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The Effect of Ginger Plant (*Zingiber officinale*) Aqueous Extract on Function and Histological Structure of Kidney in Mice Treated with Carbon Tetrachloride

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Abstract : The percent work was designed to determine the effect of ginger plant aqueous extract on function and histological structure of kidney in mice treated with carbon tetrachloride (CCl_4). Ginger plant caused a protective effect against CCl_4 induced kidney damage and improved the kidney weight and biochemical parameters including urea, uric acid and creatinine. The ginger plant has a protective effect against injury in the kidney of mice treated with CCL_4 , because the ginger plant protects the tissues of kidney from toxic effect of CCL_4 . The kidney of CCL_4 treated mice showed many histological alterations in the kidney included: atrophy, vascular degeneration and hemorrhage, death cell, degeneration of epithelial cells, destruction of basement membrane and reduce of interstitial connective tissue. **Key words :** Kidney, CCl_4 , Urea.

1. Introduction:

The kidney is a vital organ present in vertebrates and some other animals. It has a wide range of functions such as the excretory organ and it filtered the blood to remove waste ^{1, 2}. The kidneys regulate the electrolytes balance including: salt, acid-base and fluid and they are also responsible for the reabsorption of amino acids, glucose and water ^{3, 4}. During the development, three kidney systems are formed in cranial to caudal sequence: the pronephros, (It is rudimentary and nonfunctional), mesonephros (It may function for a short time during early fetal period) and metanephros (It forms the permeant kidney) ^{5, 6}.

The kidneys were composed of renal tubules (nephrons), which developed from intermediate mesoderm (nephromeres)⁷. Mammalian kidneys were compact organ divided into a cortex and medulla ^{8,9}. The cortex and medulla contain the nephrons, which can be considered the functional units of the kidney; in human, each kidney composed of 1 to 4 million nephrons^{2, 3}.

The medical plants act as antiradicals and DNA cleavage protector's phytochemical are compounds found in plants that have beneficial effect on health or play an active role in amelioration of diseases. Ginger *(Zingiber officinale)* is a plant common ingredient in Asian and Indian cuisine. It is used for its medical properties for centuries among many cultures for over 5000 years^{10, 11}.

Ginger has been used for centuries to reduce inflammation and treat inflammatory conditions and antioxidant effect ¹². Toxic chemicals cause the kidney damages, which are induced by oxidative damages. Carbon tetrachloride (CCL₄), a well-known model compound for producing chemical injury ^{13, 14}.

 CCL_4 is regarded as highly toxic, it is known animal carcinogen and potential human carcinogen, CCL_4 is a clear, nonflammable, heavy liquid that evaporates readily, producing sweat odor. Its chemical stability results in an atmospheric half-life of 30 to 100 years about 1% of the total CCL_4 found in the environment is dissolved in surface waters and oceans. The CCL_4 evaporates easily, most of it released to the environment, it can remain in air for several years before it broken down and small amounts of it are found in surface water. CCL_4 can enter the body through lungs, brain, liver, skeletal muscles, kidney, stomach, intestine and skin and accumulates in the body fat. The aim of this study was to elevate the effect of ginger plant treatment on CCL_4 in mice using biochemical and histopathological parameters.

2. Materials and Methods

Twenty-four adult male albino mice (*Mus musculus*), weighting about (26-32 g) and aged (50-90 day), obtained from Al-Nahrain University, were housed in plastic cages at (20-24°C) temperature, 12 hour light/ 12 hour dark cycle. The animals feeding with laboratory diet and tap water *ad libitum*. The mice were divided randomly into experimental (18) and control (6) groups. The experimental group divided into three groups, were treated with aqueous ginger extract, CCL₄ and CCL₄ with aqueous ginger extract.

Ginger (*Zinger officiale*) plant was purchased from local market and botanical identification was confirmed at the herbarium of Baghdad University, preparation of aquatic extract of ginger, the root plant was dried and ground in an electric mill (10 gram of ginger by 100 ml distilled water).

