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Synthesis, characterization, molecular docking and evaluation of antimicrobial activity of some 3-heteroaryl substituted chromen-2-one derivatives

Rajesh B. Patil*, Sanjay D. Sawant

Sinhgad Technical Education society's, Smt. Kashibai Navale College of Pharmacy, Pune-Saswad Road, Kondhwa (Bk), Pune-411048, Maharashtra, India

Abstract: Chromen-2-ones commonly called coumarins are important synthetic and phytogenic bioactive molecules. Owing to their diverse biological activities, synthesis of substituted coumarin derivatives has been tried as antimicrobial agent in current investigation. Series of 1-(substituted phenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea derivatives (CTU1-12) and 3-(5-substituted phenyl-1,3,4-oxadiazol-2-yl)-2H-chromen-2-one derivatives (COD1-9) were synthesized. Docking studies were carried out with Autodock vina, using MGL Tools 1.5.6. Docking studies in Autodock vina suggested hydrogen bond interaction between carbonyl group of coumarin ring or carbonyl group of urea substituent with key residue Asn46 for compounds from CTU series and hydrogen bond interaction between nitrogen of oxadiazole ring and Asn46. Compounds CTU1, CTU4, CTU7 and CTU10 showed moderate antibacterial activity against gram positive organisms. **Keywords:** Coumarin, 1,3-thiazole, 1,3,4-oxadiazole, Autodock vina, Antimicrobial.

Introduction

Chromen-2-ones commonly called as coumarins are widely found phyto constituents. Its presence in some plants confer them defence against pathogens, insects, pests and herbivores¹. Coumarins possess derivatives diverse pharmacological activities including antitumor ^{2, 3}, antivascular ⁴, antimicrobial⁵, antioxidant ⁶, TNF- α inhibitor⁷, antifungal⁸, anticoagulant⁹, estrogenic¹⁰, antiviral¹¹, anthelmintic¹², anti-HIV¹³, antitubercular¹⁴, anti-inflammatory¹⁵, herbicidal¹⁶, analgesic¹⁷ and anticonvulsant^{18, 19} activity. Anticoagulant coumarin derivatives like dicoumarol, warfarin, phenoprocoumon and acenocoumarol, photosensitizing agents used in psoriasis and vitiligo like methoxsalen, trioxsalen, antibacterial agents like novobiocin and clorobiocin and antioxidant and antiproliferative agent ellagic acid have been approved as a drugs ²⁰. Substituted 1, 3-thiazole derivatives^{21, 22} and 1, 3, 4- oxadiazole^{23, 24} derivatives have been reported as antimicrobial agents. Microbial resistance has posed a series threat to mankind as many new strains of bacteria have been reported which are resistant to currently available antibacterials and antibiotics ²⁵. In view of this, 1, 3- thiazole substituted and 1, 3, 4- oxadiazole substituted coumarin derivatives were synthesised and characterized by FT-IR, ¹H-NMR and mass spectrometry. The docking studies, evaluation of antimicrobial activity against two gram-positive organisms *viz. Bacillus subtilis* and *Staphylococcus aureus* and two gram-negative organisms *viz. Escherichia coli* and *Pseudomonas aeruginosa* of the synthesised compounds are reported in this paper.

Materials and Methods

Docking studies

In the present investigation, the X-ray crystal structure of the antimicrobial agent Clorobiocin bound to topoisomerase II DNA gyrase was obtained from the RCSB Protein Data Bank (PDB ID: 1KZN). The protein with resolution 2.30 A^0 with 205 amino acid residues was processed by removing water and clorobiocin.

