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Bioaccumulation And Translocation Potential Of Na⁺ And K⁺ In Native Weeds Grown On Industrially Contaminated Soil

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Abstract: Use of the naturally growing plants to remove the inorganic contaminants from the soil as well as water is a sound technology in the recent years. The study was performed in salinity affected soil to examine the efficacy of bioaccumulation potential of sodium and potassium by 12 naturally grown weeds. The wasteland soil was found having higher bulk density (BD), water holding capacity (WHC), electrical conductivity (EC), organic carbon (OC) and nitrogen, phosphorus and potassium (NPK) than the control (garden) soil. The average values of Na⁺ and K⁺ were found in plant parts 750 and 1094 μ g g⁻¹ dry weight respectively. The Na⁺/K⁺ ratio was found highest in the *B. pertusa* (0.8). Translocation factor (TF) ranged from 0.6 to 1.2 for Na⁺ and 0.5 to 0.9 for K⁺. Na⁺ and K⁺ removal from the wasteland soil and waste substrates was most effective with the use of *P. panicum, S. munja, S. spontanum, P. purpureum* and *D. clorata,* because these plants accumulated significant higher amount of Na⁺ and K⁺ in their tissue. These weed plants found to have a great potential for the phytoremediation of industrial wasteland in the investigated area.

Keywords: Bioaccumulation; enrichment factor; industrial area; phytoremediation; salinity; translocation factor.

Introduction

The environmental stresses trigger a wide range of plant responses, ranging from altered gene expression and cellular metabolism to changes in growth, biomass accumulation, and plant productivity¹⁻⁸ and it may also affect the uptake and transport of toxic substances inside the plants. Salinity is a major problem of the soil ecosystem in the arid, semi-arid and coastal regions throughout the world⁹⁻¹¹. The global range of primary salt-affected soils is about 955 M ha, while secondary salinisation affects about 77 M ha, with 58% of these in irrigated areas. About 21.5 million hectare of land is salt affected in Asia alone and India having 8.6 million hectare out of them. The major ions responsible for salinization of soil are Na⁺, K⁺, Ca²⁺, Mg²⁺ and Cl⁻. High salinity is one of the most important environmental stresses impeding crop growth and severely reduces agricultural yields and productivity¹². The soil salinity affects seed germination; alter plants metabolisms through osmotic stress, ion specific effects and oxidative stress¹³. Several reports are available which indicate that many crops have tolerance power towards the salinity e.g. Barley¹² and Chick pea etc.¹⁴. Softum ion can compete with K⁺ and inhibit transport and metabolic processes depending on K⁺. Since Na⁺ is frequently present at higher concentrations than K⁺ in soils, most plants must have transport systems with high selectivity for K⁺ against Na⁺. The objective of present study was planned to evaluate the phytoextraction potential of different naturally growing plant species especially weeds for the reclamation of salinity affected wasteland.

Materials and methods

Site description

The study area is located at Sandila Industrial Area, Uttar Pradesh, India, located between 26° 53' and 27° 46' north latitudes and between 79° 41' and 80° 46' east longitudes (www.mapsofindia.com), average temperature 23°C, average rainfall 970 mm.^{3,5} Certain industries *i.e.* Cotton mill, Vegetable oil mill, Steel Industry (all closed now) have discharged and safe yeast industry, milk powder factory, chemicals factory etc. (all operating), discharging their effluents in the water and soil of the area. The soil was completely barren at the time of sampling was performed during January, May and September month of the year.

Sampling and analysis of metals in soil and plants

On the basis of the existing information and agricultural background 8 locations were selected for soil and plant sampling. Composite soil samples were collected from 0 to 20 cm soil layer, mainly from the root zone (rhizosphere) from each site. The concentrations of metals in soils were determined in the Environmental Laboratory of BBA University of Environmental Science, Lucknow, India. For the metal analysis, soil and plant samples were dried in oven to constant weight and digested in conc. HNO₃ and HClO₄ in 5:1 for soil and 3:1 for plants (v/v) ratio at low temperature till a white residue was obtained. Double distilled water was used to maintain final known volume¹⁵. The samples were analyzed using VARIAN AA240FS make Fast sequentional AAS with flame (FAAS).

