



Gamma irradiation effect on some morphological and chemical characters of Sudani and Masri Roselle varieties

Al-Ansary A.M.F.¹, Nagwa R. Abd-El Hamied¹, M.E.S. Ottai¹
and R.A. El-Mergawi²

¹Department of Genetic and Cytology, National Research Centre, Dokki, Cairo, Egypt.

²Department of Botany, National Research Centre, Dokki, Cairo, Egypt.

Abstract: Roselle or Karkadaih (*Hibiscus sabdariffa* L.) is one of the most important medicinal plants used for various nutritional, medicinal and pharmaceutical purposes. Seeds of two Roselle varieties (Sudani and Masri) were sown for the plant parent generation, then the harvested seeds were exposed (in dry and in soaked seed categories) to gamma rays with different doses. The irradiated seeds were re-sown for M₂ and M₃ generations to evaluate the responsibility to gamma radiation. All the studied morphological characters indicated significant variability between varieties in the plant parent generation except the number of main branches/plant. The Sudani plant parents exhibited the higher values for all characters than of Masri variety. Moreover, in the M₂ and M₃ generations all the morphological characters were affected significantly by the seed categories (dry or soaked) radiation doses and their interaction as well as the varietal variation. Gamma rays improved the values of all morphological traits than of control. The dose of 40 Gray in the soaked seed category and 240 Gray in the dry seed category stimulated the highest trait values. Furthermore, genotypic and phenotypic coefficients of variation, broad sense heritability and expected genetic advance estimation presented higher values in M₂ than M₃ for most characters. On the other hand, the phytochemical screening of Roselle sepals showed greater acidity, anthocyanins, phenolics and antioxidant activity for Sudani than Masri variety. The responsibility of soaked seed category was more sensitive than of dry seed category. Total soluble solids and pH values had slight responsibility to gamma rays. Irradiation dose of 60 Gray in soaked seed category and 240 in dry seed category stimulated the highest values for most evaluated chemicals in Masry variety. While 20 Gray gave the highest sugars, anthocyanins and antioxidant activity and 60 Gray gave the maximum acidity and phenolics at the soaked seed category. No characterized doses effects were noticed in the dry seed category of Sudani variety. All the morphological and chemical results indicated that there is a store of genetic variability between the studied varieties that can be exploited for the improvement of Roselle yield through the selection and/or the hybridization between Sudani and Masri to produce a new variety that can share the valuable characters.

Key words: Roselle, *Hibiscus sabdariffa*, varieties, generations, Gamma irradiation, sepals phytochemical screening.

Introduction

Genus *Hibiscus* under Malvaceae family consists of about 300 species. More than half of them originated in the parts of central and eastern of Africa^{1,2}. *Hibiscus sabdariffa* L. also known as Roselle or

Karkadaih is one of the most important species of *Hibiscus* in Egypt. Roselle is best grown in tropical and sub-tropical regions^{3,4}.

Roselle is cultivated for its leaves, seeds and calyces. Nutritionally young leaves of Roselle contain nutrients such as phosphorus, calcium, magnesium and potassium⁵. The leaves are consumed as a green vegetable and prepared like spinach⁶. However, the seed of Roselle is a valuable food resource on account of its protein, calorie and also substantial amount of fiber and valuable micro-nutrients⁷. The seeds contain 17 to 20% edible fixed oil which is similar in its properties to cotton seed oil^{8,9,10}. On the other hand, the color extract from the dry calyces is rich in anthocyanin¹¹, amino acids, organic compounds, mineral salts¹² and source of vitamin C¹³. Calyces extract is also a potential source of natural colorant to replace red synthetic coloring agents for carbonated soft drinks, jams, juices, jellies, sauces, chutneys, wines, preserves and other acidic foods^{2,6,14}. In fact, some Roselle varieties/cultivars are identified according to calyces anthocyanin content. For example, the Sudani Roselle variety has dark red calyces, while the calyx has light red color in Masri variety^{9,10}.

In several countries, Roselle is also considered to be one of the most famous folk medicinal plants. Where, many chemical components present in Roselle have potential health benefits and support the ethno medicinal use of Roselle in promoting cardio-vascular health and preventing hypertension¹⁵, pyrexia and liver disorders¹⁶, microorganism growth limitation¹⁷, as well as a diuretic, digestive and sedative¹⁸. The red varieties of Roselle have antioxidant and cyclooxygenase inhibitory activity¹⁹. Also, Roselle enters in pharmaceutical and cosmetic industries²⁰.

On the other hand, irradiation induces several cytological, genetic²¹, morphogenetic²², biochemical²³ and physiological alteration in cell and tissues of plants^{24,25}. Gamma ray treatments to plants with high doses disturb the leaf gas-exchange, hormone balance, water exchange and enzyme activities^{26,27}. These effects include changes in the plant cellular structure and cell metabolism such as alteration in photosynthesis, dilation of membranes of thylakoids, modulation of the antioxidant systems and accumulation of phenol compounds. Irradiation has proven an adept mean of encouraging the expression of recessive genes and producing new genetic variations²⁸. Irradiation also been successfully used for mutation in breeding of various plants^{22,29,30}. Mutation induction is one approach for creating genetic variation in the plants³¹. The technology of mutation induction has become an established tool in plant breeding in order to supplement existing germplasm and to improve cultivars in specific traits. Improved varieties of many crops have been released to forms as a result of induced mutation which have been used directly as new cultivars or in cross breeding programs³². Also, gamma radiation significantly affected the plant active ingredient biochemical contents such tannins and phenols...etc.³³

The aim of the present work is to evaluate the response of some important breeding characters and sepal biochemical screening for Sudani and Masri Roselle varieties affected by different doses of gamma irradiation in dry and soaked categories.

Materials and Methods

Materials

Air dried seeds of two Roselle (*Hibiscus sabdariffa* L.) varieties; Sudani and Masri were obtained from the Genetics and Breeding of Medicinal and Aromatic Plants Group, Genetics and Cytology Dept., National Research Centre (NRC), Egypt.

Seed radiation

The seeds of each Sudani and Masri Roselle varieties were divided into two categories; dry category where the air dry seeds were exposed to radiation directly, and Soaked category where the seeds has been soaked in water for 10 hours before exposing to rays. The seeds were exposed to gamma irradiation under Gamma Cobalt 60 Apparatus at the Nuclear Research Centre. The applied doses were 20, 40, 60 and 80 Gray for the soaked seeds category, and 80, 160, 240, and 320 Gray for the dry seeds category. The doses rate of gamma rays was one Gray per 1.613 second. Unexposed seeds were used as control.

Cultivation method

Irradiated and non-irradiated (control) seeds were sown in a new reclaimed sandy land at Wadi El-Natroun Village, Behira Governorate on April for three successive summer seasons 2012-2014. A randomized complete block design with three replications was used. Each replicate had five lines 3.5 m length and 60 cm in between. The distance between hills was 40 cm and each hill was thinned at one plant. Normal agronomic recommended practices of Roselle growing were followed to obtain maximum yield. A representative random sample of 10 individual plants from each plot were selected for recording the data of five traits: plant height (PH) cm, number of main and total branches per plant (NMB and NTB, respectively), number of capsules per plant (NC) and air dry sepals weight per plant (DSW) g.

Statistical analysis

The general statistical procedures were applied using version 11 of SPSS software³⁴. The statistical procedures were practiced according to standard methods given by³⁵. The analysis of variance (ANOVA) and broad sense heritability (h^2b) were generally assigned according to³⁶. The phenotypic and genotypic coefficients of variance (PCV and GCV %) were computed according to³⁷. The expected genetic advance (GA %) was computed according to³⁸.

