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Optimization Studies for Defluoridation of Water using Aspergillus niger Fungal Biosorbent

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Abstract: Fungal biosorbent prepared from *Aspergillus niger* for removal of fluoride was investigated for the various factors influencing defluoridation. Calcium and alkali treated biomass was effective in removal of fluoride. The extent of defluoridation was dependent on the initial pH of fluoride containing water and decreased with increasing pH. The capacity was found to be between 30% at pH 5.0 and 21% at pH 8.0. Fluoride removal decreased with increased bicarbonate concentration, but was independent of the presence of chloride and sulphate. The kinetics of fluoride removal exhibited a rapid phase of binding for a period of 1.0 hour and a slower phase of binding during the subsequent period. The potential use of this fungal biosorbent for biodefluoridation is discussed in this communication. **Keywords:** Fluoride, Biomass, Biosorption, *Aspergillus niger*.

Introduction

Excess amount of fluoride in drinking water has been known to cause adverse effects on human health. Membrane separation techniques were investigated for the effective separation of fluoride¹. Garmes et al. $(2002)^2$ have investigated a hybrid process containing adsorption and dialysis for defluoridation of water. Adsorption process was reported to be effective,

environmental friendly and economical³. Use of biosorbents/biomass from various microbial sources, leaf based sorbents, and water hyacinth, for fluoride removal was reported by various investigators^{4,5,6}. Laxmaiah et al (2002)⁷ have used fungal biosorbent for removing of fluoride from water. Apart from this report, not much information regarding the use of fungal biosorbents for removal of fluoride are not

available. Hence, the present investigation was done and the results are discussed.

Material and Methods

The Aspergillus niger strain used in these experiments was isolated from Mahatma Gandhi University Campus, Nalgonda, A.P., India. The alkali extracted biomass of A.niger (biosorbent) was prepared according to the method of Akthar et al (1995)⁸. Finely powdered biosorbent was first washed with glass-distilled water and then suspended in fluoride containing waters. The Biosorbent (1 g) was suspended in 100 mL glassdistilled water or tap water and pH adjusted to 7.0, stirred and centrifuged. The supernatant was discarded and the process repeated two more times. The final pellet was suspended in 100 mL (200µg/L) of fluoride containing water. After specified period of exposure, the suspension was centrifuged and the fluoride content of the supernatant determined. 1.0 g of dried biomass was suspended in test tubes containing10 ml of aqueous CaCl2, sodium bicarbonate solution in the concentration varying from 40 to 160 µg/ml and allowed to stand for 5 hours after putting it mixed for 5 minutes on rotor. Influence of the aqueous phase pH on fluoride adsorptive uptake was studied by adjusting the reaction mixture to different initial pH values from 3.0 to 8.0 and analyzed for residual fluoride after equilibrium contact time. The fluoride content of the supernatants was determined colorimetrically using SPADNS method (APHA, 1998)⁹.

Results and Discussion:

The surface characteristics of the fungus are responsible for sorptive defluoridation. The sorption is characterized under different experimental conditions and discussed. Perusal of Table 1 shows the ability of the biosorbents to bind to fluoride ions. Initially 2.0 hrs incubation was tested to check the amount of biosorbent which would be ideal for binding to the fluoride. Biosorbent concentration at 1.0 g could bind about 20% of fluoride present in the water. More amounts of the biosorbent could not enhance the absorption of fluoride and hence 1.0 g of the biosorbent was used for further investigation. Effect of time, that is duration of exposure to the fluoride containing water was also studied. Table 2 shows the sorption profile, which clearly indicates that the sorption process attains equilibrium in 12 hrs Twelve hours of incubation could bind about 25% of the total fluoride present in water. Although there was a slight increase in the absorption, it was not much. Hence, 12 hrs incubation was use for further studies. The total amount of fluoride removed was found to be 26% at the end of twenty hours incubation.

Mass dependence and pH influence on the biodefluoridation by the biosorbent was investigated (Table 3). At pH 5.0, 30% of fluoride was bound to the biosorbent while at pH 8.0 21% of the biosorbent was bound. With increase in pH the ability of the sorbent to bind to the fluoride ions gradually decreased. Overall acidic pH was amenable than basic pH for removal of fluoride. Lower binding at higher pH could be due to competition between F- and OHfor fluoride binding sites. The mechanism of fluoride binding by the biosorbent is not clear and may be due to the protonation of primary amino groups at acidic pH which could bind to fluoride.