 CCL_4 was purchased from sigma-Aldrich Co. St. USA.A single dose of CCL_4 (diluted in olive oil 1:1) at (0.2% ml/kg) body weight (BW). All groups were injection intraperitoneal (IP) (0.1 ml) for each dose, the control group received single dose of normal saline at one day for ten days, while three experimental groups, the first group received single dose of aqueous ginger extract (500 mg/kg) at day for ten days, the second group received single dose of CCL_4 (0.2%) for two days, the third group received single dose of CCL_4 (0.2%) for two days, the third group received single dose of CCL_4 (0.2%) for two days.

Mice of all groups were given ether anesthesia and the animals were scarified. The blood samples were collected by cardiac puncture, serum was collected for determine the urea, uric acid and creatinine^{15,16}.

The kidney was excised from animals and weighted and then fixed in 10% formalin for 24 hours ¹⁷ and washed by tap water. Paraffin wax methods employed in histological study according to the methods of Vacca (1985); Suvarn et al. (2013)^{18,19}. The collected data were analyzed by SPSS. The histological sections were observed by compound light microscope (Olympus, Japan).

3. Results

The current study is designed to demonstrate the effect of ginger plant (*Zinger officiale*) aqueous extract on kidney in albino mice (*Mus musculus*) treated with carbon tetrachloride (CCL_4), the results showed as following:

* Weight Study:

The CCL₄ treatment affected on the kidney tissues. It was found that significant decrease at (P<0.05) in kidney weight of CCL₄ treated mice 0.25 ± 0.003) gm compared with control group where the mean weight of kidney in treated group with CCL₄ was (0.14 ± 0.003) gmwile control group was (0.14 ± 0.01) gm(Table 1, Figure 1). However, a significant increase at (P<0.05) was observed in the kidney weight of mice treated with CCL₄ + ginger plant compared with CCL₄ treated group where the mean weight of kidney in treated group with CCL₄+ginger plant was (0.21 ± 0.01) gm while treated group with CCL₄ was (0.14 ± 0.003) gm are presented in (Table 1, Figure 1).

Table (1): The statistical analysis of the mean and stander error (Mean±S.E) of kidney weight in control and treated groups.

	Control group (Mean±S.E.)	Treated groups (Mean ±S.E.)		
		Ginger plant	CCL ₄	CCL ₄ + Ginger plant
Weight of kidney (gm)	0.25±0.01	0.23±0.002*	0.14±0.003*	0.21±0.01*

^{*}Significant differences (P<0.05)



Figure (1): The mean weight of kidney in control and treated groups.

Table (2): The statistical analysis of mean and stander error (Mean±S.E.) of level of urea, uric acid and creatinine in the serum blood of control and treated groups.

	Control group	Treated groups (Mean±S.E.)		
	(Mean±S.E.)	Ginger plant	CCL ₄	CCL ₄ +Ginger plant
Urea (µmol/L)	42.2±0.86	30.8±1.24*	54.8±2.6*	32.60±1.73*^
Uric acid (µmol/L)	2.15±0.04	2.02±0.04*	3.42±0.02*	2.25±0.02*^
Creatinine (µmol/L)	0.42±0.01	0.34±0.01*	0.45±0.01*	0.36±0.02*^

*^ Significant differences (P<0.05)



Figure (2): The concentration of urea in the serum blood of control and treated mice.



Figure (3): The concentration of uric acid in the serum blood of control and treated mice.





* Biochemical Study

In current study, the levels of urea, uric acid and creatinine were estimated in serum blood as the kidney function biomarkers. Urea and uric acid levels showed significant increase at (P<0.05) in the treated group with CCL₄compared with control group, where the mean levels of urea and uric acid in treated groups with CCL₄ were(54.8±2.6, $3.42\pm0.02 \mu mol/L$) while control group was (42.2 ± 0.89 , $2.15\pm0.04 \mu mol/L$) (Table 2, Figure 2, 3).Non-significant differences were noticed (P<0.05) in creatinine level of treated group with CCL₄compared with control group are presented in (Table 2, Figure 4).

