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Stability Indicating Thin-Layer Chromatographic Determination of Brivarecetam as Bulk Drug: Application to Forced Degradation Study

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Abstract : A new, accurate, selective, simple and precise HPTLC method for estimation of briveracetam in bulk drug as well as tablet dosage form was developed and validated. The drug was well separated using mobile phase of ammonium acetate: methanol:n-propanol (8:1.6:1.6 v/v/v) with densitometric quantification of brivaracetam at 242nm. The TLC parameters were standardized and the Rfof brivaracetam was determined to be 0.40. The values of Linearity (200-1200ng/spot), Method precision (intra-day RSD 0.5-0.8% and inter-day RSD 0.25-0.46%), Accuracy (% recovery 97.53%-102.84%) and specificity were determined according to ICH guidelines. Brivaracetam was exposed to various stress condition to study degradation profile. Degradation was seen in acidic, basic and oxidative condition. Brivaracetam was found to be stable in photolytic condition. The experiment gives us satisfactory result for method validation and development indicates the successful validation of HPTLC method for quantitative determination of brivaracetam. The HPTLC method is simple, rapid, economic andmore suitable for routine analysis of brivaracetam in bulk and tablet dosage forms. **Keywords :** Brivaracetam, HPTLC, Densitometric estimation, Method development, Validation, Stability indicating method.

Introduction

Brivaracetam is a third-generation antiepileptic racetam derivative and a 4-n-propyl analogue of levetiracetam [Fig.1]. It was granted Food and Drug Administration (FDA) approval as an add-on therapy in February 2016.Brivaracetam is the drug which binds to the synaptic vesicle glycoprotein 2A (SV2A).¹⁻³In phase II clinical trials the adult patients shows promising effect on refractory partial seizures. Results of stage III trials have been recorded; it is proved that brivaracetam is 10 times more potent for the prevention of certain types of seizure in mouse models than levetiracetam, of which it is an analogue. It shows activity as a novel high-affinity synaptic vesicle protein 2A (SV2A) ligand, displays inhibitory activity at neuronal voltage-dependent sodium channels, data which are collected from animal models suggested potent and broad-spectrum antiepileptic activities.⁴⁻⁶

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There is no known information in the literature about development and validation of brivaracetam by HPTLC. Literature survey reveals data about brivaracetam viz. pharmacokinetics and metabolism of 14C-brivaracetam⁷, metabolism studies of brivaracetam and gemfibrozil⁸, clinical trials of adjunctive brivaracetam for refractory partialonsetseizures⁹, identification of drug metabolites in human plasma or serum integrating metabolite prediction by LC–HRMS methods, HPLC Studies on degradation behavior of brivaracetam¹⁰, a meta-analysis of placebo-controlled studies and development of validated stability indicating method by HPLC and UPLC of brivaracetam¹¹⁻¹².

Currently, HPTLC is used for the identification and quantification of pharmaceuticals. HPTLC offers several advantages include abbreviated analysis time, parallel processing of samples, minimal maintenance cost, modest consumption of mobile phase, etc. Because of the high sensitivity and its utility HPTLC test results was also included in pharmacopoeia; thus HPTLC getting wide acceptance.¹³⁻¹⁶

The main aim was to develop simple, precise and robust analytical procedure with degradation studies for estimation of brivaracetam and formulation by HPTLC. The proposed method was validated as per ICH guidelines.¹⁷⁻¹⁸



Figure 1: Structure of brivaracetam

Materials and Methods

Reagents and chemicals

Brivaracetam (99.85%) of pharmaceutical grade was procured from Manus Akttevva BiopharmaLLP, Ahmedabad, India. Silica gel 60F $_{254}$ TLC plates (10×10 cm, layer thickness 0.2 mm, Merck, Germany) were used as stationary phase. The pharmaceutical dosage form (Brivact, tablet 25mg, Sanofi India) was procured from local market. All chemicals and reagents were of analytical grade were purchased from Merck (Pvt) Ltd.