Translocation Factor (TF) and Enrichment Coefficient (EC)

The translocation factor (TF) or mobilization ratio was calculated to determine relative translocation of metals from the growing medium to other parts (root and shoot) of the plant species¹⁶⁻¹⁷. The enrichment coefficient (EC) has been calculated to derive the degree of contamination and heavy metal accumulation in growing medium (drain water) and in plants growing on contaminated site¹⁸.

Statistical analysis

All treatments were replicated for six times (n=6). Results were analyzed using One-way ANOVA (SPSS statistical package and MS excel). The difference between treatments were considered significant at p<0.05.

Results and discussion

Soil properties

The bulk density (BD) of wasteland soil was 2.05, which was higher than that of BBA University campus soil used as control. The water holding capacity (WHC) of Sandila industrial area (SIA) soil was 64.17 in comparison to 51.91 observed for the control soil. Soil pH plays an important role in maintaining the availability of nutrients and thus affects the plant growth. The pH of SIA was alkaline in nature (8.42). Electric Conductivity (EC) of the SIA soil was 2.14dS m⁻¹, which was about 5 fold higher than that of the control soil. Organic Matter (OM) is an important component of the soil fertility, which provides nutrients to the vegetation after decomposition. The SIA soil had about 20 fold higher % organic carbon than that of the garden soil. The % organic carbon in wasteland soil indicates higher accumulation of organic substances in the soil. Soil quality was defined in many different ways, ¹⁹ have defined soil quality as the ability of soil to support crop growth, including factors such as tilth, aggregation, organic matter content, soil depth, water holding capacity, infiltration rate, pH changes and nutrient capacity²⁰⁻²³. The water content and availability of ions of a soil solution are, indeed, the major factors affecting its apparent electrical conductivity, which justifies the assessment of salinity from apparent EC measurements²⁴⁻²⁵.

Na concentrations in plant parts

The accumulation of Na in the weeds and their parts naturally grown in the industrial wasteland soil were shown in Table 1. The maximum accumulation of Na was found in *P. panicum* and minimum in *C. arcutus*. The sodium concentration range in the shoot of the weeds were 891.67 to 476.95 ($\mu g g^{-1} d wt$), however, in the roots it was 1158.51 to 601.73 ($\mu g g^{-1} d wt$). The weeds, *S. munja, S. spontanum, D. ciliaris, E. burmanica* were accumulated higher amount of Na in the shoots as compared to the roots.

Potassium concentrations in plant parts

The highest K concentration was measured in *P. panicum* (1298.16 μ g g⁻¹ d wt in plant), while the lowest were in *C. arcuatus* (780.71 μ g g⁻¹d wt). The concentrations of K in plant shoots were ranged from 859.3 to 446.32 μ g g⁻¹d wt with an average value 6.52.5 μ g g⁻¹d wt, whereas in plant root ranged 1900.39 to 1004.1 μ g g⁻¹d wt with an average value 1457.24 μ g g⁻¹d wt (Table 2).

Table 1. Sodium accumulation ($\mu g g^{-1} \mu g g^{-1} d wt$) in weeds and their parts (shoots and roots) growing in industrial wasteland soil

Plants name	Plants ($\mu g g^{-1} d wt$)	Shoot ($\mu g g^{-1} d wt$)	Root ($\mu g g^{-1} d wt$)
B. pertusa	804.02 ± 59.81^{bc}	587.39±46.21 ^{def}	1020.65±73.41 ^b
C.cambogiensis	798.88±51.14 ^{cd}	738.87 ± 40.50^{bc}	858.88 ± 61.78^{cd}
C. arcuatus	614.23±45.79 ^{de}	476.95±37.52 ^f	751.51±54.05 ^{de}
C. dactylon	630.98±46.49 ^{de}	496.66±37.94 ^f	765.3±55.04 ^{de}
C. difformis	731.79±55.70 ^{cd}	$509.91 \pm 38.95^{\rm f}$	953.68 ± 72.45^{bc}
C. rotundus	816.60 ± 60.02^{bc}	598.1±41.41 ^{def}	1035.1 ± 78.64^{ab}
D. aegyptium	725.35±53.25 ^{cd}	550.34±38.10 ^{ef}	900.37 ± 68.40^{bcd}
D.colorata	902.11±70.22 ^{ab}	645.72±52.43 ^{cde}	1158.51±88.01 ^a
D. ciliaris	749.39±61.94 ^{cd}	832.46±73.26 ^{ab}	$666.32 \pm 50.62^{\text{ef}}$
E. burmanica	634.15±52.19 ^{de}	666.56±58.66 ^{cd}	$601.73 \pm 45.71^{\text{f}}$
P.flavidum	643.88±52.02 ^{de}	$514.78 \pm 45.30^{\text{f}}$	772.97±58.72 ^{de}
P. panicum	930.67 ± 65.74^{a}	891.67±61.73 ^a	969.68 ± 69.7^{bc}
S. munja	833.55±58.74 ^{bc}	902.69 ± 62.49^{a}	764.41±54.98 ^{de}
S.spontanum	720.85±51.72 ^{cd}	792.44±56.73 ^{ab}	649.26±46.70 ^{ef}