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Energy minimization of this clean protein was carried out UCSF Chimera ²⁶ with Amber ff12SB force field and combination of 10,000 steepest descent and conjugate gradient steps with 0.02 A⁰ step size. 2D and 3D structures of all the synthesized compounds were drawn in Marvin Sketch (a structure drawing program). Geometry optimization of these 3D structures was carried out in ArgusLab 4.0.1 (from Thomson and Planaria Software LLC) on semi empirical quantum mechanical basis with parameterized model number 3 (PM3) hamiltonian, until restricted closed shell hartree-fock self consistent field formalism converses to 10⁻¹⁰ kcal/mol and steepest descent geometry search criteria until gradient converses to 10⁻⁶ kcal/mol. Gasteiger partial atomic charges for optimized molecules were computed in UCSF chimera. The energy minimized protein and geometry optimized structures of compounds were pre processed in MGLtools1.5.4²⁷. Docking simulation was carried out in Autodock Vina ²⁸ using the grid box of size 18 x 18 x 18 with 1 A⁰ spacing was defined along x, y and z axis. The analysis of binding free energy and interactions of ligands with residues at active site was carried out by using Pymol and Discovery studio 3.5.

Chemistry

The reagents and solvents used during synthesis were of laboratory grade obtained from Thomas Baker and Loba Chemie. The melting point of the compound was determined by open capillary method, expressed in^oC. The reactions were monitored preparative TLC from Merck with the solvent system Chloroform: methanol in the ratio of 9:1. IR spectra were recorded on Shimadzu FT-IRAffinity-1 spectrophotometer by KBr pellet technique and are expressed in cm⁻¹. ¹H-NMR spectra was recorded on Bruker Avance 400 MHz FT-NMR spectrometer using CDCl₃ or DMSO as solvent and TMS as internal standard (δ ppm). The chemical shifts are expressed in δ ppm and splitting patterns are designated as s: singlet; d: doublet; q: quartet; m: multiplet. Mass spectra were recorded using Waters Quatropole-TOF Micro Mass (Electro spray ionization) Spectrometer. The strategy adopted in the synthesis of compounds is shown in **Scheme 1** and **Scheme 2**.



Scheme 1: Synthesis of 1-(substituted phenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea derivatives; (i) Ethylacetoacetate, piperidine, RT; (ii) Bromine, dioxane; (iii) Thiourea, reflux; (iv) Ar-COOH, Phenyl chloroformate, Dimethoxyethane, Sodium azide, 70 °C



Scheme 2: Synthesis of 3-(5-substituted phenyl-1,3,4-oxadiazol-2-yl)-2H-chromen-2-one; (i) Diethylmalonate, piperidine, glacial acetic acid; (ii) Hydrazine hydrate (99%), reflux; (iii) Ar-COOH, POCl₃

Synthesis of 3-acetylcoumarin (1a)

Piperidine (0.1 ml) was added dropwise with stirring to the mixture of salicylaldehyde (18 mmol) and ethylacetoacetate (24 mmol) in 1 ml ethanol. The solid obtained was recrystallized from ethanol to yield 72 % pale yellow solid of 1a; m.p. 118-119 $^{\circ}$ C.

Synthesis of 3-(2-bromoacetyl)-2H-chromen-2-one (1b)

In the solution of 15 mmol of **1a** in 15 ml dioxane, bromine (0.7 ml, 15 mmol) was added with continuous stirring. The solid separated was filtered and air dried. The crude **1b** was recrystallized from benzene-petroleum ether mixture in 60 % yield, m.p. 122-125 $^{\circ}$ C.

Synthesis of 3-(2-amino-1,3-thiazol-5-yl)-2H-chromen-2-one (1c)

Compound **1b** (10 mmol) and thiourea (10 mmol) was dissolved 30 ml of absolute ethanol and refluxed for 4 hrs. Solvent was evaporated on rotary evaporator and the residue was poured over crushed ice. Ammonia was added to make the solution distinctly basic, the solid obtained was filtered and washed with cold water. The crude **1c** was recrystallized from ethanol to obtain pale yellow solid in 55 % yield; m.p. 138-140 $^{\circ}$ C. ¹H NMR (CDCl₃): (ppm) 4.97 (2H, s, -NH₂), 7.28-7.78 (5H, m, Ar-H), 8.51 (1H, pyran-H); MS: m/z = 245.0520 (M⁺+1).