Phytochemical screening

Total anthocyanins

Total anthocyanins were determined according to³⁹, with some modifications. 0.1 g for each replicate, was added to 20 mL of methanol containing HCl (0.5%, v/v), homogenized for 3 min at 1500 rpm using a homogenizer Ultraturrax Turratec TE102E (Tecnal, Brazil), and held at 4 °C for 1 h in the darkness. The slurry was centrifuged at $17,600 \times g$ for 15 min at 4 °C. The absorbance of the supernatant was recorded at 515 nm. Total anthocyanins content was calculated using the extinction coefficient equal to $3.6 \times 10^4 \text{ mol}^{-1} \text{ m}^{-1}$. Total anthocyanin content was expressed as mg pelargonidin-3-glucoside eq. g^{-1} DW.

Water extraction

Powdered sepals were subjected to extraction with water. One gram of powdered sample was shaken with 100 ml of distilled water for 12 hrs using shaking incubator at room temperature. Solids were separated by centrifugation and filtration. Total soluble solids, pH, total titratable acidity, soluble sugars, total phenolics, antioxidant activity, were then determined in the extracts.

Total soluble solids, pH and total titratable acidity (TTA)

Total Soluble solids were determined by evaporating water from 20ml of water extract using oven at 80 °C. Determination of pH for water extract was carried out using pH meter. Total titratable acidity was determined by titrating 20 ml of water extract. Titration was carried out to pH 8.2 using a 0.1 mol L^{-1} NaOH solution according to⁴⁰. Results were expressed as mg citric acid /g of dry weight (DW).

Soluble sugars

Soluble sugars in water extract were determined by phenol-sulfuric acid method according to⁴¹.

Total phenolics

Total phenolics was determined using Folin-Ciocalteu's reagent according⁴². Briefly, 1 ml of the extract was mixed with 1.5 ml of deionised water followed by 0.25 ml of Folin-Ciocalteu's reagent and allowed to react for 6 min. Then, 2.5 ml of 7% sodium carbonate was added and allowed to stand for 1 hr, and then the absorption was measured colorimetrically at 765 nm. Total phenolic was expressed as mg gallic acid/g dry sample using standard curve of gallic acid solution.

Antioxidant activity

Antioxidant was determined using Diphenyl-2-picrylhydrazyl (DPPH) radical-scavenging activity according to the method of⁴³ with some modifications. The stock solution was prepared by dissolving 24 mg 1, 1-diphenyl-2-picrylhydrazyl (DPPH) with 100 ml methanol and then stored at -20°C until needed. The working solution was obtained by mixing 10 ml stock solution with 45 ml methanol to obtain an absorbance of 1.1 ± 0.02 units at 515 nm using the spectrophotometer. Water extracts (750 μl) were allowed to react with 1500 μl of the DPPH solution for 5 min in the dark. Then the absorbance was taken at 515 nm. The standard curve was linear between 25 and 800 μmol Trolox. Results are expressed in μmol Trolox /g DW. Additional dilution was needed if the DPPH value measured was over the linear range of the standard curve.

Results and Discussion

Data of five morphological characters of the plant parents generation (the first season) of Sudani and Masri Roselle varieties were recorded, analyzed and illustrated in Table (1).

Table 1. Analysis of variance for five quantitative characters of the plant parent generation of two Roselle varieties

Source of variance	Df	Plant height	No. main branches /plant	No. total branches /plant	No. Capsules /plant	Dry sepals weight /plant
Varieties	1	16245.00**	26.45	396.05**	3892.05*	426.43**
Replicates	9	264.89	4.72	16.45	527.38	39.46
Error	9	546.56	7.78	5.61	692.27	35.61

The traits of plant height (PH), number of total branches (NTB), number of capsules (NC) and dry sepals weight (DSW) per plant indicated significant differences between the studied Roselle varieties. While the varieties differed non-significantly in the trait of number of main branches per plant (NMB). The plant parents of Sudani variety exhibited the higher range and mean values for all characters than of Masri variety. The plant parents of both varieties presented considered coefficient of variance CV% (more than 10%) for all studied traits except PH trait in Sudani variety (Table, 2).

Table 2. Four statistical items of five quantitative characters for the plant parent generation of Masri and Sudani varieties

variety	Items	Plant height	No. main branches /plant	No. total branches /plant	No. Capsules/plant	Dry sepals weight /plant
Sudani	Range	170-220	9-18	20-32	57-118	21.95-40.25
	X	202.5	12.2	25.3	88.9	27.7
	SE	4.73	0.92	1.33	7.04	1.71
	CV%	7.38	23.76	16.67	25.03	19.50
Masri	Range	100-190	7-13	13-19	30-92	11.5-29.5
	X	145.5	9.9	16.4	61.0	18.48
	SE	7.67	0.64	0.66	8.51	2.14
	CV%	16.66	20.45	12.6	44.12	36.64

The significant differences of average value addition to the considered CV% values for the morphological characters between the plant parent generation of Sudani and Masri Roselle varieties indicated that there is a store of genetic variability that can be exploited for the improvement of Roselle yield and thus suggesting the possibility of evolving higher Roselle yield variants through proper selection². The ranges of PH trait (170-220 and 100-190 cm for Sudani and Masri, respectively) are near to that of^{2,4}. Meanwhile,^{44,45} found that Roselle plant is about 3.5 m tall.⁹ recorded average of 150 and 147 cm for the height of Sudani and Masri Roselle plants, respectively. However, the range of Masri NTB trait was (13-19) lower than (18.47-36.73) of⁴ and (22.22-24.5) of². Meanwhile, NTB range of Sudani variety (20-32) was nearest to⁴ and wider than². But,⁹ found average of 5.7 and 4.3 branches per plant for Sudani and Masri Roselle varieties. A great diversity

between the varieties was also recorded in both NC and DSW traits agree with the finding of^{2,46,47}. This diversity indicated the possibility to increase calyx fruit production (quantity and weight) through selection.

Data of the above studied morphological characters (after radiation exposing) for M₂ and M₃ generations were analyzed and shown in Table (3). From this table, each of seed categories, radiation dose and their interaction affected significantly in all traits for both M₂ and M₃ generations. Also, NMB presented significant differences between varieties in both generations. The varieties affected significantly by radiation for NTB in M₂, and PH in M₃. The varieties interacted significantly with seed categories for NC and DSW in M₂ as well as NMB and NC for M₃. Meanwhile, the relationship of varieties with radiation doses was significant only for NTB in M₂ and NMB in M₃. Furthermore, the triple relationship among varieties, seed categories and irradiation doses had non-significant interaction in all traits for both generations except PH in M₂ which was significant (Table, 3).

Table 3. Analysis of variance for five quantitative characters of two generations (M₂ and M₃) of two Roselle varieties treated with five radiation doses for two radiation seed categories

G.	Source of variance	Df	Plant height	No. main branches /plant	No. total branches /plant	No. Capsules /plant	Dry sepals weight /plant
M ₂	Replicates	2	3.63	2.23	15.05	31.85	3.73
	Varieties (V)	1	79.3	16.03*	1530.15**	163.35	19.66
	Radiation Category (C)	1	9101.97**	673.37**	728.02**	2394.02**	290.01**
	Dose (D)	4	5303.45**	306.65**	1529.86**	13607.69**	1665.04**
	V x C	1	0.06	1.33	0.41	904.81**	110.65**
	V x D	4	51.16	2.56	159.11**	22.39	1.77
	C x D	4	764.32**	46.89**	258.23**	3055.47**	372.98**
	V x C x D	4	119.22*	0.98	17.04	219.61	28.50
Residual	38	41.69	2.86	10.93	96.59	11.73	
M ₃	Replicates	2	9.05	1.40	1.25	42.02	8.63
	Varieties (V)	1	7526.40**	228.15**	36.82	109.35	18.60
	Radiation Category (C)	1	504.60**	36.82**	331.35**	770.41**	78.44*
	Dose (D)	4	6120.71**	268.39**	3105.23**	14174.23**	1726.78**
	V x C	1	106.67	25.35*	14.01	442.82*	46.45
	V x D	4	99.69	17.86**	16.32	107.14	13.76
	C x D	4	699.14**	40.77**	496.27**	1513.54**	176.31**
	V x C x D	4	61.29	2.31	11.18	86.53	13.36
Residual	38	56.31	4.51	40.95	99.12	12.14	

