

In vitro antidermatophytic effects of the methanolic extract of the *Amygdalus eburnea*

Mehdi Rezaeifar¹, Maryam Rezaeifar^{2*}

¹Department of Pharmaceutics, Faculty of Pharmacy, Kerman University of Medical Sciences, Kerman, Iran

²School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

Abstract : Dermatophytes are one of the main significant groups of fungi, which lead to globally human and animal's infections. The present study aims to determine the antidermatophyte activity of *A. eburnea* methanolic and aqueous extracts on *in vitro*. Minimum inhibitory concentration (MIC) of extracts against tested dermatophytes (*Trichophyton mentagrophytes* and *T. interdigitale*) was determined by broth macrodilution method, according to the protocol M38-A2 of the Clinical and Laboratory Standards Institute (CLSI) for filamentous fungi with some modifications. The results demonstrated that all the extracts had fungistatic activities with the MIC values 3.3 to 6.6 mg/ml. Methanolic extract of *A. eburnea* was much more effective than aqueous extract of plant once they showed lower MIC for tested dermatophyte strains. To conclude, the obtained findings demonstrated that *A. eburnea* extracts were found to be more active against some dermatophytic fungi strains and thus provided the evidence for its traditional use value and it is suitable substitute for treatment of fungi infections.

Keywords: Dermatophyte; MIC; Extract: *Trichophyton*.

Introduction

Dermatophytes are one of the main significant groups of fungi, which lead to globally human and animal's infections^{1, 2}. Dermatophytes include the capacity to invade keratinized tissues, such as hair, skin and nails to produce dermatophytosis^{3, 4}. Various types of the disease including tinea corporis, tinea pedis, capitis, barbae, cruris, manum and onychomycosis are present. Nowadays, treatments for these infections are still restricted to a few antifungal drugs. However, these drugs have limited in use due to having high toxicity and the emergence of drug resistance in their antifungal activities⁵⁻⁷.

Historicalys, plant extracts and their compounds, due to having fewer side effects, low cost and high availability, are valuable sources that are commonly used to treat a wide range of disease conditions including infectious diseases⁸⁻¹⁰.

One of these interesting plants is *Amygdalus eburnea* Spach. (called "Ghosk" in Persian) from family of Rosaceae as a type of almond which is naturally grown and distributed in Iran¹¹. In folk Iranian medicine *A. eburnea* has been used as laxative and anti-worm. Moreover, brew of dermal tissue are used for cough, respiratory distress and paregoric¹¹. The present study aims to determine the antidermatophyte activity of *A. eburnea* methanolic and aqueous extracts on *in vitro*.

Experimental

Collection of plant materials

The shell root of *A. eburnean* was collected from rural regions of from Baft district, south east of Iran, in April 2013. They were identified by a botanist of the Botany Department of Shahid Bahonar University, Kerman, Iran¹². A voucher specimen of the plant materials was deposited at the Herbarium of Department of Pharmacognosy of School of Pharmacy, Kerman University of Medical Science, Iran.

Preparing of extracts

One hundred gram of powdered plant material was separately extracted by percolation method with methanol (80%) and water successively for 72 h. in room temperature. The extracts were passed through filter paper (Whatman No.3, Sigma, Germany) to remove plant debris. The extracts were finally concentrated in vacuum at 50°C using a rotary evaporator (Heidolph, Germany) and stored at -20°C, until testing¹³⁻¹⁵.

Minimum inhibitory concentration (MIC) determination

The fungal strains used in this work are *Trichophyton mentagrophytes* (ATCC 9533), and *T. interdigitale* (ATCC 200099). The MIC of extracts against tested dermatophytes was determined by broth macrodilution method, according to the protocol M38-A2 of the Clinical and Laboratory Standards Institute (CLSI) for filamentous fungi with some modifications¹². For the broth macrodilution method, 0.9 ml of the final conidia suspensions were mixed with 0.1 ml⁻¹ of the different concentrations of extract (0.0625-16 mg/ml) in test tubes and incubated at 30°C for 7 days. The positive control tube contained 0.9 ml of conidial suspension and 0.1 ml of RPMI-1640, and the negative one contained 1 ml of RPMI-1640 only. The minimum concentrations at which no visible growth was observed were defined as the MIC, which were expressed in mg/ml¹⁶⁻¹⁸.