HPTLC Instrumentation and optimization of chromatographic condition

Pre-coated aluminum plates of silica gel $60F_{254}$ plates (10 cm ×10cm) with 250µm thickness;(E. Merk, Darmstadt, Germany), supplied by Anchrom technologist, Mumbai were used as stationary phase. The standard solutions were spotted in the form of bands and were applied with the help of Camag 100µl sample (Hamilton, Bonaduz, Switzerland) syringe, on the silica gel plates, using a CamagLinomat V (Switzerland) sample applicator. Prewashing of plates was done with methanol and activated at 110°C for 5min in oven prior to chromatography. Bands were applied with constant application rate of 0.1µl/s with the spaceof 6mm. Thedimensionof slit was kept at (5mm×0.45mm) and the scanning speed was 10mm/s. Baseline correction was done and each track was scanned three times.

Optimization of mobile phase

Various solvent systems were tried like mixture of a) Methanol: acetonitrile(7:3 v/v) b) Acetonitrile: methanol: toluene (2:2:1v/v/v) c) Toluene: ethyl acetate: methanol (3:1:1 v/v/v) d) Toluene: methanol: ammonia (1:3:0.5 v/v/v) e) Butanol: water (4:1 v/v) f) Ammonium sulphate 0.5% : methanol (8:2 v/v) to separate and resolve spot of brivaracetam from its impurities and other excipients of formulation. The mixture of ammonium acetate: methanol (8:1.6 v/v) could resolve brivaracetam but R_f value was found in range of (6 to

7). Hence to improve R_f value n-propanol 1.6 mlwas added in the mixture. The mixture of ammonium acetate: methanol: n propanol showed satisfactory separation and gives well resolved peak. R_f value of drug was found to be 0.40 ±0.03.

Development of plates was carried out as linear ascending direction in a $(20 \text{ cm} \times 10 \text{ cm})$ twin trough glass chamber (Camag, Muttenz, Switzerland) saturated with the mobile phase. The optimized time period required for chamber saturation was 20min at room temperature $(25^{\circ}\text{C}\pm2)$ and relative humidity of $60\%\pm5$.

Forced Degradation Studies

Acid and Base Induced Degradation

Acid and base hydrolysis studies were carried out by exposing drug solution to 0.01N HCl and 0.01N NaOH solution refluxed at 50°C for 15 mins. These solutions was applied silica plate and developed in the mobile phase. Changes were observed on the denisitogram.

Hydrogen Peroxide-Induced Degradation

The sample was treated with 3% hydrogen peroxide solution for 2 hours at room. This solution was applied on silica plate and developed in the mobile phase. Changes were observed on the denisitogram.

Photochemical Degradation

Stock solution (1000 μ g/ml) was exposed to direct sunlight for 48 hrs. This solution was applied on silica plate and run the mobile phase. Changes were observed on the denisitogram. The photochemical study was also carried out in photostability chamber for 48 hour.

Experimental

Preparation of standard stock solutions

For the preparation of stock solution accurately weighed 10mg of brivaracetam was transferred to a 10ml volumetric flask and was diluted with methanol ($1000\mu g/ml$). From this stock solution0.1ml was pipetted out and was diluted with 10ml methanol ($10\mu g/ml$).

Prewashing of plates

The pre-coated silica gel $60F_{254}$ plates (10 cm ×10cm) from E. Merck were pre-washed with methanol, dried and activated for 30min at 110°C.

Sample application

The standard and formulation samples of brivaracetam were spotted in the form of bands of length 6mmon pre-coated silica gel TLC plates. With distance of 10mm from the bottom and left margin and 10mm distance between two bands. Samples were applied with the help of continuous drying stream of nitrogen gas at constant application rate of 150nl/s.

Method Validation

As per ICH guidelines Q2 9(R1)the developed HPTLC method was validated for various parameters such as linearity, accuracy, precision, limit of detection, limit of quantification, repeatability, specificity and robustness.

Linearity and calibration curve

In linearity the method was evaluated with the help ofsix concentration levels of calibration curves. Aliquots of standard working solution of different concentration in the range 200-1200ng/spot of brivaracetam

were spotted on the plate. The calibration curve was plotted peak area vs concentration by using Win-CATS software. After 20min chamber saturation time was over the plate was developed in twin trough glass chamber. The length of the run was 80mm. The experiment was repeated three times to generate the calibration equation and then least square regression analysis was performed.