General procedure for the synthesis of 1-(substituted phenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea derivatives (CTU1-12)

To the solution of sodium azide (0.110 gm, 1.7 mmol), sodium acetate (12.30 mg, 0.15 mmol) and appropriate aromatic acids (1 mmol) in dimethoxyethane (DME) (10 ml); phenylchloroformate (140 μ l, 1.10 mmol) was added at room temperature and the mixture was stirred for 8 hrs. Compound **1c** (366 mg, 1.50 mmol) was added at 75 $^{\circ}$ C and the solution was stirred at this temperature for 16 hrs in Radlys Carousel 6 place reaction station. The reaction mixture was allowed to cool to room temperature. Hexane 40 ml and water 10 ml was added to the cooled reaction mixture and stirred for 20 min. The solid obtained was filtered and air dried.

Synthesis of ethyl 2-oxo-2H-chromene-3-carboxylate (2a)

The mixture of salicyladehyde 3.6 mmol and diethylmalonate 7.2 mmol was stirred at room temperature. Piperidine 0.25 ml was slowly added to the mixture under stirring and stirring was continued for 30 min. The resulting solution was acidified with glacial acetic acid. The solid obtained was filtered and recrystallized from ethylacetate to obtain 2a in 60 % yield.

Synthesis of ethyl 2-oxo-2H-chromene-3-carbohydrazide (2b)

The mixture of compound 2a (1.9 gm), hydrazine hydrate (2 ml) in 15 ml absolute ethanol was refluxed for 6hr. The solid obtained on cooling the reaction mixture was filtered and recrystallized from ethanol to obtain 2b in 40 % yield.

General procedure for the synthesis of 3-(5-substituted phenyl-1,3,4-oxadiazol-2-yl)-2H-chromen-2-one (COD1-9)

Compound **2b** (2.4 mmol) was dissolved in 10 ml phosphorous oxychloride. Appropriate aromatic acids (2.4 mmol) were added and the mixture was refluxed for 6 hrs. The reaction mixture was cooled and poured over crushed ice and the solution was neutralized with 20% sodium bicarbonate solution. The solid separated was filtered, air dried and recrystallised from methanol to afford 37-55% of **COD1-9**.

3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]-1-phenylurea (CTU1)

Pale green solid, Yield: 45 %, m.p. 201-203 0 C, Mol. Wt. 363.39, 1102 (C-O-C, str.), 1506 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3100 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.28 (2H, s, NH₂), 7.00-7.92 (9H, m, Ar-H), 8.14 (1H, s, thiazole-H), 8.51 (1H, s, pyran-H). MS: m/z = 363 (M⁺)

1-(4-chlorophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU2)

Pale yellow solid, Yield: 57 %, m.p. 168-170 °C, Mol. Wt. 397.02, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.27 (2H, s, NH₂), 7.04-8.18 (8H, m, Ar-H), 8.50 (1H, s, thiazole-H), 8.59 (1H, s, pyran-H).

1-(4-bromophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU3)

Pale green solid, Yield: 43 %, m.p. 175-177 ⁰C, Mol. Wt. 442.28, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.29 (2H, s, NH₂), 6.99-7.72 (8H, m, Ar-H), 8.13 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(2-chlorophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU4)

Pale green solid, Yield: 45 %, m.p. 158-160 °C, Mol. Wt. 397.83, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.29 (2H, s, NH₂), 7.00-8.12 (8H, m, Ar-H), 8.49 (1H, s, thiazole-H), 8.57 (1H, s, pyran-H).

1-(4-methoxyphenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU5)

Pale green solid, Yield: 50 %, m.p. 210-212 ⁰C, Mol. Wt. 393.41, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 2.54 (3H, s, CH₃), 3.28 (2H, s, NH₂), 7.01-7.71 (8H, m, Ar-H), 7.73 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(4-nitrophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU6)

Pale green solid, Yield: 57 %, m.p. 196-198 ⁰C, Mol. Wt. 408.38, 1102 (C-O-C, str.), 1506 (Ar C=C), 1665 (N-H, bend), 1604 (-C=N), 1696 (C=O), 3100 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.28 (2H, s, NH₂), 7.03-7.73 (8H, m, Ar-H), 8.16 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(3-chlorophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU7)

Pale green solid, Yield: 40 %, m.p. 181-183 ⁰C, Mol. Wt. 397.83, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.28 (2H, s, NH₂), 7.00-7.72 (8H, m, Ar-H), 8.14 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(3-hydroxyphenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU8)