Mean values of the five morphological traits of M₂ and M₃ generations for Masri and Sudani Roselle varieties treated with five radiation doses in a soaked seed category are shown in Table (4). The table shows that all gamma radiation doses stimulated the higher values than that of control (untreated plants) for all characters, generations and varieties to prove that all used doses improved the morphological traits of Roselle plants but with different responses. The dose of 40 Gray stimulated the highest morphological traits value for both generations and varieties. The highest radiation dose (80 Gray) stimulated the lowest morphological traits value comparing with the other doses. Generally, doses arrangement was 40, 60, 20 and 80 Gray, respectively related to the morphological traits improving. On the other hand, the plants of M₃ generation gave the higher morphological traits values than those of M₂ generation. Sudai was the better variety for all characters than Masri variety at M₂, while inverse pattern was noticed for PH, NMB and NTB of M₃ generation.

The same result pattern was noticed in the dry seed category (Table, 5). Gamma radiation improved the values for all studied traits in both generations and varieties than control. The dose of 240 Gray gave the highest trait value more than 160, 320 and 80 Gray, respectively. The plants of M₃ generation had the higher mean value in all traits than those of M₂ generation, except the mean value of NMB in Masri variety addition to PH and NMB traits of Sudani variety which had higher values in M₂ plants for all radiation doses.

Table 4. Mean value of five morphological traits of M₂ and M₃ generations for Masri and Sudani Roselle varieties treated with five radiation doses in a soaked seed category

Traits	Radiation doses	M ₂ generation		M ₃ generation	
		Masri variety	Sudani variety	Masri variety	Sudani variety
Plant height (cm)	Control	118.3 ± 4.41	111.3 ± 3.67	147.7 ± 6.23	139.7 ± 2.60
	20 Gy	144.0 ± 2.0	150.3 ± 2.19	190.0 ± 2.89	165.0 ± 2.89
	40 Gy	168.3 ± 4.41	183.0 ± 4.58	223.3 ± 9.28	203.3 ± 3.33
	60 Gy	153.7 ± 3.76	160.0 ± 1.15	207.7 ± 5.36	176.3 ± 2.73
	80 Gy	132.0 ± 1.15	123.3 ± 3.76	172.7 ± 2.67	158.3 ± 1.67
Number of main branches/plant	Control	8.3 ± 0.33	8.3 ± 0.88	12.0 ± 0.58	11.0 ± 1.53
	20 Gy	14.3 ± 0.33	15.3 ± 0.33	17.3 ± 0.33	15.3 ± 0.33
	40 Gy	18.7 ± 1.20	22.0 ± 0.58	27.3 ± 1.20	24.3 ± 3.93
	60 Gy	15.3 ± 0.33	17.3 ± 0.88	23.0 ± 1.00	17.0 ± 0.00
	80 Gy	12.3 ± 1.67	12.7 ± 0.67	15.0 ± 0.58	14.0 ± 0.58
Number of total branches/plant	Control	13.7 ± 0.88	17.3 ± 0.67	30.3 ± 3.84	30.3 ± 1.86
	20 Gy	20.3 ± 0.88	26.3 ± 2.85	61.3 ± 2.19	58.0 ± 2.31
	40 Gy	30.0 ± 2.00	48.0 ± 2.00	83.3 ± 1.20	78.0 ± 2.89
	60 Gy	23.3 ± 0.33	41.3 ± 4.18	70.0 ± 2.08	65.7 ± 0.88
	80 Gy	17.7 ± 0.67	21.7 ± 0.33	48.3 ± 3.38	48.7 ± 1.45
Number of capsules/plant	Control	42.0 ± 3.61	59.7 ± 4.37	72.3 ± 11.05	61.3 ± 1.33
	20 Gy	73.3 ± 1.86	93.0 ± 0.58	116.3 ± 1.20	120.3 ± 4.26
	40 Gy	120.0 ± 3.46	122.7 ± 10.4	163.0 ± 2.52	170.0 ± 0.00
	60 Gy	98.0 ± 1.15	99.0 ± 1.53	148.7 ± 9.84	151.3 ± 3.76
	80 Gy	57.7 ± 4.33	72.0 ± 2.52	106.0 ± 4.73	89.7 ± 5.17
Dry sepals weight (g)	Control	14.7 ± 1.26	20.9 ± 1.53	25.3 ± 3.87	21.5 ± 0.45
	20 Gy	25.7 ± 0.68	32.6 ± 0.20	39.1 ± 2.04	42.2 ± 1.39
	40 Gy	42.0 ± 1.18	42.9 ± 3.64	57.0 ± 0.89	59.2 ± 0.15
	60 Gy	34.4 ± 0.46	34.7 ± 0.52	52.0 ± 3.44	52.9 ± 1.30
	80 Gy	20.2 ± 1.53	25.2 ± 0.89	37.1 ± 1.65	31.5 ± 1.87

Table 5. Mean value of five morphological traits of M₂ and M₃ generations for Masri and Sudani Roselle varieties treated with five radiation doses in a dry seed category

Traits	Radiation doses	M ₂ generation		M ₃ generation	
		Masri variety	Sudani variety	Masri variety	Sudani variety
Plant height (cm)	Control	142.0 ± 7.51	147.0 ± 3.79	154.3 ± 6.36	136.0 ± 4.93
	80 Gy	155.7 ± 1.20	154.3 ± 0.33	173.0 ± 1.53	150.0 ± 2.89
	160 Gy	184.3 ± 2.96	182.0 ± 4.16	201.7 ± 4.41	170.0 ± 2.89
	240 Gy	193.0 ± 1.53	199.7 ± 5.49	211.7 ± 1.67	187.0 ± 5.86
	320 Gy	164.7 ± 2.91	168.0 ± 3.61	185.0 ± 2.89	157.3 ± 2.33
Number of main branches/plant	Control	12.0 ± 1.73	12.0 ± 0.58	12.0 ± 0.58	8.7 ± 0.88
	80 Gy	18.7 ± 1.33	18.7 ± 0.33	14.0 ± 0.00	12.3 ± 0.67
	160 Gy	24.0 ± 1.00	25.0 ± 0.58	22.3 ± 0.88	15.3 ± 0.33
	240 Gy	27.7 ± 1.67	29.3 ± 1.45	26.7 ± 2.19	17.3 ± 0.33
	320 Gy	21.7 ± 0.33	22.7 ± 0.88	18.3 ± 0.33	13.7 ± 0.33
Number of total branches/plant	Control	14.7 ± 2.33	16.7 ± 1.67	37.0 ± 3.51	33.3 ± 3.18
	80 Gy	22.7 ± 0.33	27.3 ± 2.67	42.3 ± 0.67	45.0 ± 3.00
	160 Gy	33.0 ± 2.52	45.3 ± 1.20	60.0 ± 1.53	59.3 ± 2.19
	240 Gy	42.3 ± 2.73	64.7 ± 1.33	77.0 ± 9.50	72.7 ± 4.37
	320 Gy	26.3 ± 1.20	36.3 ± 2.03	48.7 ± 1.76	51.7 ± 1.20
Number of capsules/plant	Control	50.3 ± 8.41	39.0 ± 3.79	73.3 ± 9.77	76.0 ± 4.16
	80 Gy	77.7 ± 3.84	65.3 ± 6.77	93.7 ± 1.20	97.7 ± 0.67
	160 Gy	109.7 ± 4.70	119.7 ± 11.14	120.3 ± 7.31	134.3 ± 5.33
	240 Gy	162.7 ± 9.33	162.0 ± 4.62	154.3 ± 7.88	162.7 ± 3.84
	320 Gy	92.7 ± 4.26	84.7 ± 4.37	101.7 ± 2.40	113.3 ± 7.26
Dry sepals weight (g)	Control	17.7 ± 2.89	13.7 ± 1.31	25.9 ± 3.39	26.6 ± 1.47
	80 Gy	27.2 ± 1.33	22.8 ± 2.35	32.9 ± 0.30	34.2 ± 0.24
	160 Gy	38.3 ± 1.60	41.9 ± 3.89	42.1 ± 2.53	47.1 ± 1.54
	240 Gy	56.9 ± 3.27	56.7 ± 1.62	53.9 ± 2.71	57.0 ± 1.33
	320 Gy	32.5 ± 1.49	29.6 ± 1.53	35.5 ± 0.76	39.7 ± 2.39