Significant decrease at (P<0.05) was noted in urea, uric acid and creatinine levels in the CCL₄ +ginger plant treated mice compared with CCL₄ treated group, where the mean levels of urea, uric acid and creatinine in the treated group CCL₄ +ginger plant were (32.60 ± 1.73 , 2.35 ± 0.02 , 0.36 ± 0.02 µmol/L) while CCL₄ treated group were (54.8 ± 2.6 , 3.42 ± 0.02 , 0.45 ± 0.01 µmol/L) (Figure 2, 3, 4).

*Histological Study

The microscopic observation of kidney sections of control group that the kidneys were located, one on either side of the vertebral column, the outer surface of the kidney was a layer of connective tissue called the renal capsule, this capsule covers the kidney.

The kidney composed of two regions: the outer layer, was cortex and the inner layer, was medulla (Figure 5). The cortex and medulla contain functional units (nephrons), which are represented by renal corpuscles (Bowman's capsules). The initial portions of a nephrons were the renal corpuscles (Bowman's capsules), which were located in the cortex of kidney. Bowman's capsule consists of double wall, the outer wall was parietal layer (capsular epithelium) and inner wall was visceral layer (glomerular layer). The epithelial tissue of Bowman's capsule was composed of simple squamous.

The thin cavity between parietal layer and visceral layer was the capsular space (Bowman's capsule) (Figure 6). The afferent arterioles form the glomerulus, which are formed a network of capillaries enclosed in Bowman's capsules (Figure 6).

The followed by renal corpuscle (Bowman's capsule) was renal tubule, that passes from the cortex to the deep part of medulla and called proximal convoluted tubule. The epithelial tissue lining of this tubule was composed of simple cuboidal epithelial tissue and its bearing a brush border in the free surface. The cytoplasm of the cells was light colored and the nuclei of cells were dark colored in central location (Figure 6).



Figure (5): Cross section through mice kidney of control group showing: Capsule (CA), Cortex (C), Medulla (M), (H&E, 4x).



Figure (6): Cross section through mice cortex kidney of control group showing Bowman's capsule (BC), Blood vessel glomeruli (BVG), Capsule space (CS), proximal convoluted tubule (PCT), distal convoluted tubule (DCT). (H&E, 40x).

The distal convoluted tubule was formed from the thick segment of the ascending limb. The lining epithelial tissue of distal tubules was simple cuboidal. The cells contain nuclei in the central and dark-colored.

The lumen of distal convoluted tubule was larger than in proximal convoluted tubule. There was no brush border. The cells were more numerous than in proximal convoluted tubule (Figure 6).

The distal tubules were continuous with collecting tubules (connecting tubules). The cells of collecting tubules were simple cuboidal epithelial tissue and have nuclei and clear cytoplasm (Figure 7). The kidney of ginger plant only treated mice did not reveal any histopathological alternations (infiltration, congestion, enlargement of epithelial cells, atrophy and necrosis) (Figure 8).



Figure (7): Cross section through mice medulla kidney of control group showing collecting tubules (CT), interstitial connective tissue (ICT), (H&E, 40x).



Figure (8): Cross section through mice kidney of treated group with ginger plant showing normal cortex (C) and medulla (M) region (H&E 4x).



Figure (9): Cross section through mice kidney of CCL_4 treated group showing atrophy of glomerulus (AT) (H&E, 40x).



Figure (10): Cross section through mice kidney of CCL₄ treated group showing cell death (CD) of convoluted tubules (H&E, 40x).

The kidney of CCl₄ treated mice showed many histological alternations included atrophy of glomerulus and vascular degeneration and decrease in the diameter of glomerulus (Figure 9). Degeneration of epithelial cells and cell death of renal tubules (Figure 10).Thickening of epithelial cells layer of renal tubules (Figure

11).Destruction of basement membrane in convoluted tubule (Figure 11).Partial detachment of epithelial cells in convoluted tubule (Figure 11).Increase interstitial space with reduce interstitial connective tissue of kidney (Figure 11).



Figure (11) : Cross section through mice kidney of CCL_4 treated group showing cell layer thickening (T) of convoluted tubules, destruction of basement membrane (D), detachment (DE) of epithelial cells in convoluted tubule and increase interstitial space (IS)(H&E, 40x).