All the values are means of three replicates \pm S.D. Different letters indicate significant differences between mean at *p*<0.05 (DMRT) analyzed by one way ANOVA

Table 2. Potassium accumulation ($\mu g g^{-1} d wt$) in weeds and their parts (shoots and roots) growing in industrial wasteland soil

Plants name	Plants ($\mu g g^{-1} d wt$)	Shoot ($\mu g g^{-1} d wt$)	Root ($\mu g g^{-1} d wt$)
B. pertusa	996.76±98.59 ^{cd}	756.92±99.90 ^{ab}	1236.6±97.29 ^{bc}
C.cambogiensis	1240.80 ± 114.60^{a}	636.95 ± 84.07^{bcd}	$1844.64 \pm 145.12^{\circ}$
C. arcuatus	780.71±76.28 ^e	557.32±73.56 ^{cde}	$1004.1 \pm 78.99^{\circ}$
C. dactylon	806.58±77.39 ^{cde}	567.55±74.91 ^{cde}	1045.6±79.87 ^c
C. difformis	890.43±89.41 ^{cde}	707.37±96.81 ^{abc}	$1073.49 \pm 82.00^{\circ}$
C. rotundus	1013.46±96.13 ^{bc}	767.76 ± 105.08^{ab}	1259.16±87.17 ^{bc}
D. aegyptium	913.22±85.81 ^{cd}	667.83±91.40 ^{bc}	1158.6±80.20 ^{bc}
D.colorata	1109.36±113.99 ^b	859.3±117.61 ^a	1359.41±110.38 ^b
D. ciliaris	1123.39±110.94 ^b	494.23±67.64 ^{de}	1752.54±154.23 ^b
E. burmanica	924.80±92.29 ^{cd}	446.32±61.09 ^e	1403.28±123.49 ^b
P.flavidum	828.54±86.92 ^{cde}	573.34±78.47 ^{cde}	1083.74±95.374 ^c
P. panicum	1298.16±112.43 ^a	719.12±94.91 ^{abc}	1877.21±129.95 ^a
S. munja	1233.64±103.19 ^a	566.89±74.82 ^{cde}	1900.39±131.56 ^a
S.spontanum	1074.90±91.49 ^{bc}	481.5±63.55 ^{de}	1668.3±119.43 ^a

All the values are means of three replicates \pm S.D. Different letters indicate significant differences between mean at *p*<0.05 (DMRT) analyzed by one way ANOVA.

Potassium (K^+) is an essential element for plant growth and is an extremely dynamic ion in the soil system. As anion, potassium is highly mobile in the plant system but only moderately mobile in the soil system ²⁶⁻²⁷. Along with other elements like chlorine, sodium and lithium, potassium is called a non-constitutive element, as it does not form component/compounds in the plant system²⁸. Potassium plays an important role to

enhance tolerance capacity of the plants such as rice²⁹, tomato³⁰⁻³¹, cucumber and pepper³² and strawberry³³. Under salinity stress, potassium translocated back to the root in excess of the plant's requirements³⁴. One author has reported synergy between cation (K⁺ or Ca²⁺) accumulation and myo-inositol for breaking of the symmetry of bud growth³⁵. The results of the present study showed a variation in sodium and potassium levels within plan species. For the relationship between salt tolerance and ion effects, it was suggested that plant species differ in the degree of Na⁺ and K⁺ toxicity affecting their growth³⁶. Salinity not only caused high Na⁺ accumulation in plants but also influenced the uptake of essential nutrients such as K⁺ and Ca²⁺ through the effects of ion selectivity. Decline in K⁺ accumulation because of salinity stress, has been widely reported in many crops like wheat, sorghum etc.³⁷⁻³⁸. In those species that retain Na⁺ in woody roots or stems, there is a strong correlation between Cl⁻ exclusion and salt tolerance. This indicates the higher ratio of Na⁺/K⁺ resulted in the decrease in the growth rates.