These results proved that all the used gamma doses enhanced (with different responses) the traits of Roselle plants comparing with the control plants. This stimulatory effect of gamma rays is due to the fact that mutagens stimulate the role of enzyme and growth hormone responsible for growth and yield, in addition to stimulate the cell division, alteration of metabolic processes that affect synthesis of nucleic acids^{22, 27, 48}. Improvement of Roselle plant characters as a result of gamma radiation was recorded by⁴⁹ in cotton plant as well as^{50, 51, 52} in okra plant. The superior gamma dose in each category (40 and 240 Gray in the soaked and dry seed categories, respectively) stimulated the maximum values for all studied traits. ⁵³ assumed the stimulation of gamma radiation to its impact on the auxins balance within the plant tissues. The dose of 80 Gray was the weaken dose compared with the other gamma doses might be due to reduced mitotic division in meristematic tissues and reduced moisture content^{27, 54, 55, 56}. ²⁶ suggested that gamma radiation disturb the leaf gas-exchange, hormone balance, water exchange and enzyme activities. These effects include changes in the plant cellular structure and cell metabolism such as alteration in photosynthesis, dilation of membranes of thylakoids, modulation of the antioxidant systems and accumulation of phenolic compounds. Generally, the studied characters were beter enhanced in M₃ plants than in M₂ plants to confirm that the Roselle plant can be improved and enhanced its income through selection. However, Sudani was the best variety in M₂ generation, while Masri was the best one for PH, NMB and NTB traits in M₃ generation. These results can be utilized for improving the Roselle plant in Egypt through the hybridization between Sudani and Masri to produce a new variety that could share the valuable characters.

On the other hand, Table (6) shows the assessed genotypic and phenotypic coefficients of variation (GCV% and PCV%, respectively) as well as broad sense heritability (h^2_b) and expected genetic advance (GA%) for the five studied traits of M₂ and M₃ generations for Masri and Sudani Roselle varieties exposed to different gamma radiation doses in case of soaked seeds. Both varieties presented greater GCV% in M₂ than M₃ generation for all traits except NC and DSW of Sudani variety affected by 20 and 80 Gray and PH of Masri variety affected by 80 Gray. However, the dose of 60 Gray stimulated the highest GCV% (9.98 and 10.55%) for NMB at M₂ of Masri and Sudani, respectively. While, the lowest GCV% value (1.62 and 1.57%) related to PH character for Masri M₂ and Sudani M₃, respectively. Higher PCV% was also assessed for M₂ than M₃ generation for both varieties in all traits and doses, except for Sudani NMB exhibited low PCV% affected by 20, 60 and 80 Gray addition to Masri DSW affected by 40 Gray. The greatest PCV% was affected by 80 Gray and related to NMB (25.23%) of Masri M₂ and (24.74%) of Sudani M₃. While the lowest PCV% (5.89 and 6.24% for Masri and Sudani, respectively) was exhibited for PH affected by 40 Gray in the M₃ generation (Table 6).

Exposing to 40 Gray of gamma rays produced the highest h^2_b for PH, NMB and NTB traits (17.68, 35.51 and 37.16%, respectively) in Sudani M₂ and (14.57, 29.82 and 36.84%, for the same respecting) in Masri M₃ generation. The maximum h^2_b for NC and DSW traits (38.24 and 38.24%, respectively) were obtained by 40 Gray in Masri M₂ and (37.15 and 36.89%, respectively) for Sudani M₃. The lowest h^2_b values were obtained affecting by 80 Gray for PH, NC and DSW (3.72, 11.07 and 11.07%, respectively) in Masri M₂ and for PH, NTB, NC and DSW (3.47, 7.83, 6.43 and 6.40%, respectively), in Sudani M₂. The lowest h^2_b of NMB was exhibited by exposing to 80 Gray in M₃ of both Masri (7.69%) and Sudani (8.33%), addition to 3.39% of the lowest h^2_b caused by 60 Gray in Masri M₂. Furthermore, higher GA values were recorded in M₃ generation for Masri PH, NTB and NC and Sudani NTB, NC and DSW, while Sudani PH and NMB presented the higher GA at M₂ generation. The dose of 40 Gray gamma rays stimulated the highest GA for Masri NC and DSW (6.50 and 3.84%, respectively) in M₂ and for PH, NMB and NTB (3.95, 2.54 and 5.26%, respectively) in M₃. Same stimulation was recorded for Sudani PH and NMB (4.23 and 2.62%) at M₂ and NTB, NC and DSW (4.82, 7.56 and 4.44%, respectively) at M₃ generation. Gamma dose 80 Gray exhibited the lowest GA values for all traits of both varieties and both generations (Table 6).

The resulted genetic parameters for the irradiated dry seeds of both varieties are shown in Table (7). Both varieties presented higher GCV% in M₂ than M₃ generation for NTB, NC and DSW affected by all gamma radiation doses. Meanwhile Sudani PH and NMB had the lower GCV% in M₂ affected by all gamma radiation doses. The dose of 80 and 320 Gray for PH trait and 240 Gray for NMB of Masri variety exhibited the higher GCV% in M₃ generation. The trait of NMB had the highest GCV% (8.33%) for Masri affected by 160 Gray in M₂ and (9.79%) for Sudani affected by 240 Gray in M₃ generation. While, the lowest GCV% was exhibited by the lowest dose (80 Gray) for PH of both Masri and Sudani (1.37 and 1.01%, respectively) in M₂ generation. Higher PCV% was assessed for Masri PH and NTB affected by all gamma doses in M₂. While higher PCV% was obtained for Masri NMB affected by all gamma doses (except by 240 Gray) as well as NC and DSW

(except by 80 Gray) in M₃ generation. However, Sudani PH and NMB had the higher PCV% affected by all gamma doses in M₃. Higher PCV% was obtained for Sudani NTB affected by all radiation doses (except by 240 Gray) as well as NC and DSW (except by 160 and 240 Gray) in M₂ generation. The maximum PCV% was obtained by 80 Gray in NMB trait of both Masri (25.42%) and Sudani (25.58%) in M₃ generation. Meanwhile NC trait showed the lowest PCV% (5.76 and 5.52% affected by 240 Gray for both Masri and Sudani, respectively at M₂ generation.)