Figure (12): Cross section through mice kidney of CCL_4 +ginger planttreated group showing cortex (C) and Medulla (M) (AT) (H&E, 4x).

Congested blood vessels and some of blood vessels were dilation in the kidney medulla of the treated group with CCL_4 and hemorrhage (Figure 12). The vascular degeneration in the treated group with CCL_4 compared with control, treated with ginger plant and CCL_4 +ginger plant (Figure 12). The kidney of CCL_4 +ginger plant treated mice showed clear recovery characterized, and the kidney sections showed intact glomerulesand normal convoluted tubules structure (Figure 12) when compared with control group (Figure 5, 6, 7).

Discussion

In the present study, we investigated the effect of ginger aqueous extract ginger plant was reported to be present the increase in glomeruli damage. The preventive effect of ginger plant induced oxidative stress in the mice is due to its antioxidant properties. The ginger plant contains high concentrations of vitamins A, B, C and E and is very rich in magnesium and other trace elements, these vitamins act as antioxidants and preventing toxicant induced tissue injury ¹². CCL₄ is used as solvent for oils and fats, and widely used as a cleaning fluid (dry cleaning) and used in fire extinguishers and used as pesticide kill insects in grain, but this was stopped in 1986. CCL₄ is poisoning by the skin absorption, ingestion and inhalation. Inhalation of CCL₄ vaporous can depress central nervous system activity and cause degeneration of the kidneys ^{13, 14}.

After analyzing different sections from control and treated groups animals by using compound light microscope, the result declared difference in the mice kidney associated with administration of CCL_4 in a dose (0.2% ml/kg) on albino mice (*Mus musculus*).

In the kidney of mice treated with CCL_4 induced severe changes, glomeruli look small, atrophied and loosely arranged in Bowman's capsule ²⁰.

In the present work showed detachment of epithelial cells from basement membrane of convoluted tubules in mice kidney treated with CCL_4 , this result agrees with Althnaian *et al.* (2013)²¹ who pointed detachment of cells associated with impairment in meiosis ^{21,22}.

In this study, it was found that destruction of basement membrane of the convoluted tubules this result agrees with Cordeiro and Kaliwal $(2013)^{20}$ who showed the similar change in basement membrane of convoluted tubules (Cordeiro and Kaliwal, $2013)^{20}$.

Also, degeneration of proximal and distal tubules and interstitial connective tissue in mice kidney treated with CCL4, this result agree with Brattin et al. $(1985)^{13}$, who documented that the CCL₄ cause histopathological effect, represented by degeneration and destruction of interstitial connective tissue other studies suggest the cause of degeneration due to CCL₄ accumulation in lysosomes then rapture the cells and death or due to inhibition of protein synthesis ²¹.

Also, cell death in mice kidney treated with CCL_4 , other studies documented that the cell death due to the DNA repair synthesis was detected, the CCL_4 is bio transformed to trichloromethyl free radicals (CCL_3) is considered the initial step in a chain of events, that lead to bind to lipids, contributing to the breakdown of cell structure and protein synthesis ad disrupting cell energy process lead to cell apoptosis (necrosis)^{22, 23}.

The histological sections showed congested blood vessels, which caused by inflammation that causes changes in blood flow and gets relaxation and dilation in the blood vessels that lead to blood accumulates in dilated capillaries and venules^{24, 25}.

The present study showed hemorrhage in mice kidney treated with CCL_4 this result agree with Ogeturk*et al.* $(2005)^{26}$ who revealed hemorrhage occur in the rat kidney.

Ginger root inhibit the induction of genes encoding cytokines and chemokines that synthesized and secreted at sites of inflammation, ginger reported to inhibit amyloid AB peptide induced cytokine and chemokine expression in cultured THP-1 monocytes ²⁷. The ginger inhibits platelet aggregation ²⁸.

