction Expression Vector Consist of Two Reporter Gene as a Fusion Protein. Aust. J. Basic & Appl. Sci., 9(7): 586-595, 2015
2. Aljeboree A. M., Alshirifi A. N., and Alkaim A. F. (2014). "Kinetics and equilibrium study for the adsorption of textile dyes on coconut shell activated carbon." Arabian J. Chem. 10.1016/j.arabjc.2014.01.020.
3. Dallas, Q.; Jiang, G.; Sladek, M. (1998) Digestion of terminal restrictionendonuclease recognition site on PCR products, BioTechniqes 24, 582-584
4. David Benton, (1996) Bioinformatics - principles and potential of a new multidisciplinary tool
5. Dieffenbach, C.W., Lowe, T.M.J., Dveksler, G.S.,(1995) General Concepts forPCR Primer Design, in *PCR Primer, A Laboratory Manual*, Dieffenbach, C.W, and Dveksler, G.S., Ed., Cold Spring Harbor Laboratory Press, 133-155.
6. EvrogenpTurboGFP-N vector (2013) http://www.evrogen.com/products/vectors/pTurbo_GFPN/pTurbo_GFP-N.shtml
7. Evrogen, pFusionRed-C vector (2013). <http://www.evrogen.com/products/vectors/pFusionRedC/pFusionRed-C.shtml>
8. Haff,L.A. (1994)Improved quantitative PCR using nested primers. *Genome Res.* 3: 332-337
9. Kaufman J.R. (2000) Overview of Vector Design for Mammalian GeneExpression, Molecular Biotechnology, 16,151-161.
10. Luscombe, N. M. ;Greenbaum, D, and Gerstein, M. (2001). What is bioinformatics? An introduction and overview, Yearbook of Medical Informatics 83-100.
11. The European Molecular Biology Laboratory (EMBL) (2015)http://www.embl.de/pepcore/pepcore_services/cloning/pcr_strategy/primer_design/
12. The new England biolabsInc, restriction endonuclease, (2014) <https://www.neb.com/products/restriction-endonucleases/restriction-endonucleases>.
13. The nonprofit plasmid repository addgene, analysis sequence (2015) Error! Hyperlink reference not valid.
14. The sequence manipulation site , PCR primer states (2014).
15. The sequence manipulation site, PCR product (2014) Error! Hyperlink reference not valid.
16. The sequence manipulation site, restriction digested (2014).Error! Hyperlink reference not valid.
17. Aljebori, A. M. and A. N. Alshirifi (2012). "Effect of Different Parameters on the Adsorption of Textile Dye Maxilon Blue GRL from Aqueous Solution by Using White Marble." Asian journal of chemistry 24(12): 5813-5816.
18. Al-Saadi, A. H., K. I. Zaidan, et al. (2015). "Dental sex determination by multiplex PCR in iraqi samples." Research Journal of Pharmaceutical, Biological and Chemical Sciences 6(6): 1572-1577.
19. Al-Terehi, M., Al-Saadi, A. H., Al-Sherefi, A. N., and Zaidan, H. K. (2016). "Optimization Polyplex Stability in Different Glucose Concentrations." International Journal of ChemTech Research 9(3): 396-401.
20. Al-Terehi, M., A. H. Al-Saadi, et al. (2015). "Some herbal medicinal plants activity against Candida spp which resistance to antifungal drugs." International Journal of PharmTech Research 8(10): 146-150.

21. Al-Terehi, M., A. H. Al-Saadi, et al. (2015). "Some plants extracts synergism effects in pathogenic bacteria." International Journal of PharmTech Research 8(10): 158-165.
22. Al-Terehi, M., H. K. Zaidan, et al. (2015). "Effective of different factors on trace elements concentrations in Iraqi lactating mother'smilk." International Journal of PharmTech Research 8(10): 151-157.
23. Aljeboree, A. M. (2016). International Journal of ChemTech Research 9(3): 412-423.
24. Aljeboree A. M. (2015). Research Journal of Pharmaceutical, Biological and Chemical Sciences 6(4): 778-788.
25. Alqaragully, M. B., AL-Gubury, H. Y., Aljeboree, A. M., Karam, F. F., and Alkaim, A. F. (2015). Research Journal of Pharmaceutical, Biological and Chemical Sciences 6(5): 1287-1296.
26. Al-Terehi, M., , al-kilabi, I. H. AL-Mamoori, A. M. J. Al-Jboori, M. J., Al-Saadi, A. H., and Zaidan, H. K. (2016). International Journal of ChemTech Research 9(3): 407-411.
27. Al-Terehi, M., Al Saadi, A. H., Zaidan, H. K., and Al-Harbi, S. J. (2016). "Protective Effects of Glycyrrhiza glabra Plant Extract against Cyclophosphamide in Kidney and Liver Tissues in White Albino Rats." International Journal of ChemTech Research 9(3): 402-406.
28. Hadi Z. A., Aljeboree A. M. and Alkaim A. F. (2014). Int. J. Chem. Sci. 12(4): 1273-1288.