Table 6. Four genetic items estimated for five quantitative characters of M₂ and M₃ generations for Masri and Sudani Roselle varieties treated with four radiation doses in a soaked seed category

Traits	variety	Item	M ₂ generations				M ₃ generations			
			20G	40G	60G	80G	20G	40G	60G	80G
Plant height (cm)	Masri	GCV	2.03	2.43	2.23	1.62	1.98	2.25	2.15	1.67
		PCV	7.82	6.90	7.42	8.40	6.69	5.89	6.23	7.23
		h ² _b	6.75	12.35	9.07	3.72	8.71	14.57	11.93	5.34
		GA	1.57	2.96	2.13	0.85	2.28	3.95	3.18	1.37
	Sudani	GCV	2.39	2.67	2.52	1.62	1.76	2.26	1.98	1.57
		PCV	7.41	6.35	7.06	8.71	7.38	6.24	6.99	7.63
		h ² _b	10.39	17.68	12.73	3.47	5.69	13.18	8.03	4.25
		GA	2.38	4.23	2.96	0.77	1.43	3.44	2.04	1.06
Number of main branches /plant	Masri	GCV	9.89	9.96	9.98	9.38	7.69	8.27	8.33	6.67
		PCV	22.44	18.35	21.31	25.23	21.45	15.15	17.21	24.04
		h ² _b	19.42	29.48	21.92	13.81	12.85	29.82	23.42	7.69
		GA	1.28	2.08	1.47	0.88	0.98	2.54	1.91	0.57
	Sudani	GCV	9.98	9.72	10.55	9.55	7.82	8.66	8.32	7.14
		PCV	21.31	16.31	19.71	24.61	23.04	16.17	21.21	24.74
		h ² _b	21.92	35.51	28.63	15.05	11.50	28.71	15.38	8.33
		GA	1.47	2.62	2.01	0.97	0.84	2.32	1.14	0.59
Number of total branches /plant	Masri	GCV	7.31	7.77	7.68	6.52	5.24	5.05	5.20	5.07
		PCV	19.64	14.58	17.64	21.90	10.40	8.31	9.43	12.47
		h ² _b	13.84	28.38	18.93	8.85	25.42	36.84	3.39	16.53
		GA	1.14	2.56	1.60	0.71	3.34	5.26	4.13	2.05
	Sudani	GCV	6.59	6.66	6.85	5.59	5.24	5.11	5.23	5.08
		PCV	17.13	10.93	12.18	19.97	10.84	8.71	9.88	12.39
		h ² _b	14.78	37.16	31.62	7.83	23.35	34.42	28.03	16.83
		GA	1.37	4.02	3.28	0.70	3.02	4.82	3.75	2.09
Number of capsules /plant	Masri	GCV	4.41	4.25	4.41	3.96	3.29	3.37	3.39	3.16
		PCV	9.88	6.87	7.95	11.91	8.02	6.21	6.65	8.62
		h ² _b	19.89	38.24	30.77	11.07	16.87	29.48	26.05	13.44
		GA	2.97	6.50	4.94	1.57	3.24	6.15	5.31	2.53
	Sudani	GCV	3.58	3.73	3.66	2.81	3.69	3.54	3.62	3.43
		PCV	9.05	7.32	8.62	11.09	7.48	5.81	6.32	9.38
		h ² _b	15.68	26.02	17.99	6.43	24.29	37.15	32.86	13.38
		GA	2.72	4.82	3.16	1.06	4.50	7.56	6.47	2.32
Dry sepals weight /plant (g)	Masri	GCV	7.45	7.18	7.45	6.70	5.49	5.70	5.74	5.34
		PCV	16.68	11.62	13.41	20.13	13.98	14.42	11.24	14.57
		h ² _b	19.98	38.24	30.89	11.07	15.38	15.64	26.02	13.45
		GA	1.76	3.84	2.93	0.93	1.73	2.65	3.14	1.50
	Sudani	GCV	6.06	6.31	6.18	4.75	6.22	5.99	6.12	5.79
		PCV	15.28	12.39	14.55	18.75	12.63	9.86	10.69	15.82
		h ² _b	15.73	25.97	18.04	6.40	24.30	36.89	32.75	13.41
		GA	1.61	2.84	1.88	0.62	2.67	4.44	3.81	1.38

GCV = Genotypic coefficient of variation, PCV = Phenotypic coefficient of variation, h²_b = Broad sense heritability%, GA = Genetic advance

Table 7. Four genetic items estimated for five quantitative characters of M₂ and M₃ generations for Masri and Sudani Roselle varieties treated with four radiation doses in a dry seed category

Traits	variety	Item	M ₂ generations				M ₃ generations			
			80G	160G	240G	320G	80G	160G	240G	320G
Plant height (cm)	Masri	GCV	1.37	2.04	2.14	1.67	1.44	1.97	2.07	1.73
		PCV	7.78	6.78	6.53	7.43	7.32	6.47	6.22	6.93
		h ² _b	3.12	9.03	10.69	5.06	3.88	9.29	11.03	6.22
		GA	0.78	2.32	2.78	1.28	1.01	2.50	2.99	1.64
	Sudani	GCV	1.01	1.88	2.10	1.57	1.44	1.98	2.20	1.69
		PCV	7.92	6.92	6.42	7.59	7.91	7.14	6.61	7.60
		h ² _b	1.63	7.35	10.68	4.55	3.32	7.69	11.11	4.96
		GA	0.41	1.91	2.82	1.16	0.81	1.92	2.83	1.22
Number of main branches /plant	Masri	GCV	7.99	8.33	8.26	8.28	5.85	8.31	8.29	7.92
		PCV	20.17	16.67	14.99	17.98	25.42	17.61	15.40	20.52
		h ² _b	15.67	25.00	30.35	21.21	5.29	22.23	28.99	14.89
		GA	1.22	2.06	2.60	1.70	0.39	1.80	2.46	1.15
	Sudani	GCV	7.99	8.32	8.20	8.32	8.91	9.69	9.79	9.43
		PCV	20.17	16.16	14.39	17.38	25.58	21.58	19.66	23.51
		h ² _b	15.67	26.52	32.47	22.93	12.12	20.18	24.81	16.10
		GA	1.22	2.21	2.82	1.86	0.79	1.37	1.74	1.07
Number of total branches /plant	Masri	GCV	7.20	7.48	7.17	7.48	3.15	4.62	4.74	4.06
		PCV	18.36	13.82	11.56	16.39	14.72	11.14	9.21	13.13
		h ² _b	15.37	29.33	38.49	20.84	4.57	17.17	26.49	9.54
		GA	1.32	2.73	3.88	1.85	0.59	2.36	3.87	1.26
	Sudani	GCV	6.88	6.81	6.18	7.04	4.39	4.97	4.98	4.79
		PCV	16.48	11.31	8.84	13.28	13.55	10.92	9.37	12.15
		h ² _b	17.45	36.33	48.93	28.11	10.48	20.66	28.28	15.55
		GA	1.62	3.83	5.76	2.79	1.32	2.76	3.97	2.01
Number of capsules /plant	Masri	GCV	3.89	4.06	3.76	4.06	2.78	3.29	3.37	3.03
		PCV	9.92	7.63	5.76	8.66	9.55	7.84	6.49	8.95
		h ² _b	15.36	28.25	42.69	21.93	8.49	17.61	26.92	11.44
		GA	2.44	4.87	8.24	3.63	1.57	3.42	5.55	2.14
	Sudani	GCV	4.54	4.33	3.95	4.61	2.75	3.28	3.30	3.11
		PCV	10.58	6.78	5.52	8.69	9.34	7.27	6.30	8.30
		h ² _b	18.36	40.82	51.25	28.08	8.69	20.36	27.55	14.06
		GA	2.61	6.83	9.44	4.26	1.63	4.10	5.81	2.72
Dry sepals weight /plant (g)	Masri	GCV	6.55	6.84	6.35	6.83	4.64	5.52	5.67	5.04
		PCV	16.80	12.94	9.75	14.64	16.15	13.29	11.01	15.20
		h ² _b	15.19	27.96	42.48	21.79	8.25	17.25	26.48	11.00
		GA	1.43	2.86	4.85	2.13	0.90	1.99	3.24	1.22
	Sudani	GCV	7.63	7.32	6.68	7.78	4.65	5.55	5.58	5.27
		PCV	17.94	11.47	9.34	14.73	15.78	12.28	10.63	14.02
		h ² _b	18.11	40.69	51.12	27.89	8.69	20.43	27.58	14.11
		GA	1.53	4.03	5.58	2.50	0.97	2.43	3.44	1.62