Enrichment coefficient (EC)

The Enrichment coefficient (EC) values of all studied samples for Na and K which varied between 0.30 to 1.30 for Na and 0.20 to 0.65 for K. The highest EC were observed in *S. spontanum* and *D. ciliaris* for Na and for K in *D. colorata* and *C. difformis*. Enrichment coefficient is an important factor when considering the phytoremediation potential of a plant species $^{39.40}$. The enrichment coefficient>1 shows a special ability of the plant to absorb metal ions from soils and transport it to aerial parts⁴¹. Plants can immobilize heavy metals through absorption and accumulation by roots, adsorption onto roots, or precipitation within rhizosphere⁴².

Translocation factors (TF) of Na and K in weeds grown in industrial wasteland soil

The translocation factors for Na were ranged from 0.70 to 1.20, with the highest value in the sample of *P. panicum* and *S. munja* while the lowest was found in *C. arcuatus*. A plant's ability to translocate metals from the roots to the shoots is measured using the TF, which is defined as the ratio of metal concentration in the shoots to the roots. TF higher than 1 indicates a very efficient ability to transport nutrients from roots to shoots, most likely due to efficient metal transport systems⁴³. The results indicated that the translocation factors of plant species were higher than 1 in this study (Fig.1). Na can be toxic to photosynthetic activity, chlorophyll synthesis and antioxidant enzymes if available in higher amount⁴⁴. Translocation factors for K ranged from 0.45 to 0.90, with the highest value in the sample of *D. colorata* and the lowest in *E. burmanica* (Fig. 2). Tolerant plants tend to restrict soil–root and root–shoot transfers, and therefore have much less accumulation in their biomass, while hyperaccumulators actively take up and translocate metals into their aboveground biomass.

In the present investigation, 12 different plant species naturally grown on the industrially contaminated soil was studied and found *C. cambogiensis, D. aegyptium and D. ciliaris* species were found efficient for Na and K reclamation from contaminated soils. These plants can be grow easily on contaminated wasteland soil and subsequently may be used as a raw material for commercial extraction of metals.



Figure 1. Translocation factors of Na in weeds grown in industrial wasteland soil



Figure 2. Translocation factor of Na in weeds naturally grown in industrial wasteland soil.

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References

- 1. Jaleel CA, Manivannan P, Kishorekumar A, Sankar B, Gopi R, Somasundaram R, Panneerselvam R. Alterations in osmoregulation, antioxidant enzymes and indole alkaloid levels in *Catharanthus roseus* exposed to water deficit. Colloids and Surfaces B: Biointerfaces, 2007 59: 150–157.
- Farooq M, Basra SMA, Wahid A, Cheema ZA, Cheema MA Khaliq A. Physiological role of exogenously applied glycinebetaine in improving drought tolerance of fine grain aromatic rice (*Oryza sativa* L.). J Agron Crop Sci., 2008, 194: 325–333.
- 3. Ghavri SV, Singh RP. Phytotranslocation of Fe by biodiesel plant *Jatropha curcas* L. grown on iron rich wasteland soil. Braz J Plant Physiol., 2010, 22(4): 235-243.
- 4. Ghavri SV, SK Rawat, RP Singh. Comparative study of growth and survival rate of *Jatropha curcas* clones (BTP-A, BTP-N and BTP-K) in the contaminated waste land soil from Sandila Industrial Area (SIA). Pollut Res., 2010, 29(3): 519-522.
- 5. Ghavri SV, Singh RP. Growth, biomass production and remediation of copper contamination by *Jatropha curcas* plant in industrial wasteland soil. J Environ Biol., 2012, 33: 207-214.
- 6. Bauddh K, Singh RP. Differential toxicity of cadmium to mustard (*Brassica juncea* L.) genotypes is not maintained at higher metal levels. J Evniron Biol., 2011, 32: 355-362.
- 7. Bauddh K, Singh RP. Cadmium tolerance and its phytoremediation by two oil yielding plants *Ricinus communis* (L.) and *Brassica juncea* (L.) from the contaminated soil. Int J Phytorem., 2012, 14: 772-785
- Bauddh K, Singh RP. Growth, tolerance efficiency and phytoremediation potential of *Ricinus communis* (L.) and *Brassica juncea* (L.) in salinity and drought affected cadmium contaminated soil. Ecotox Environ Safety, 2012, 85:13-22 DOI:10.1016/j.ecoenv.2012.08.019.