Precision

To evaluate intra-day precision, samples were analyzed on same day. Three different samples of three concentrations(200ng/spot, 400ng/spot and 800ng/spot) were analyzed. The inter-day precision study was carried out by comparing the data of three different days.

Repeatability

Repeatability of sample application was ensured by spotting six spots of 600ng/spot standard drug solution on a TLC plate at different times on same day with the help of sample applicator, followed by development of plate and recording of the peak areas for six spots.

Accuracy

For determining accuracy of the developed method recovery studies of the drug were carried out. Accuracy was assessed by mixing known quantity of standard drug'Briviact' which is manufactured by Sanofi India, and the contents were analyzed by the standard method. Recovery studies were carried out at 80-120% levels. Percentage recovery and percentage RSD were then calculated.

Limit of detection and limit of quantitation

To calculate the limit of detection (LOD) and limit of quantitation (LOQ) methanol was spotted six times. LOD studies were carried out by taking different concentrations as 20, 40, 60, 80, 100ng/spot. Spot was not detected up to concentration 40ng/spot. The peak was observed at 60ng/spot and a signal-to-noise ratio is 3:1. The LOQ was performed by taking different concentrations as 20, 40, 60, 80, 100ng/spot. The peak was observed with quantifiable area at 600ng/spot with a signal-to-noise ratio of 10:1.

Specificity

To confirm the specificity, brivaracetam was spotted on TLC plate, developed and scanned. The standard UV spectrum of brivaracetam was compared with spectrum of brivaracetam extracted from tablet. The peak purity of brivaracetam was assessed by comparing their respective spectra at different position of spot.

Robustness

For the study of robustness various parameters were selected viz. mobile phase composition, chamber saturation time and solvent migration distance. The small change in one factor was introduced and the effect on the results was examined.

Result and discussion

Optimization of mobile phase

The most suitable mobile phase is ammonium acetate: methanol: n-propanol (8:1.6:1.6 v/v/v). It gave better resolution and better reproducibility in migration of brivaracetam [Fig.2].

Forced degradation condition	R_{f} valve of degradation products	% Degradation	% Drug	Figure
Acid, 0.01 N HCl, 50°C	Peak 1- 0.03	6.1±0.29	96.9±0.24	Fig.3 A
for 15 min	Peak 2- 0.47			
Alkaline 0.01 N NaOH,	Peak 1- 0.06	8.8±1.12	91.2±1.01	Fig.3 B
50°C for 15 min	Peak 2- 0.47			
	Peak 3- 0.84			
H_2O_2 3% w/v, 2 hrs	Peak 1- 0.06	12.9±2.7	90.1±2.15	Fig.3 C
	Peak 2- 0.47			
	Peak 3- 0.62			
	Peak 4- 0.64			





Figure 2: Densitogram of brivaracetam formulation (800 ng/spot)



A.Densitogram of Acid Degradation



B. Densitogram of Base Degradation



(A- Acid induced Degradation, B- Base Induced Degradation, C- Oxidative Degradation)

C. Densitogram of Oxidative Degradation



Forced Degradation Studies

A forced degradation study was carried out. Degradation was observed in acid, alkaline and oxidative conditions. The drug was stable in photochemical studies.[Fig.3] [Table 1]

Method validation

Linearity

The calibration curve was plotted as peak area of compound against the concentration over the range of 200 to 1200ng/spot. The slope, intercept and correlation co-efficient value was calculated and it is given in [Table 2]. Graph showed that their exhibits a good correlation between regression coefficient and concentration of the drug [Fig.4].





Figure 4: Calibration curve of brivaracetam (200-1200ng/spot)

Table 2: Linearity and range

Linearity and range	Brivaracetam
Range (ng/spot)	200-1200
Regression coefficient (r ²)	0.997
Linearity equation	y = 3.793x - 43.7

Precision

The intra-day and inter-day was performed and relative standard derivations were found in the range of 0.54-0.87% and 0.25-0.46% respectively. Smaller the values of intra-day and inter-day variation in the analysis indicated that the method was precise. [Table 3]

Table 3:Intra-day and Inter-day precision study (n=3)

Amount	Intra-day precision		Inter-day precision	
(ng/spot)	Mean area	%RSD	Mean area	%RSD
40	1617	0.8749	1615	0.2501
60	2271	0.5468	2231	0.3731
80	2952	0.5420	2948	0.4629