GCV = Genotypic coefficient of variation, PCV = Phenotypic coefficient of variation, h²_b = Broad sense heritability%, GA = Genetic advance

Greater h²_b was estimated for all traits (except PH) of both Roselle varieties in M₂ than M₃ generation affected by all gamma doses. But, the greater h²_b for PH was estimated in M₃ of both varieties affected by all radiation doses. The highest h²_b value was computed in NC trait affected by 240 Gray for both Masri (42.69%) and Sudani (51.25%) in M₂ generation. However, the higher GA% was received for all traits (except PH) of both varieties in M₂ than M₃ generation affected by all gamma doses. The greater GA% for PH was obtained in M₃ of both varieties affected by all gamma doses. The maximum GA% was obtained for NC trait affected by 240 Gray for both Masri (8.24%) and Sudani (9.44%) in M₂ generation. Hence the lowest GA% was estimated for Masri NMB (0.39%) affected by 80 Gray in M₃ and for Sudani PH (0.41%) affected by 80 Gray in M₂ generation (Table 7).

In general, greater GCV% values were computed in M₂ than M₃ generation for most of studied traits of both Roselle varieties affected by the different gamma ray doses in both soaked and dry categories. Irradiated application in soaked seed category gave higher PCV% in M₂ than M₃ for all traits of both varieties except Sudani NMB and Masri DSW. M₃ had the higher PCV% in dry seed category for Masri NMB, NC and DSW, as well as Sudani PH and NMB. Opposite to PCV% of Masri PH and NTB addition to Sudani NTB were higher in M₂ generation. Gamma exposed dry seed category presented greater h²_b in M₂ for all characters of both varieties except PH trait. Same trend was noticed in soaked seed category for Masri NC and DSW as well as Sudani PH and NMB, but higher h²_b was noticed for Masri PH and NTB as well as Sudani NC and DSW in M₃ generation. GA% results had approximately the same response of h²_b to gamma radiation. In this conception, ^{57, 58, 59} found greater GCV%, h²_b and GA% in the second season. They attributed this difference to lesser effect of the environmental factors. On the other hands, highest GCV% exhibited in NMB affected by 60 Gray for both varieties at M₂ generation. NMB in M₂ of Masri and M₃ of Sudani had the highest PCV% affected by 80 Gray. ⁵⁷ found that the trait of seed yield/plant exhibited the highest GCV% in the first and second seasons. Whereas, the plant height trait exhibited the lowest GCV% value in both seasons. Furthermore, the highest value of heritability broad sense (h²_b) was achieved in the trait of NC affected by 40 Gray in Masri M₂ and Sudani M₃. Therefore, it can be indicated that this trait (NC) possessed a wide range of genetic variability, so it can be improved by the mass selection. ⁶⁰ concluded that more variable conditions reduce heritability, whereas uniform conditions increase it. Meanwhile, ^{57, 61, 62} concluded that any plant character depends on many components which are greatly influenced by environment exhibits low heritability. However, NC character had the highest GA% affected by 40 Gray in M₂ generation of both varieties, whereas the lowest GA% was obtained by Masri NMB and Sudani PH affected by 80 Gray. These results indicate that the GA% for a trait depends on the amount of genetic variability of such trait. Similar conclusions have been drawn by ⁵⁷ who stated that there was no definite trend between genetic coefficients of variation and heritability or between heritability and genetic advance. Therefore, conjunction of heritability estimates with genetic advance in a selection program is essential.

Phytochemical screening of Roselle calyces (sepals) for M₃ generation of Masri and Sudani varieties response to gamma irradiation was presented in Table (8). The responsibility of Soaked seed category to gamma irradiation was higher than dry seed category for most evaluated chemicals in both Roselle varieties. Moreover, Sudani variety had the higher values for most evaluated chemicals in both soaked and dry seed categories. Total soluble solids (ranged from 47.5 to 56.5 g/100gDW) and pH value (ranged from 2.0 to 2.3 pH.) had slight response to gamma ray exposing in both varieties and categories. Irradiation doses 60 Gray in soaked seed category and 240 Gray in dry seed category stimulated the highest values for total acidity, soluble sugars, total anthocyanins, total phenolics and antioxidant activity in Masri variety. However the dose of 20 Gray stimulated the highest values of sugars, anthocyanins and antioxidant activity, but the highest acidity and phenolics were achieved by 60 Gray in Sudani variety at soaked seed category. Meanwhile, the dry seed irradiated category of Sudani variety did not show general dose effect, where 320 Gray stimulated the highest phenolics and antioxidant activity, the highest sugars was obtained by 240 Gray and the highest acidity was obtained by 80 Gray (Table 8).

Generally, greater acidity, anthocyanins, phenolics and antioxidant activity were recorded for Sudani variety comparing with those of Masri variety. The responsibility of soaked seed category to gamma irradiation was higher than dry seed category in both Roselle varieties. Irradiation doses 60 Gray in soaked seed category and 240 Gray in dry seed category stimulated the highest values for most evaluated chemicals in Masri variety. In Sudani variety the highest sugars, anthocyanins and antioxidant activity were stimulated by 20 Gray, but the highest acidity and phenolics were achieved by 60 Gray at soaked seed category, while the dry seed category did not show general dose effect. These results are in agreement with ⁶³ who found greatest anthocyanins, sugars and pH in the sepals of Sudani variety comparing with Masri and White Roselle varieties. They returned the differences in sepal chemicals to the varietal genetics and environmental conditions which exerted a great influence on the metabolism of biochemical components. Furthermore ^{33, 64, 65} reported that gamma irradiation can modify the tannin and the phenol contents. This modification is very favorable, since this anti-nutritional factor had the capacity for decreasing protein digestibility ⁶⁶. On the other hand, gamma irradiation resulted in a significant tendency to decreasing DPPH (Diphenyl-2-picrylhydrazyl) radical-scavenging activity of different methanolic extracts ³³. However ⁵³ reported that the dose of 40 Gray had the capacity to enhance total flavones content and coloring matters (anthocyanins) in Roselle plants.

Table 8. Seven chemical evaluations of the plant calyces (DW) for M3 generation of Masri and Sudani Roselle varieties treated with five radiation doses in soaked and Dried seed categories

Category	Variety	Dose	Total soluble solids g/100g	pH	Total acidity mg acid/g	Soluble sugars g/100g	Total anthocyanins mg/g	Total phenolics mg/g	Anti-oxidant activity $\mu\text{mol/g}$
Soaked seed	Masri	Control	56.0	2.1	127.8	10.4	4.5	38.8	93.2
		20 Gy	47.5	2.2	61.5	5.1	3.1	32.1	75.9
		40 Gy	53.0	2.1	84.5	10.0	5.0	39.2	97.9
		60 Gy	55.0	2.2	115.6	15.7	6.2	64.1	183.4
		80 Gy	54.4	2.2	57.0	15.8	6.1	56.3	145.0
	Sudani	Control	55.8	2.1	104.1	15.0	6.2	61.3	169.0
		20 Gy	56.5	2.2	54.1	14.2	6.4	59.3	179.5
		40 Gy	54.2	2.1	113.3	13.6	5.8	47.7	112.8
		60 Gy	51.2	2.2	128.3	13.7	6.3	64.0	150.4
		80 Gy	50.9	2.1	65.5	13.6	6.2	57.7	163.2
Dried seed	Masri	Control	51.5	2.1	82.0	6.1	2.5	29.3	54.8
		80 Gy	48.2	2.0	103.7	14.2	4.3	39.8	86.4
		160 Gy	51.2	2.0	96.0	7.8	2.2	31.4	67.2
		240 Gy	53.1	2.0	120.3	15.3	5.3	39.5	110.4
		320 Gy	50.0	2.0	74.9	8.3	4.3	37.2	102.7
	Sudani	Control	51.3	2.1	103.5	12.9	6.2	61.1	163.7
		80 Gy	52.6	2.0	127.4	10.1	5.7	46.6	93.1
		160 Gy	51.9	2.2	77.5	11.8	6.2	58.7	163.2
		240 Gy	55.1	2.3	49.3	14.2	6.2	60.7	176.7
		320 Gy	49.7	2.1	70.4	12.6	6.2	64.8	188.1