The kidney has a higher affinity to CCL_4 than liver due to the predominance of cytochrome p-450 in the renal cortex ^{20, 29}. The kidney is sensitive to CCL_4 leading to build up of water in the body and waste products in the blood causing kidney failure ²⁹. The biochemical finding was confirmed by histological observation. The

changes include cellular necrosis, blood vessel damage and infiltration of inflammatory cells, atrophy and other histological changes which were consistent with the other authors ^{13, 20, 21}.

Consistent with previous studies, the administration of CCL_4 induced a renal failure indicated by elevation of serum urea, creatinine and uric acid ^{30,31}. In the present study serum blood biomarkers, urea and uric acid levels were increased (P<0.05) and no significant change in serum creatinine in mice group with CCL_4 compare to mice control group.

This result agree with other studies Khan *et al.* $(2009)^{29}$ and Mbarki *et al.* $(2016)^{33}$, and it disagree with Ogeturk *et al.* $(2005)^{26}$ in the level of serum urea, while this result agree with Althnaian *et al.* $(2012)^{32}$, and it disagree with Mbarki *et al.* $(2016)^{33}$ in the level of serum uric acid, the level of creatinine in this study agree with Khan and Al-Zohairy (2011) and Althnaian *et al.* $(2012)^{32, 34}$, while it disagree with Khan *et al.* $(2009)^{29}$ and Mbraki *et al.* $(2016)^{33}$, the increased serum blood level of renal markers have been attributed to the kidney injury ²⁰.

The present study declared a significant decrease in weight of kidney mice treated with CCL_4 compared with control group, this decrease may be due to decrease mass of tissues lead to decrease weight of kidney.

References:

- 1. Kardong, K. V. (1998). Vertebrates: comparative anatomy, function, evolution. 4th ed., McGraw Hill, New York.
- 2. Young, B.; Low, J.; Steven, A. and Heath, J. W. (2006). Functional histology. Churchill Langston, London.
- 3. Kent, G. C. and Carr, R. K. (2001). Comparative anatomy of vertebrates. 9th ed., McGraw Hill, New York.
- 4. Yari, A. and Gharzi, A. (2013). Anatomical and histological study of the excretory system in the Bosc's Fringe-Toed Lizard (*Acanthodactyiusboskianus*). Asian J. Animal Sci., 7: 30-35.
- 5. Sadler, T. W. (2006). Medical embryology. 10th ed. Lippincott Williams and Wilkins, New York: 371 pp.
- 6. Keith, L. M.; Persaud, T. V. and Mark, G. T. (2016). The developing human: clinically oriented embryology. 10th ed. Elsevier, Philadelphia: 260-262 p.
- Sadler, T. W. (2015). Langman's medical embryology. 13th ed. Wolters Kluwer Healt, New York: 261-264 p.
- 8. Leake, L. D. (1975). Comparative histology. Academic Press Inc., New York: 738 pp.
- 9. Ross, M, H.; Romerell, L. J. and Kay, G. I. (1995). Histology: a text and atlas. 3rd ed. Williams and Wilkins, USA: 678-638 p.
- 10. Cobley, L. S. (1976). An introduction the botany of tropical crops. Logman, London.
- 11. Purseglove, J. S.; Brown, E. G.; Green, C. L. and Robbins, S. R. (1981). Species, Vol. 1 Longman Group Limited, London.
- 12. Terry, R.; Rosadzki, P.; Watson, L. K. and Emst, E. (2011). The use of ginger (*Zingiber officinale*) for the treatment of pain: a systemic review of clinical trials. Pain Med., 12(12): 1808-1818.
- 13. Brattin, W. J.; Glende Jr. E. A. and Recknagel, R. O. (1985). Pathological mechanisms in carbon tetrachloride hepatotoxicity. J. Free Radic. Biol. Med., 1: 27-38.
- 14. Al-Assaf, A. H. (2013). Hepatoprotective and antioxidant effect of corsolic acid on carbon tetrachloride induced hepatoxicity. Acad. J., 7(12): 673-678.
- 15. Buege, I. A. and Aust, S. D. (1978). Microsomal lipid peroxidation. Methods Enzymol., 52: 302-310.
- 16. Tabacco, A.; Meiatim, F.; Moda, E. and Tarli, P. (1979). Simplified enzymic colorimetric serum urea nitrogen determination. Clin. Chem., 25(2): 336-337.
- 17. Luna, L. G. (1968). Manual of histological staining methods. 3rd ed., McGraw-Hill Book Co. Inc. New York: 258 pp.
- 18. Vacca, L. (1985). Laboratory manual of histochemistry. Raven Press, New York: 238 pp.
- 19. Suvarn, K.; Layton, C. and Bancroft, J. (2013). Bancroft's theory and practice of histological techniques 7th ed. Churchill Livingston, New York: 654 pp.
- 20. Cordeiro, M. C. and Kaliwal, B. B. (2013). Protective role of bark extract of *Bridelia retusa* Spreng on CCL₄ induced histological toxicity in mice. J. Pharm. Phyto., 2(4): 142-148.