Pale green solid, Yield: 43 %, m.p. 163-165 ⁰C, Mol. Wt. 379.38, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3200 (O-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 2.55 (1H, s, OH), 3.30 (2H, s, NH₂), 6.92-7.68 (8H, m, Ar-H), 8.05 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(2, 4-dichlorophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU9)

Pale green solid, Yield: 51 %, m.p. 178-180 ⁰C, Mol. Wt. 432.28, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.30 (2H, s, NH₂), 6.93-7.69 (7H, m, Ar-H), 8.07 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(4-fluorophenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU10)

Pale green solid, Yield: 40 %, m.p. 116-118 °C, Mol. Wt. 381.38, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.29 (2H, s, NH₂), 6.97-7.68 (8H, m, Ar-H), 7.70 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(4-hydroxyphenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU11)

Pale green solid, Yield: 48 %, m.p. 149-151 ^oC, Mol. Wt. 379.38, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3200 (O-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 2.55 (1H, s, OH), 3.31 (2H, s, NH₂), 6.86-7.66 (8H, m, Ar-H), 8.00 (1H, s, thiazole-H), 8.50 (1H, s, pyran-H).

1-(2-methoxyphenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea (CTU12)

Pale green solid, Yield: 42 %, m.p. 198-200 ⁰C, Mol. Wt. 393.41, 1102 (C-O-C, str.), 1539 (Ar C=C), 1665 (N-H, bend), 1696 (C=O), 3154 (Ar-C-H), 3384 (N-H).; ¹H NMR (CDCl₃): (ppm) 3.28 (2H, s, NH₂), 3.86 (3H, s, CH₃), 6.96-7.68 (8H, m, Ar-H), 7.68 (1H, s, thiazole-H), 8.49 (1H, s, pyran-H).

3-(5-phenyl-1,3,4-oxadiazol-2-yl)-2H-chromen-2-one (COD1)

Pale green solid, Yield: 52 %, m.p. 172-175 0 C, Mol. Wt. 290.27, 1152 (C-O-C, str.), 1570 (Ar C=C), 1623 (N-H, bend), 1740 (C=O), 3100 (Ar-C-H).; 1 H NMR (CDCl₃): (ppm) 6.92-8.14 (9H, m, Ar-H), 8.92 (1H, s, pyran-H). MS: m/z = 291 (M+H)

3-[5-(4-chlorophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD2)

Pale yellow solid, Yield: 45 %, m.p. 198-200 ^oC, Mol. Wt. 324.71, 1195 (C-O-C, str.), 1573 (Ar C=C), 1696 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 6.93-8.17 (8H, m, Ar-H), 8.96 (1H, s, pyran-H).

3-[5-(4-fluorophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD3)

Pale yellow solid, Yield: 40 %, m.p. 186-188 ⁰C, Mol. Wt. 308.26, 1195 (C-O-C, str.), 1573 (Ar C=C), 1696 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 6.96-8.83 (9H, m, Ar-H), 8.95 (1H, s, pyran-H).

3-[5-(2,4-dichlorophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD4)

Pale yellow solid, Yield: 55 %, m.p. 201-203 ^oC, Mol. Wt. 359.16, 1255 (C-O-C, str.), 1581 (Ar C=C), 1696 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 6.93-8.15 (7H, m, Ar-H), 8.98 (1H, s, pyran-H).

3-[5-(4-methoxyphenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD5)

Pale yellow solid, Yield: 48 %, m.p. 138-140 ^oC, Mol. Wt. 320.29, 1174 (C-O-C, str.), 1490 (Ar C=C), 1734 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 3.31 (3H, s, -OCH₃), 6.96-8.13 (8H, m, Ar-H), 8.90 (1H, s, pyran-H).

3-[5-(3-chlorophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD6)

Pale yellow solid, Yield: 37 %, m.p. 145-147 ^oC, Mol. Wt. 324.71, 1152 (C-O-C, str.), 1573 (Ar C=C), 1700 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 6.93-8.17 (8H, m, Ar-H), 8.96 (1H, s, pyran-H).