Table 4: Repeatability study

Amount (ng/spot)	Peak area	%RSD
	2214	0.5989
	2244	
60	2246	
00	2235	
	2250	
	2246	

Repeatability

In repeatability study of sample application, the %RSD for the peak area of given sample was found to be 0.59%. The RSD value is for the measurement of peak area and sample application, both of these are the instrumental specifications (i.e.1%); which ensure the proper functioning of the system. [Table 4]

Accuracy

The % recovery of brivaracetam was found to be 97.53, 99.86, and 102.84% (at 80%, 100% and 120% respectively). The recovery studies results indicated that the given method was accurate for estimation of drug in a tablet dosage form [Table 5].

Table 5: Recovery studies of brivaracetam tablet (n=3)

%Level	Concentration of drug added	Concentration of drug	%	Average Recovery
	ng/spot	found	Recovery	(%)
80	324	316	97.53	
100	360	359	99.86	100.07
120	396	407	102.84	
	1.0.0			

LOD and LOQ

The LOD gives a measurable response of signal to noise ratio of 3:1. The LOD for brivaracetam found to be 60ng/spot. The LOQ gives response that can be accurately quantified (signal to noise ratio of 10:1). The LOQ was found to be at 600ng/spot for brivaracetam. Therefore it was conclude that the developed method was sensitive.

Analysis of formulation

The content of the brivaracetam tablet (Sanofi India, Powai, Mumbai) was calculated from the peak area which were recorded from densitogram. Analysis of brivaracetam(25mg)tablet was performed and % label claim was found to be 98.64%. [Fig.5] [Table 6]

Table 6: Analysis of formulation

% KSD
0.46

*n= no of times procedure repeated



Figure 5: Densitogram of brivaracetam tablet sample





Figure 6: UV spectrum of standard brivaracetam

Specificity is the method for analyzing Rf value and spectra pattern of drug. The good correlation among spectra gives the peaks at start (s), apex (m), and end (e) indicates the peak purity of brivaracetam [correlation r (s, m) = 0.20, 0.26, r (m, e) =0.26, 0.31]. Therefore it can be assured that no impurities or degradation products migrated with the peaks obtained from standard solutions of the drug. It was observed that excipients present in formulation did not interfere with peak of drug ($R_f 0.40\pm003$). Hence, above HPTLC method was found to be specific. [Fig.6]

Robustness

Robustness is the effect of operational parameters examination on the analysis results. Small changes in mobile phase composition were introduced and deviation was observed. 2% RSD deviation was observed which confirmed the proposed method is robust.

Conclusion

The present research work gives satisfactory result for method validation and development of stability indicating HPTLC method for quantitative determination of brivaracetam. The HPTLC method is simple, rapid, economic andmore flexible for routine analysis of brivaracetamin bulk and tablet dosage forms. Thus, the proposed method is of considerable importance and has sound industrial applicability for quality control and analysis of brivaracetam from bulk drug and formulations.

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Conflict of Interest

The authors have no conflict of interest to declare.

References

- Kenda, B. M., Matagne, A. C., Talaga, P. E., Pasau, P. M., Differding, E., Lallemand, B. I., & Michel, P. (2004). Discovery of 4-substituted pyrrolidonebutanamides as new agents with significant antiepileptic activity. *Journal of Medicinal Chemistry*, 47(3), 530–549. doi:10.1021/jm030913e.
- Lynch, B. A., Lambeng, N., Nocka, K., Kensel-Hammes, P., Bajjalieh, S. M., Matagne, A., & Fuks, B. (2004). The synaptic vesicle protein SV2A is binding site the antiepileptic drug levetiracetam. *Proceedings of the National Academy of Sciences of the United States of America*, 101(26), 9861–9866. doi:10.1073/pnas.0308208101.
- 3. Ryvlin, P., Werhahn, K., Blaszczyk, B., Johnson, M., & Lu, S. (2014). Adjunctive brivaracetam in adults with uncontrolled focal epilepsy: results from a double-blind, randomized, placebo-controlled trial. *Epilepsy Currents*, 55(1), 47–56. doi:10.1111/epi.12432.
- 4. Biton, V., Berkovic, S., Abou-Khalil, B., Sperling, M., Johnson, M. E., & Lu, S. (2014). Brivaracetam as adjuctive treatment for uncontrolled partial epilepsy in adults:a phase III randomized, double-blind, placebo-controlled trial. *Epilepsy Currents*, 55(1), 57–66. <u>doi:10.1111/epi.12433</u>.
- 5. Taylor, P., Pantaleone, D., Senkpeil, R., & Fotheringham, I. (1998). Novel biosynthetic approaches to the production of unnatural amino acids using transaminase. *Trends in Biotechnology*, 16(10), 412–418. doi:10.1016/S0167-7799(98)01240-2.
- Jacobs, P. L., Ridder, L., Ruijken, M., Rosing, H., Jager, N. G. L., Beijnen, J. H., ..., & van Dongen, W. D. (2013). Identification of drug metabolites in human plasma or serum integrating metabolite prediction, LC_HRMS and untargeted data processing. *Bioanalysis*, 5(17), 2115–2128. doi:10.4155/bio.13.178.