- 21. Althnaian, T.; Albokhadaim, I. and El-Bahr, S. M. (2013). Biochemical and histopathological study in rats intoxicated with CCL₄ and treated with camel milk. Springer Plus, 2(1): 57.
- 22. Williams, A. T. and Burk, R. F. (1990). Carbon tetrachloride hepatotoxicity: an example of free radical mediated injury. Semir. Liver Dis., 10:279-284.
- 23. Basu, S. (2003). Carbon tetrachloride-induced lipid peroxidation eicosanoid formation and their regulation by antioxidant nutrient. Toxicology, 189: 113-127.
- 24. Junqueria, L. C.; Carnerio, J. and Kelly, R. O. (1998). Basic histology. 8th ed. McGraw-Hill Com., New York: 494 pp.
- 25. Harrison, d. G.; Widder, J.; Grumbach, I. (2006). Endothelial mechanotransduction nitric oxide and vascular inflammation. J. Intern. Med., 259: 351-363.
- 26. Ogeturk, M.; Kus, I.; Kavakli, A.; Oner, J.; Kukner, A. and Sarsilmaz, A. (2005). Reduction of carbon tetrachloride induction nephropathy by melatonin administration. Cell Biochem. Funct., 23(2): 85-92.
- Grzanna et al. (2004). Ginger Extract Inhibits β-Amyloid Peptide–Induced Cytokine and Chemokine Expression in Cultured THP-1 Monocytes. The Journal of Alternative and Complementary Medicine. 10(6): 1009-1013
- 28. Liao, Y. R.; Leu, Y. L.; Chan, Y. Y.; Kuo, P. C. and Wu, T. S. (2012). Anti-platelet aggregation and vasorelaxing effects of the constituents of the rhizomes of *Zingiber officinale*. Molecules, 17(8): 9928-8937.
- 29. Khan, M. R.; Rizvi, W. and Shaheen, S. (2009). Carbon tetrachloride induced nephrotoxicity in rats: protective role of *Digera muricata*. J. Ethnopha., 122(1): 91-99.
- 30. Abdel Moneim, A. E. and El-Deib, K. M. (2012). The possible protective effects of *Physalis peruviana* on CCL₄-induced nephrontoxify in male albino rats. Life Sci. J., 2012(9): 1038-1052.
- 31. Al-Yahya, M.; Mothana, R.; Al-Said, M.; Al-Dosari, M. (2013). Attenuation CCL₄-induced oxidative stress and hepatonephrotoxity by Saudi Sidr honey in rats. Evidence-Based Complement. Altern. Med., 115-1165.
- 32. Althnaian, T.; Albokhadaim, I. and El-Bahr, S. M. (2012). Biochemical and histopathological study in rats intoxicated with carbon tetrachloride and treated with camel milk. Springer Plus., 2(57).
- 33. Mbarki, S.; Dhibi, S.; Bouaenna, H.; Elfeki, A. and Hfaiedh, N. (2016). Effect of MgCl₂ supplementation on blood parameters and kidney injury of rats exposed to CCL₄. Pen Life Sci., 11: 250-258.
- 34. Khan, A. A. and Al-Zohairy, M. (2011). Hepatoprotective effects of camel milk against CCL₄ induced hepatotoxicity in rats Asian J. Biochem., 6(2): 171-180.