3-[5-(4-nitrophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD7)

Pale yellow solid, Yield: 55 %, m.p. 193-195 ⁰C, Mol. Wt. 335.27, 1111 (C-O-C, str.), 1540 (Ar C=C), 1683 (C=O), 3100 (Ar-C-H).; 1H NMR (CDCl3): (ppm) 6.96-8.44 (8H, m, Ar-H), 8.70 (1H, s, pyran-H).

3-[5-(4-bromophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD8)

Pale yellow solid, Yield: 51 %, m.p. 171-173 ⁰C, Mol. Wt. 369.16, 1123 (C-O-C, str.), 1570 (Ar C=C), 1681 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 6.97-8.31 (8H, m, Ar-H), 8.96 (1H, s, pyran-H).

3-[5-(2-chlorophenyl)-1,3,4-oxadiazol-2-yl]-2H-chromen-2-one (COD9)

Pale yellow solid, Yield: 46 %, m.p. 127-129 ^oC, Mol. Wt. 324.71, 1152 (C-O-C, str.), 1573 (Ar C=C), 1700 (C=O), 3100 (Ar-C-H).; ¹H NMR (CDCl₃): (ppm) 6.96-8.21 (8H, m, Ar-H), 8.97 (1H, s, pyran-H).

Evaluation of antimicrobial activity

The antimicrobial activity of all the synthesized compounds was examined against different Grampositive (*Bacillus subtilis* and *Staphylococcus aureus*) and Gram-negative (*Escherichia coli* and *Pseudomonas aeruginosa*) by measuring zone of inhibition. The antimicrobial activity was carried out by agar cup plate method at the concentration level 25 μ g/ml. Ofloxacin was used as standard at concentration 25 μ g/ml. Nutrient agar was used as culture media for antibacterial activity. Twenty four hrs old culture of bacterial pathogen was placed in nutrient agar and spread throughout the plate by spread plate technique. Wells were bored using sterile borer at equidistance. The plates were kept at room temperature for 30 minutes. The test compounds, standard and control were placed in respective wells and plates were incubated at 37°C for 36 hrs. Zone of inhibition was measured by zone reader.

Results and Discussion

Docking

The docking protocol adopted in this investigation was validated by docking of clorobiocin to the energy minimized gyrase protein. The residues Asp73, Asn46 and Arg136 are important in making hydrogen bond and Arg76, Pro79, Ile78, Ile90, Thr165, Val43, Val71, Ala47, Val120 and Gly77 are important in hydrophobic interactions. The best conformer generated in docking showed same interactions as shown in Figure 1.



Figure 1: Docked conformer of Clorobiocin (docked conformer shown in cyan and original pose of clorobiocin shown in megenta color)

After docking designed molecules (CTU1-12 & COD1-9) most of the compounds show interaction with Asn46 along with other hydrophobic interactions. The extra hydrogen bonding interactions were not observed for designed compounds as these compounds may not access deep hydrophobic pocket of gyrase protein. The binding free energy in Kcal/mole and interactions are presented in (Table 1). Compounds CTU 1, CTU4, CTU7, CTU10 showed two hydrogen bond interactions. Compounds CTU8, CTU9, COD5, COD8 showed no hydrogen bond interactions. The 2D digram of important interactions between active site residues and compound atoms is shown in figure 2 and 3.

Sr.	Compound	Docking score (Binding	Interactions
No.		free energy) kcal/mol	
1	CTU1	-8.3	Asn46 (H), Arg76
2	CTU2	-8.3	Asn46 (H)
3	CTU3	-8.0	Asn46 (H)
4	CTU4	-8.2	Asn46 (H), Arg76
5	CTU5	-7.9	Asn46 (H)
6	CTU6	-7.9	Asn46 (H)
7	CTU7	-8.5	Asn46 (H), Arg76
8	CTU8	-7.5	Asn46
9	CTU9	-8.6	-
10	CTU10	-8.2	Asn46 (H), Arg76
11	CTU11	-9.0	Asn46 (H)
12	CTU12	-7.9	Asn46 (H)
13	COD1	-8.7	Asn46 (H)
14	COD2	-9.0	Asn46 (H)
15	COD3	-8.9	Asn46 (H)
16	COD4	-9.3	Asn46 (H)
17	COD5	-8.0	Arg76
18	COD6	-8.9	Thr165 (H), Arg76
19	COD7	-8.7	Asn46 (H)
20	COD8	-8.7	-
21	COD9	-9.0	Asn46 (H)
22	Clorobiocin	-6.4	Asn46 (H), Asp73 (H), Arg136 (H), Arg76