Statistical analysis

Data analysis was done using SPSS statistical package, version 16.0 (SPSS Inc., Chicago, IL, USA). To assess the interaction of time and the experimental group, repeated measures analysis test was used. Differences were significant when the *p*-value was lower than 0.05^{19, 20}.

Results and Discussion

Table 1 shows the results of *in vitro* antifungal activity of *A. eburnean* extracts against *T. mentagrophytes* and *T. interdigitale*. The results demonstrated that all the extracts had fungistatic activities with the MIC values 3.3 to 6.6 mg/ml. Methanolic extract of *A. eburnean* was much more effective than aqueous extract of plant once they showed lower MIC for tested dermatophyte strains. Among the tested dermatophytes, *T. interdigitale* was the most sensitive to the extracts of *A. eburnean*, while *mentagrophytes* was the less effective. Moreover, fluconazole as control drug revealed both fungistatic and fungicidal activities with the MIC value of 0.0184 to 0.0368 mg/ml against tested dermatophytes. Based on the statistical analysis, the difference in antidermatophytic effects between the extracts and the control drug was not statistically significant (*p*>0.05).

Table 1. Mean of minimum inhibitory concentration (MIC) of *A. eburnean* extracts against some pathogenic dermatophyte species.

Tested samples	MIC (mg/ml)	
	<i>Trichophyton interdigitale</i>	<i>Trichophyton mentagrophytes</i>
Methanolic extract	3.3	5.3
Aqueous extract	5.3	6.6
Fluconazole	0.0184	0.0364

Regarding antimicrobial effects of *A. eburnean*, previous study have reported that the extract from the *A. eburnean* were found to be more active against Gram-positive bacteria as an available and inexpensive herb possesses wound healing activity, and thus provided the evidence for its traditional use value and it is suitable substitute in healing of burn wounds²¹. Recently, Rezaeifar et al have demonstrated that *A. eburnean* extracts were found to be more active against some pathogenic fungi strains (*Aspergillus flavus* and *Candida albicans*) and thus provided the evidence for its traditional use value and it is suitable substitute for treatment of fungi infections²².

According to the previous studies, in phytochemical screening of the crude extract of all of plants there are some compounds such as terpenoids, phenols, flavonoids, fatty acids and sterols²³⁻²⁷. Reviews have reported biological and antimicrobial activities of these components²⁸⁻³³. Thus, we can suggest that these components are responsible for the antifungal activity of *A. eburnean*; however their exact action mechanism is poorly understood. To conclude, the obtained findings demonstrated that *A. eburnean* extracts were found to be more active against some dermatophytic fungi strains and thus provided the evidence for its traditional use value and it is suitable substitute for treatment of fungi infections.

Declaration of Interest

The author declares that there is no conflict of interest in this study.