Temperature can also effect the rates of defluoridation. Temperature at 30°C was found to bind more amounts of fluoride. About 35% of the fluoride was absorbed at this temperature. Increase in temperature above 30°C resulted in decrease in the rates of defluoridation. Only 25% of fluoride could be removed with increase in the temperature upto 50°C. The effects of coexisting anions such as chloride, sulphate and bicarbonate on fluoride adsorption by the fungal adsorbent were examined and the results are given in Table 5. Chloride and sulphate did not perceptibly interfere with fluoride removal at a concentration of 160 µg/L. However, bicarbonate showed great competitive adsorption with fluoride. The fluoride adsorption amount decreased quickly from 36 to 27% with the increase of bicarbonate concentration from 40 to 160 μ g/L. This may be

due to the competition of bicarbonate ions with the fluoride ions at the active site present on the surface of the sorbents. The order of interference for fluoride removal observed as in the following order, HCO3->SO42-, Cl- for the adsorbent

The present results indicated that the addition of co-ions, in the concentration ranges investigated had no appreciable effect on the amount of fluoride ions removed by these adsorbents except for bicarbonates.

Biosorbent	pН	Duration of	Fluoride	Capacity
(conc in g)		exposure	in water	(µg F-/g
		(hours)	(µg/L)	biosorbent)
0.25	7.0	2.0	200	26
0.50	7.0	2.0	200	32
0.75	7.0	2.0	200	34
1.0	7.0	2.0	200	38
1.5	7.0	2.0	200	38
2.0	7.0	2.0	200	38
2.5	7.0	2.0	200	38
3.0	7.0	2.0	200	38

Table 1; Effect of adsorbent concentration on binding of fluoride

Table 2: Time course of fluoride binding by the biosorbent

Biosorbent	рН	Duration of	Fluoride	Capacity
(conc in g)		exposure	in water	(µg F-/g
		(hours)	$(\mu g/L)$	biosorbent)
1.0	7.0	1.0	200	36
1.0	7.0	2.0	200	38
1.0	7.0	4.0	200	42
1.0	7.0	6.0	200	45
1.0	7.0	8.0	200	45
1.0	7.0	10.0	200	48
1.0	7.0	12.0	200	52
1.0	7.0	14.0	200	53
1.0	7.0	16.0	200	53
1.0	7.0	18.0	200	52
1.0	7.0	20.0	200	52

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Biosorbent	pН	Duration of	Fluoride	Capacity
(conc in g)		exposure(hours)	in water(µg/L)	(µg F-/g biosorbent)
1.0	3.0	12	200	54
1.0	3.5	12	200	54
1.0	4.0	12	200	58
1.0	4.5	12	200	64
1.0	5.0	12	200	66
1.0	5.5	12	200	60
1.0	6.0	12	200	58
1.0	6.5	12	200	54
1.0	7.0	12	200	50
1.0	7.5	12	200	46
1.0	8.0	12	200	42

Table 3: Effect of pH on fluoride binding by the biosorbent

Table 4: Effect of temperature on removal of fluoride

Biosorbent	Temp	pН	Duration of	Fluoride	Capacity
(conc in g)	(°C)		exposure(hours)	in water(µg/L)	(µg F-/g biosorbent)
1.0	25	5.0	12	200	68
1.0	30	5.0	12	200	70
1.0	35	5.0	12	200	64
1.0	40	5.0	12	200	66
1.0	45	5.0	12	200	58
1.0	50	5.0	12	200	50

Biosorbent	Coions	Duration of	Fluoride	Capacity
(conc in g)	(µg/ml)	exposure(hours)	in water(µg/L)	(µg F-/g biosorbent)
	Chloride			
1.0	40	12	200	70
1.0	80	12	200	72
1.0	120	12	200	70
1.0	160	12	200	72
	Sulphates			
1.0	40	12	200	68
1.0	80	12	200	70
1.0	120	12	200	68
1.0	160	12	200	70
	Bicarbonates			
1.0	40	12	200	72
1.0	80	12	200	68
1.0	120	12	200	62
1.0	160	12	200	54

Conclusions:

Aspergillus niger used in this study could clearly remove fluoride at a rate of 36%. The present results reported here agree with those of Laxmiah et al $(2002)^7$ where alkali-treated fungal biosorbent was used for defluoridation. The organic matrix of the biosorbent contains Ca++ ions after treatment with calcium. It is possible that the Ca++ ions are responsible for binding fluoride. It may be possible that cations on the surface of biomass may be used to removal anions that are not usually removed as the cell envelops carry negative charges or their surface. The mechanism of binding is not clear and further work is required to understand the molecular aspects for large scale defluoridation of water supplies. Hence, biosorption can therefore provide a solution to control fluoride pollution

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