Table 1: Docking score (Binding free energy in kcal/mol) and important interaction with residues



Figure 2: 2D interaction diagram for CTU series compounds



Figure 3: 2D interaction diagram for COD series compounds

Antimicrobial activity

The synthesised compounds were evaluated for antibacterial activity using agar cup plate method. Ofloxacin, a well known topoisomerase II DNA gyrase inhibitor, was used as a standard. The results are presented (Table 2). The results show that compounds form CTU series are more active than COD series. The presence of thiazole ring with hydrophobic urea substituent may be the possible reason for better antimicrobial activity of these compounds. CTU 7 was found most active against *S. Aureus*. Docking scores and hydrogen bonds formation with important residues are in good agreement with the antimicrobial activity. All the compounds were found poorly active against gram negative *E. coli and P. aeruginosa*. Compound CTU 1 possess good activity against all the test organisms used in the study, suggesting the importance of unsubstituted phenyl ring on urea nucleus. When compared with the standard ofloxacin, all the synthesised compounds exhibited poor to moderate activity against gram positive and gram negative organisms.

Zone of Inhibition in mm (millimetre)							
Compounds	B. subtilis	S. aureus	E. coli	P. aeruginosa			
CTU1	17.3 ± 0.3	16 ± 0.4	16.4 ± 0.6	15.5 ± 0.4			
CTU2	13.2 ± 0.2	14.1 ±0.8	11.0 ± 1.0	11.9 ± 0.3			
CTU3	13.6 ± 0.3	13.3 ± 0.1	9.2 ± 0.9	11.6 ± 0.6			
CTU4	17.0 ± 1.7	17.8 ± 0.3	11.4 ± 1.0	14.5 ± 0.4			
CTU5	13.8 ± 0.2	14.7 ± 0.2	8.4 ± 0.1	11.5 ± 0.2			
CTU6	14.7 ± 0.2	15.6 ± 0.5	7.1 ± 0.3	11.5 ± 0.4			
CTU7	18.3 ± 0.2	16.9 ± 0.5	13.2 ± 0.7	14.7 ± 0.2			
CTU8	13.8 ± 0.9	14.4 ± 0.5	7.5 ± 0.9	11.3 ± 0.2			
CTU9	12.6 ± 0.1	13.1 ± 0.6	6.6 ± 0.5	9.7 ± 0.1			
CTU10	17.8 ± 0.7	17.1 ± 0.2	13.5 ± 0.9	15.1 ± 0.5			
CTU11	14.4 ± 0.1	13.3 ± 0.5	9.3 ± 1.0	12.0 ± 0.1			
CTU12	15.0 ± 0.2	14.5 ± 0.3	11.3 ± 0.6	12.9 ± 0.1			
COD1	12.2 ± 0.3	12.9 ± 0.6	10.7 ± 0.2	11.4 ± 0.3			
COD2	13.8 ± 0.6	13.2 ± 0.9	9.8 ± 0.3	11.6 ± 0.2			
COD3	13.9 ± 0.7	14.5 ± 0.3	8.9 ± 0.4	11.6 ± 0.1			
COD4	12.2 ± 0.3	13.4 ± 0.4	9.2 ± 0.05	10.9 ± 0.1			
COD5	10.3 ± 0.4	10.9 ± 0.8	9.7 ± 1.0	9.6 ± 0.4			
COD6	9.4 ± 0.3	10.6 ± 0.4	11.6 ± 1.0	9.7 ± 0.4			
COD7	14.1 ± 0.1	13.2 ± 0.4	12.1 ± 0.2	12.1 ± 1.2			
COD8	9.7 ± 0.4	10.6 ± 0.2	9.6 ± 0.7	9.5 ± 0.5			
COD9	13.2 ± 0.3	12.4 ± 0.3	10.3 ± 0.1	11.3 ± 0.3			
Ofloxacin	31.0 ± 0.711	28.8 ± 0.849	28.2 ± 0.205	27.9 ± 0.216			