- 7. Maria, L., Sargentini, M., & Pascal, E. (2008). Pharmacokinetics and metabolism of ¹⁴c-brivaracetam, a novel SV2A ligand, in Healthy subject. *Drug Metabolism and Disposition: the Biological Fate of Chemicals*, 36(1), 36–45.
- 8. Nicolas, J., Chanteux, H., Rosa, M., Watanabe, S., & Stockis, A. (2012). Effect of gemfibrozil on the metabolism of brivaracetam in vitro and in human subject. *Drug Metabolism and Disposition: the Biological Fate of Chemicals*, 40(8), 1466–1472. doi:10.1124/dmd.112.045328.
- 9. Nicolas, J., Hannestad, J., Holden, D., Kervyn, K., Nabulsi, N., Tytgat, D., . . ., & Klitgaard, H. (2016). Brivaracetam a selective high affinity synaptic vesicle protein 2A (SV2A) ligand with preclinical evidence of high brain permeability and fast onset of action. *Epilepsia*, 57(2, S2), 201–209. doi:10.1111/epi.13267.
- 10. MaliN, Mhaske D. (2016). HPLC Studies on degradation behaviour of brivaracetam and development of validated stability-indicating HPLC assay method. *International Journal of Science and Research Methodology*, 4(3): 43-57.
- 11. Tian, X., Yuan, M., Zhou, Q., & Wang, X. (2015). The efficacy and safety of brivaracetam at different doses for partial-onset epilepsy: a meta-analysis of placebo-controlled studies. *Expert Opinion on Pharmacotherapy*, 16(12), 1755–1767. doi:10.1517/14656566.2015.1058360.
- 12. Vishweshwar V, Moses Babu J and Muralikrishna. (2018). Development and Validation of stabilityindicating UPLC method for the development of Brivaracetam, its related impurities and degradation products. *International Journal of Pharmaceutical Science and Research*, 9(6): 2315-2327.
- 13. Chauhan, K., & Choudhari, V. (2018). Development and Validation of stability indicating HPTLC method for estimating of Carbocisteine and Amoxicilin as bulk drug and in formulation by Derivatization. *International Journal of Pharm Tech Research*, 11(2), 108–118. doi:10.20902/IJPTR.2018.11202.
- 14. Sethi, P. D. HPTLC: Quantitative Analysis of Pharmaceutical formulation, 1st edition, New Delhi, CBS publishers and Distributors, 2013: 15-16.
- 15. Attimarad, M., Mueen Ahmed, K. K., Aldhubaib, B. E., & Harsha, S. (2011). High performance thin layer chromatography: A powerful analytical technique in pharmaceutical drug discovery. *Pharmaceutical Methods*, 2(2), 71–75. doi:10.4103/2229-4708.84436.
- 16. Upton, R. T. (2010). Use of high-performance thin layer chromatography by the American Herbal Pharmacopoeia. *Journal of AOAC International*, 93(5), 1349–1354.
- 17. ICH. Stability Testing of New Drug Substances and Products: International Conference on Harmonization, Q1A(R2), IFPMA, Geneva, Switzerland, 2003.
- 18. ICH Validation of analytical procedures; Text and methodology; Q2 (R1), International Conference onHarmonization, 2005.