- Singh RP, Dhania G, Sharma A, Jaiwal PK. Bio-technological approaches to improve phytoremediation efficiency for environments contaminants. In: Environmental bioremediation technologies (Eds.: S.N. Singh and R.D. Tripathi), 2007, p. 223-258. Springer Verlage Berlin Heidelberg.
- 10. Demir I, Mavi K. 2008. Effect of salt and osmotic stresses on the germination of pepper seeds of different maturation stages. Braz Arch Boil Technol. 51(5):897-902.
- 11. Sima NAKK, Askari H, Mirzaei HH, Pessarakali M. Genotypic differential responses of three forage species to calcium supplement in saline conditions. J Plant Nutr., 2009, 32: 579-597.
- 12. Flowers TJ, Hajiabagheri MA. Salinity tolerance in Hordeum vulgare: ion concentration on root cells of cultivars differing in salt tolerance. Plant soil, 2001, 231: 1-9.
- 13. Bhaskar P, Bauddh K, Singh RP. Differential response of two high yielding cultivars of Indian mustard (*Brassica juncea* L.) to NaCl salinity during seed germination and early seedling growth. J Ecophysiol Occup Hlth., 2009, 9: 137-144.
- 14. Kaya M, Kaya G, Kaya MD, Atak M, Saglam S, Khawar KM, Ciftci CY. Interaction between seed size and NaCl on germination and early seedling growth of some Turkish cultivars of chickpea (*Cicer arietinum* L.). J Zhejiang Univ Sci B., 2008, 9: 371-377.
- 15. Fritioff A., Greger M. 2007. Functions as a proton-coupled symporter for phytosiderophore- and nicotianamine-chelated metals. J Biol Chem., 2007, 279: 9091–9096.
- 16. Barman SC, Sahu RK, Bhargava SK, Chaterjec C. Distribution of heavy metals in wheat, mustard and weed grown in field irrigated with industrial effluents. Bull Environ Contam Toxical., 2000, 64: 489-496.
- 17. Gupta S, Nayek S, Saha RN, Satpati S. Assessment of heavy metal accumulation in macrophyte, agricultural soil and crop plants adjacent to discharge zone of sponge iron factory. Environ Geol., 2008, 55: 731-739.
- 18. Kisku GC, Barman SC, Bhargava SK. Contamination of soil and plants with potentially toxic elements irrigated with mixed industrial effluent and its impact on the environment. Water Air Soil Pollut., 2000, 120:121-137.
- 19. Power JF, Meyers RJK. The maintenance or improvement of farming systems in North America and Australia. In: Stewart J.W.B. (ed.): Soil quality in semi-arid agriculture. In: Proc. Int. Conf. Univ. Saskatchewan, Saskatoon, Canada: 1989, 273–292.
- 20. Loveland P, Webb J. Is there a critical level of organic matter in the agricultural soils of temperate regions: a review. Soil Till Res., 2003, 70: 1–18.
- 21. Chaplain V, Defossez P, Delarue G, Richard G, Tessier D, Roger-Estrade J. Effects of grassland on bulk density of soil and mechanical resistance. Geophys. Res Abstracts., 2011, 13: 2011-3121.
- 22. Grassini P, Jinsheng Y, Kenneth G, Hubbard, Kenneth G, Cassman. Soil water recharge in a semi-arid temperate climate of the Central U.S. Great Plains Agricult. Water Manage., 2010, 97: 1063-1069.
- 23. Piedallu C, Jean-Claude G, Bruand A, Seynave I. Mapping soil water holding capacity over large areas to predict potential production of forest stands. Geoderma, 2011, 160: 355-366.
- 24. Corwin DL, Lesch SM. Characterizing soil spatial variability with apparent soil electrical conductivity I. Survey protocols. Comp Elect Agri., 2005, 46: 103-133.
- 25. Friedman SP. Soil properties influencing apparent electrical conductivity: A review Computers and Electronics in Agriculture, 2005, 46: 45-70.
- 26. Loo AY, Jain K, Darah I. Antioxidant activity of compounds isolated from the pyroligneous acid *Rhizophora apiculata*. Food Chem., 2008, 107:1151-1160.
- 27. Mirhosseini H, Amid BT. A review study on chemical composition and molecular structure of newly plant gum exudates and seed gums. Food Res Int., 2012, 46: 387-398.
- 28. Ranade-Malvi U. Interaction of micronutrients with major nutrients with special reference to potassium. Karnataka J Agric Sci., 2011, 24: 106-109.