 Table 2: Antimicrobial activity of synthesized compounds

Data presented in Mean ± SD (N=3)

Conclusion

Series of 1-(substituted phenyl)-3-[5-(2-oxo-2H-chromen-3-yl)-1,3-thiazol-2-yl]urea derivatives (CTU1-12) and 3-(5-substituted phenyl-1,3,4-oxadiazol-2-yl)-2H-chromen-2-one derivatives were synthesized. Docking studies in Autodock vina suggested hydrogen bond interaction between carbonyl group of coumarin ring or carbonyl group of urea substituent with key residue Asn46 for compounds from CTU series and hydrogen bond interaction between nitrogen of oxadiazole ring and Asn46. Compounds CTU1, CTU4, CTU7 and CTU10 showed moderate antibacterial activity against gram positive organisms.

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References

- 1. Mazid M., Khan T. A., Mohammad F., Role of secondary metabolites in defense mechanisms of plants, Biology and Medicine, 2011, 3 (2), 232-249.
- 2. Stefanova T. H., Nikolova N. J., Toshkova R. A., Neychev H.O., Antitumor and immunomodulatory effect of coumarin and 7-hydroxycoumarin against Sarcoma 180 in mice, J. Exp. Ther. Oncol., 2007, 6(2), 107-15.
- 3. Chun Fen Xiao, Li Yang Tao, Hong Yi Sun, Wen Weia, Yu Chena, Li Wu Fu, Yong Zou, Design, synthesis and antitumor activity of a series of novel coumarin–stilbenes hybrids, the 3-arylcoumarins, Chinese Chemical Letters, 2010, 21 (11), 1295-1298.
- 4. Barbera M., Caputo A., Zampiron A., Gobbi S., Rampa A., Bisi A., Carrara M., The ability of coumarin-, flavanon- and flavonol-analogues of flavones acetic acid to stimulate human monocytes, Oncol. Rep., 2008, 19(1), 187-196.
- 5. Adriana Basile, Sergio Sorbo, Vivienne Spadaro, Maurizio Bruno, Antonella Maggio, Nicoletta Faraone, Sergio Rosselli, Antimicrobial and Antioxidant Activities of Coumarins from the Roots of Ferulago campestris (Apiaceae), Molecules, 2009, 14, 939-952.
- 6. Kostova I., Bhatia S., Grigorov P., Balkansky S., Parmar V. S., Prasad A. K., Saso L., Coumarins as antioxidants, Curr. Med. Chem., 2011, 18(25), 3929-51.
- Cheng J. F., Chen M., Wallace D., Tith S., Arrhenius T., Kashiwagi H., Ono Y., Ishikawa A., Sato H., Kozono T., Sato H., Nadzan A. M., Discovery and structure-activity relationship of coumarin derivatives as TNF-alpha inhibitors, Bioorg. Med. Chem. Lett., 2004, 17, 14(10), 2411-2415.
- Ahmed A., Al-Amiery, Abdul Amir Hassan Kadhum, Abu Bakar Mohamad, Antifungal Activities of New Coumarins, Molecules, 2012, 17, 5713-5723.
- 9. Lowenthal J., Birnbaum H., Vitamin K and coumarin anticoagulants: dependence of anticoagulant effect on inhibition of vitamin K transport, Science, 1969, 11, 164(3876), 181-183.
- Jacquot Y., Laios I., Cleeren A., Nonclercq D., Bermont L., Refouvelet B., Boubekeur K., Xicluna A., Leclercq G., Laurent G., Synthesis, structure, and estrogenic activity of 4-amino-3-(2methylbenzyl)coumarins on human breast carcinoma cells, Bioorg. Med. Chem., 2007, 15, 15(6), 2269-2282.
- 11. Hwu J. R., Lin S. Y., Tsay S. C., De Clercq E., Leyssen P., Neyts J., Coumarin-purine ribofuranoside conjugates as new agents against hepatitis C virus, J. Med. Chem., 2011, 14, 54(7), 2114-2126.