References

- Schippers RR. African Indigenous Vegetable, An overview of the cultivated species, Chatham, UK. Natural Resources Institute/ACP-EU Technical Centre for Agricultural and Rural Cooperation, 2000, pp. 122-133.
- Falusi OA, Dangana MC, Daudu OAY, Oluwajobi AO, Abejide DR, Abubakar A. Evaluation of some Roselle (*Hibiscus sabdariffa* L.) germplasm in Nigeria. Inter. J Biotech. Food Sci., 2014, 2:16-20.
- Fasoyiro SB, Ashaye OA, Adeola A, Samuel FO. Chemical and storability of fruits flavored (*Hibiscus sabdariffa*) drinks. World J. Agric. Sci., 2005, 1: 165-168.
- Atta S, Seyni HH, Bakasso Y, Sarr B, Lona I, Saadou M. Yield character variability in Roselle (*Hibiscus sabdariffa* L.). Afr. J Agric. Res., 2011, 6:1371-1377.
- Atta S, Diallo AB, Sarr B, Bakasso Y, Saadou M, Glew RH. Variation in macro-elements and protein contents of Roselle (*Hibiscus sabdariffa* L.) from Niger. Afr. J. Food Agric. Nutr. Dev., 2010, 10: 2707-2718.
- Delgado-Vargas F, Parcedes-Lopez O. Natural colorants for food and nutraceutical uses. CRC Press, LLC: Boca Raton, FL, 2003, p.327.
- Akanbi WB, Olaniyan AB, Togun AO, Hupeju AEO, Alaniran OA. The effects of organic fertilizer on growth, calyx yield and quality of Roselle (*Hibiscus sabdariffa* L.). Am.-Eurasian J. Sustain. Agric., 2009, 3:652-657.
- Wahba EH, Mohamed MA, Eraki MA, Mazrou MM, Afify MM, Mahfoz S. Growth and chemical components of roselle in relation to the irrigation and transparent, calcium carbonate. Egypt J. Hort., 2001, 28: 485-504.
- Ottai MES, Abd-El-Khair H. Influence of transplantation and biological control of *Fusarium* wilt on some quantitative characters of three roselle cultivars. Minufiy J. Agric. Res., 2004, 29: 1091-1108.
- Ottai MES, Aboud KA, Mahmoud IM, El-Hariri DM. Stability analysis of roselle cultivars (*Hibiscus sabdariffa* L.) under different nitrogen fertilizer environments. World J. Agric. Sci., 2006, 2: 333-339.
- Hong V, Wroslad O. Use of HPLC separation/photodiode array detection for characterization of anthocyanin. J. Agric. Food Chem., 1990, 38:708-715.
- Cissé M, Dornier M, Sakho M, N'Diaye A, Reynes M, Sock O. Le bissap (*Hibiscus sabdariffa* L.) composition et principales utilisations. Fruits, 2009, 64:179-193.

13. Babajide JM, Bodunde JG, Salami AA. Quality and sensory evaluation of processed calyces of six varieties of Roselle (*Hibiscus sabdariffa* L.). Nigerian J. Hort. Sci., 2004, 9: 110-115.
14. Wong P-K, Yusof S, Ghazali HM, Che man YB. Physicochemical characteristics of Roselle (*Hibiscus sabdariffa* L.). Nutr. Food Sci., 2000, 32: 68-73.
15. Odigie IP, Ettarh RR, Adigun S. Chronic administration of aqueous extract of *Hibiscus sabdariffa* attenuates hypertension and reverses cardiac hypertrophy in 2K-1C hypertensive rats. J Ethnopharmacol., 2003, 86: 181-185.
16. Chen CC, Hsu JD, Wang SF, Chrang HC, Yang MY, Kao ES, Ho YO, Wang CJ. *Hibiscus sabdariffa* extract inhibit the development of atherosclerosis in cholesterol-fed rabbits. J. Agric. Food Chem, 2003, 51: 5472-5477.
17. Oboh G, Elusiyam CA. Nutrient composition and antimicrobial activity of sorrel drinks (soborodo). J. Med. Food, 2004, 7: 340-342.
18. Akindohunsi AA, Olaleye MT. Toxicological investigation of aqueous-methanolic extract of the calyces of *Hibiscus sabdariffa* L. J Ethnopharmacol, 2003, 89: 161-164.
19. Hussein RM, Shahein YE, El Hakim AE, Awad HM. Biochemical and molecular characterization of three colored types of roselle (*Hibiscus sabdariffa* L.). J Ameri. Sci., 2010, 11: 726-733.
20. Ibrahim MM, Hussein RM. Variability, heritability and genetic advance in some genotypes of Roselle *Hibiscus sabdariffa* L. World J. Agric. Sci., 2006, 2:340-345.
21. Haris A, Jusoff K. Gamma Ray Radiation Mutant Rice on Local aged Dwarf. Middle East J Sci. Res., 2013, 15: 1160-1164.
22. El Sherif F, Khattab S, Goniam E, Salem N, Radwan K. Effect of gamma irradiation on enhancement of some economic traits and molecular changes in *Hibiscus Sabdariffa*. Life Sci. J, 2011, 8: 220-229.
23. Chandrashekar K, Somashekarappa H, Souframanien J. Effect of gamma irradiation on germination, growth, and biochemical parameters of *Terminalia arjuna* Roxb. Radiation Protection and Env., 2013, 36: 38-44.
24. Jan S, Parween T, Siddiqi T. Effect of gamma radiation on morphological, biochemical, and physiological aspects of plants and plant products. Envi. Rev., 2012, 20: 17-39.
25. Rahimi MM, Bahrani A. Effect of gamma irradiation on qualitative and quantitative characteristics of Canola (*Brassica napus* L.). Middle-East J Sci. Res., 2011, 8: 519-525.
26. Kiong A, Pick AL, Lai SHG, Harun AR. Physiological responses of *Orthosiphon stamineus* plantlets to gamma irradiation. Am-Eurasian J. Sustain. Agric., 2008, 2: 135-149.
27. Ilyas S, Naz S. Effect of gamma irradiation on morphological characteristics and isolation of curcuminoids and oleoresins of *Curcuma longa* L. J Animal & Plant Sci., 2014, 24: 1396-1404.
28. Schum A. Mutation breeding in ornamentals and efficient breeding method. Acta Hort., 2003, 612: 47-60.
29. Song HS, Kang SY. Application of natural variation and induced mutation in breeding and functional genomics: Papers for International Symposium; Current Status and Future of Plant Mutation Breeding. Korean J Breed. Sci., 2003, 35: 24-34.
30. Osman M, Golam F, Saberi S, Abdul Majid N, Nagoor NH, Zulqarnain M. Morpho-agronomic analysis of three roselle (*Hibiscus sabdariffa* L.) mutants in tropical Malaysia. Australian J. Crop Sci., 2011, 5:1150-1156.
31. Bhosale UP, Hallale BV. Gamma radiation induced mutations in black gram (*Vigna mungo* (L.) Hepper). Asian J Plant Sci. Res., 2011, 1: 96-100.
32. Girji M, Gnanamurthy S, Dhanavel D. Cytogenetics effect of gamma rays on root meristem cells of *Vigna unguiculata* (L.). European J Experimental Biol., 2013, 3:38-41.
33. Musa HAA, Ahmed EEA, Osman GAM, Ali HA, Ludwig-Müller J. Microbial load and stability of some phytochemical components of selected Sudanese medicinal plant materials as affected by gamma irradiation. Inter J Sci. Nature, 2011, 2:204 – 209.
34. SPSS Inc.. SPSS 11; 0 for windows. USA, Inc., 2001. Available online: <http://www.spss.com>.
35. Steel RG, Torrie JH. Principles and Procedures of Statistics. Second Ed. McGraw-Hill, Inc, New York, 1980.
36. Robinson HF, Comstock RE, Harvey PH. Genotypic and phenotypic correlation in corn and their implications to selection. Agron. J, 1951, 43: 282-287.
37. Burton GW. Quantitative Inheritance in Grasses. Proc. 6th Int. Grassland Cong., 1952, PP. 277-283.
38. Johnson HW, Robinson HF, Comstock RE. Estimates of genetic and environmental variability in soybean. Agron. J, 1955, 47: 314-318.