- 29. Bohra JS, Doerffling K. Potassium nutrition of rice (*Oryza sativa* L.) varieties under NaCI salinity. Plant Soil, 1993, 152: 299-303.
- 30. Kaya C, Kirnak H, Higgs D. Enhancement of growth and normal growth parameters by foliar application of potassium and phosphorus on tomato cultivars grown at high salinity (NaCl). J Plant Nutr., 2001, 24: 357-367.
- 31. Yurtseven E, Kesmez GD, Nlukara AU. The effects of water salinity and potassium levels on yield, fruit quality and water consumption of a native central Anatolian tomato species (*Lycopersicon esculantum*) Agr Water Manag., 2005, 78: 128-135.
- 32. Kaya C, Kirnak H, Higgs D. Effects of supplementary potassium and phosphorus on physiological development and mineral nutrition of cucumber and pepper cultivars grown at high salinity (NaCl). J Plant Nutr., 2001, 24: 1457-1471.
- 33. Kaya C, AK BE, Higgs D. Response of salt-stressed strawberry plants to supplementary calcium nitrate and/or potassium nitrate. J. Plant. Nutr., 2003. 26(3): 543-560.
- 34. Daglia M, Stauder M, Papetti A, Signoretto C, Giusto G, Canepari P, Pruzzo C, Gazzani G. Isolation of red wine components with anti-adhesion and anti-biofilm activity against *Streptococcus mutans*. Food Chem., 2010, 119: 1182-1188.
- 35. Desbiez MO, Ripoll C, Pariot C, Thellier M. 1991. Elicitation of developmental processes in higher plants by hexoses or myo-inositol in the presence of potassium or calcium. Plant Physiol Biochem. 29:457–462.
- 36. Shan Q, Liu X, Zhang J, Chen G, Liu S, Zhang P, Wang Y. Analysis on the tolerance of four ecotypes plants against copper stress in soil. Pro Environ Sci. 10, Part B, 2011,1802-1810.
- 37. Pecoraro N, Dallman MF, Warne JP, Ginsberg AB, Laugero KD, Susanne E, Hani H, Gomez F, Bhargava A, Akana SF. From Malthus to motive: How the HPA axis engineers the phenotype, yoking needs to wants. Prog Neurobiol., 2006, 79: 247-340.
- 38. Lenz CA, Ferstl CMH, Vogel RF. Sub-lethal stress effects on virulence gene expression in Enterococcus faecalis. Food Microb., 2010, 27: 317-326.
- 39. Castañeda SS, Sucgang RJ, Almoneda RV, Mendoza NDS, David CPC. Environmental isotopes and major ions for tracing leachate contamination from a municipal landfill in Metro Manila, Philippines. J Environ Radio., 2012, 110: 30-37.
- 40. Naji S, Seyed MA, Karazhiyan H. Effect of thermal treatments on functional properties of cress seed (*Lepidium sativum*) and xanthan gums: A comparative study. Food Hydroco., 2012, 28: 75-81.
- 41. Djenontin TS, Wotto, VD, Avlessi F, Lozano P, Dominique KC, Pioch D. Composition of *Azadirachta indica* and *Carapa procera* (Meliaceae) seed oils and cakes obtained after oil extraction. Ind Crop Prod., 2012, 38: 39-45.
- 42. Taskila S, Tuomola M, Ojamo H. 2012. Enrichment cultivation in detection of food-borne Salmonella. Food Cont. 26:369-377.
- 43. Zhao GQ, Ma BL, Ren CZ. Growth, gas exchange, chlorophyll fluorescence, and ion content of naked oat in response to salinity. Crop Sci., 2007, 47: 123–131.
- 44. Yamashita M, Tomita-Yokotani K, Hashimoto H, Sawaki N, Notoya M. Sodium and potassium uptake of Ulva Application of marine macro-algae for space agriculture. Adv. Space Res., 2009, 43: 1220-1223.