- 12. Mahmoud Z. F., Sarg T. M., Amer M. E., Khafagy S. M., Anthelmintic coumarin from Ethulia conyzoides var. gracilis Asch. & Schweinf., Pharmazie. 1983, 38(7), 486-487.
- 13. Kostova I., Raleva S., Genova P., Argirova R., Structure-activity relationships of synthetic coumarins as HIV-1 Inhibitors, Bioinorganic Chemistry and Applications, 2006, 1–9.
- Cardoso S. H., Barreto M. B., Lourenco M. C., Henriques M., Candea A. L., Kaiser C. R., de Souza M. V., Antitubercular activity of new coumarins, Chem. Biol. Drug. Des., 2011, 77(6), 489-493.
- 15. Fylaktakidou K. C., Hadjipavlou-Litina D. J., Litinas K. E., Nicolaides D. N., Natural and synthetic coumarin derivatives with anti-inflammatory/ antioxidant activities, Curr. Pharm. Des., 2004, 10(30), 3813-3833.
- 16. Zhang Fang-He, Wei Yan, Wang Dong, Song Wei, Wang Xian-Gang, Hao Shuang-Hong, Fungicidal and Herbicidal Activities of Coumarins, Agrochemicals, 2011, 6, 1-2.
- 17. Ghate M., Kusanur R. A., Kulkarni M. V., Synthesis and in vivo analgesic and anti-inflammatory activity of some bi heterocyclic coumarin derivatives, Eur. J. Med. Chem., 2005, 40(9), 882-887.
- Amin K. M., Abdel Rahman D. E., Al-Eryani Y. A., Synthesis and preliminary evaluation of some substituted coumarins as anticonvulsant agents, Bioorganic & Medicinal Chemistry, 2008, 16 (10), 5377-5388.
- 19. Siddiqui N., Arshad M. F., Khan S. A., Synthesis of some new coumarin incorporated thiazolyl semicarbazones as anticonvulsants, Acta Pol Pharm., 2009, 66(2), 161-167.
- 20. <u>http://www.drugbank.ca/</u>
- 21. Bassem Sadek, Moawia Mohammad Al-Tabakha, Khairi Mustafa Salem Fahelelbom, Antimicrobial prospect of newly synthesized 1,3-thiazole derivatives, Molecules, 2011, 16, 9386-9396.
- 22. Bondock S., Fadaly W., Metwally M. A., Synthesis and antimicrobial activity of some new thiazole, thiophene and pyrazole derivatives containing benzothiazole moiety, European Journal of Medicinal Chemistry, 2010, 45 (9), 3692-3701.
- 23. Sahin G., Palaska E., Ekizoglu M., Ozalp M., Synthesis and antimicrobial activity of some 1,3,4oxadiazole derivatives, Farmaco., 2002, 57(7), 539-542.

- 24. Gulay Şahin, Erhan Palaska, Melike Ekizoglub, Meral Ozalp, Synthesis and antimicrobial activity of some 1,3,4-oxadiazole derivatives, Il Farmaco, 2002, 57 (7), 539–542.
- 25. Tavares L. S., Silva C. F., deSouza V., daSilva V., Diniz C., Santos M., Strategies and molecular tools to fight antimicrobial resistance: resistome, transcriptome, and antimicrobial peptides, Frontiers in Microbiology, 2013, 4, 2-11.
- 26. Pettersen E. F., Goddard T. D., Huang C. C., Couch G. S., UCSF Chimera- a visualization system for exploratory research and analysis, J. Comput. Chem., 2004, 25, 1605-1612.
- 27. Morris G. M., Huey R., Lindstrom W., Sanner M. F., Autodock4 and AutoDockTools4: automated docking with selective receptor flexibility, J. Computational Chemistry, 2009, 16, 2785-2791.
- 28. Trott O and Olson A. J., AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization and multithreading, J. Comput. Chem., 2010, 31(2), 455-461.