39. Shin Yi, Liu RH, Nockc JF, Holliday D, Watkins CB. Temperature and relative humidity effects on quality, total ascorbic acid, phenolics and flavonoid concentrations, and antioxidant activity of strawberry. *Postharvest Biol. Technol.*, 2007, 45: 349–357.
40. Pineli LO, Moretti CL, dos Santos MS, Campos AB, Brasileiro AV, Co´ rdova AC, Chiarello M. Antioxidants and other chemical and physical characteristics of two strawberry cultivars at different ripeness stages. *J Food Composition and Analysis*, 2011, 24: 11–16.
41. Dubois M, Gilles KA, Hamilton JK, Rebers PA, Smith F. Colorimetric method for determination of sugars and related substances. *Anal. Chem.*, 1956, 28:350-356.
42. Soong Y, Barlow PJ. Antioxidant activity and phenolic content of selected fruit seeds. *Food Chem.*, 2004, 88: 411–417.
43. Brand-Williams W, Cuvelier ME, Berset C. Use of free radical method to evaluate antioxidant activity. *Lebensmittel Wissenschaft Und Technol.*, 1995, 28: 25-30.
44. Amir I, Emmy H, Khairu I, Halimatul S, Mohamed N. Roselle (*Hibiscus sabdariffa* L.) Seeds- Nutritional Composition, Protien Quality and Health Benefits. Global science books. Bailiere Tinnal Ltd., London, 2008.
45. Mahadevan N, Shivali OE, Pradeep K. *Hibiscus sabdariffa* L. An overview. *Nat. Prod. Rad.*, 2009, 8:77-83.
46. Yandong QI, Kitl C, Fatemah M, Mila B, Janet G. Biological characteristics, Nutritional and Medicinal value of Rosele, *Hibiscus sabdariffa*. *Urban for. Natl. Resour. Environ.*, 2012, p. 604.
47. Heywood VH. Flowering Plants of the World. Oxford University Press, Oxford, London, 1978, pp. 94-95.
48. Pitirmovae MA. Effect of gamma rays and mutagens on barley seeds. *Fiziol. Res*, 1979, 6: 127–31.
49. Sundaravadivelu K, Ranjithselvi P, Reddy VRK. Induced genetic variability in cotton (*Gossypium hirsutum* L.) for yield and its components. *Crop Res. Hisar.*, 2006, 32: 442-446.
50. Dubey AK, Yadav JR, Singh B. Studies on induced mutations by gamma irradiation in okra (*Abelmoschus esculentus* (L.) Monch.). *Progressive Agric.*, 2007, 7: 46-48.
51. Sharma B, Mishra K. Micro-mutations for fruit number, fruit length and fruit yield characters in gamma-irradiated generation of ANKUR-40 variety of okra [*Abelmoschus esculentus* (L.) Monech]. *Int. J Plant Sci. Muzaffarnagar*, 2007, 2: 208-211.
52. Mishra MN, Qadri H, Mishra S. Macro and micro mutations, in gamma-rays induced M2 populations of Okra (*Abelmoschus esculentus* (L) Moench). *Int. J Plant Sci. Muzaffarnagar*, 2007, 2: 44-47.
53. Abo-EI-Seoud MA, Hashim MF, Farid AM. Combined Effect of Gamma radiation and potassium fertilization on growth and coloring matter contents of roselle (*Hibiscus Sabdariffa* L.). *Second Arab Conference on the Peaceful Uses of Atomic Energy, Cairo 5-9 Nov.*, 1994, pp: 863-874.
54. Token C, Uzun B, Canci H, Ceylan FO. Effects of gamma irradiation on the shoot length of *Cicer* seeds. *Radiat. Phys. Chem.*, 2005, 73: 365-367.
55. Kon E, Ahmed OH, Saamin S, Majid NM. Gamma radiosensitivity study on long bean (*Vigna sesquipedalis*). *Am. J Appl. Sci.*, 2007, 4: 1090-1093.
56. Norfadzrin F, Ahmed OH, Shaharudin S, Rahman DA. Apreliminary study on gamma radiosensitivity of tomato (*Lycopersicon esculentum*) and okra (*Abelmoschus esculentus*). *Int. J. Agric. Res.*, 2007, 2: 620-625.
57. Ibrahim EB, Abdalla AH, Ibrahim EA, El Naim AM. Variability in some roselle (*Hibiscus sabdariffa* L.) genotypes for yield and its attributes. *Inter. J Agric. Forestry*, 2013, 3: 261-266.
58. Reddy KR, Singh KP, Rai AK. Variability and association analysis in okra. *Mdras Agric. J*, 1985, 72:478-480.
59. Vijay OP, Manohar MS. Studies on genetic variability, correlation and path analysis in okra (*Abemoschus esculentus* L. (Moench). *Indian J. Hort.*, 1990, 47:97-103.
60. Falconer DS. Introduction to Quantitative Genetics. 2nd ed. Longman, London, 1980, Pp: 322-323.
61. Mostafa MR, Islam MR, Alam ATM, Ali SM, Moll MAF. Gentic variability, heritability and correlation studies in Kenaf (*Hibiscus cannabinus* L.). *J Biol. Sci.*, 2002, 2:422-424.
62. Louis SJ, Kadams AM, Simon SY, Mohammed SG. Combining ability in roselle cultivars for agronomic traits in Yola, Nigeria. *Greener J Agric. Sci.*, 2013, 3: 145-149.
63. Ottai MES, Abdel-Moniem ASH, El-Mergawi RA. Effect of variety and location on growth and yield components of roselle (*Hibiscus sabdariffa* L.) and its infestation with the spiny bollworm *Earias insulana* (Boisd). *Archives of Phytopathol. Plant Protec.*, 2004, 37: 215-231.

64. Villavicencio ALCH, Mancini-Filho J, Delincee H, Greiner R. Effect of irradiation on anti-nutrients (total phenolics, tannins and phytate) in Brazilian beans. *Radiation Physics and Chemistry*, 2000, 57, 289-293.
65. Brigide P, Canniatti-Brazaca SG. Antinutrients and “*in vitro*” availability of iron in irradiated common beans (*Phaseolus vulgaris*). *Food Chem.*, 2006, 98, 85-89.
66. Toledo TCF, Canniatti-Brazaca SG, Arthur V, Piedade SMS. Effects of gamma radiation on total phenolics, trypsin and tannin inhibitors in soybean grains. *Radiation Physics and Chemistry*, 2007, 76, 1653-1656.