References

1. Weitzman I, Summerbell RC. The dermatophytes. Clin Microb Rev 2001; 8: 240-259.
2. Watanabe S. Present state and future direction of topical antifungals. Japanese J Med Mycol 1990; 40: 151-155.
3. Ayatollahi Mousavi SA, Kazemi A. In vitro and in vivo antidermatophytic activities of some Iranian medicinal plants. Med Mycol. 2015; 53(8):852-9.
4. Mahmoudvand H, Sepahvand A, Jahanbakhsh S, Ezatpour B, Ayatollahi Mousavi SA. Evaluation of antifungal activities of the essential oil and various extracts of *Nigella sativa* and its main component, thymoquinone against pathogenic dermatophyte strains. J Mycol Med. 2014; 24(4):e155-61.
5. Muhaimin, Syamsurizal, Anis Yohana Chaerunisaa, Meity Suradji Sinaga. Eusiderin I from *Eusideroxylon zwageri* as Antifungal agent against Plant Pathogenic Fungus. International Journal of ChemTech 2016; 9(5):418-424.
6. Ravindra S. Dhivare, S. S. Rajput. Microwave assisted solvent free synthesis and antifungal evaluation of 3, 5-bis-(4-hydroxy-3-methoxybenzylidene)-N-phenylpiperidine-2, 6-dione derived from N-phenyl glutarimides. International Journal of ChemTech Research 2016; 9(3): 325-331.
7. Meena Deviha S., Pavithra M.K.S. Antifungal activity by ethanolic extracts of medicinal plants against *Malassezia furfur*: A potential application in the treatment of Dandruff. International Journal of PharmTech Research 2015; 8(3):440-443.
8. Ezatpour B, Saedi Dezaki E, Mahmoudvand H, Azadpour M, Ezzatkhan F. In vitro and in vivo antileishmanial effects of *Pistacia khinjuk* against *Leishmania tropica* and *Leishmania major*. Evid Based Complement Alternat Med. 2015, 149707.
9. Mahmoudvand H, Saedi Dezaki E, Ezatpour B, Sharifi I, Kheirandish F, Rashidipour M. In vitro and In vivo antileishmanial activities of *Pistacia vera* essential Oil. Planta Medica 2016, 82(4), 279-284.
10. Tavakoli Kareshk A, Keyhani A, Mahmoudvan d H, Tavakoli Oliaei R, Asadi A, Andishmand M, et al. Efficacy of the *Bunium Persicum* (Boiss) essential oil against acute toxoplasmosis in mice model. Iran J Parasitol. 2015, 10 (4), 625-631.
11. Madani B, Rahemi M, Baninasab B, Mahmoodi M. Morphological Evaluation of Three Native Species of *Amygdalus* in Iran." Horticulture Environment & Biotechnol 2009, 50(1), 63-67.
12. Mahmoudvand H, Sharififar F, Saedi Dezaki E, Ezatpour B, Jahanbakhsh S, Fasihi Harandi M. Protoscolicidal effect of *Berberis vulgaris* root extract and its main compound, berberine in cystic echinococcosis. Iranian J Parasitol. 2014, 9(4), 26-34.
13. Mahmoudvand H, Nadri S, Jahanbakhsh S, Rezaeifar M. Evaluations of protoscolicidal activity of *Cardamom* extract against hydatid cyst protoscoleces. Der Pharma Chemica 2016; 8(6): 91-95.
14. Haidari M, Azargoon A, Mahmoudvand H, Almasi V, Pournia S, Shams Khorramabadi M. Complications of primary realignment of posterior urethral disruption after pelvic trauma. Trauma Mon. 2014; 19 (2): e13523.

15. Nuzhat Tabassum and Vidyasagar G. M. Synthesis, Characterization and Antimicrobial activity of Silver nanoparticles using Santalum album aqueous seeds extract. *International Journal of ChemTech Research* 2016; 9(5): 352-358.
16. CLSI/Clinical and Laboratory Standards Institute, 2002. Reference method for broth dilution antifungal susceptibility testing of conidium-forming filamentous fungi. Wayne. (Approved Standard M38-A).
17. Mahmoudvand H, Kheirandish F, Saedi Dezaki E, Shamsaddini S and Fasihi Harandi M. Chemical composition, efficacy and safety of *Pistacia vera* (var. Fandoghi) to inactivate protoscoleces during hydatid cyst surgery. *Biomed Pharmacother* 2016; 82: 393–398.
18. H Mahmoudvand; SR Mirbadie; S Sadooghian; M Fasihi Harandi; S Jahanbakhsh; E Saedi Dezaki. Chemical composition and scolicalidal activity of *Zataria multiflora* Boiss essential oil. *J Essential Oil Res.* 2016; DOI: 10.1080/10412905.2016.1201546
19. Kheirandish F, Delfan B, Mahmoudvand H , Moradi M , Ezatpoura B, Ebrahimzadehc F, Rashidipour M. Antileishmanial, antioxidant, and cytotoxic activities of *Quercus infectoria* Olivier extract. *Biomedicine & Pharmacotherapy* 2016; 82: 208–215.
20. Jahanbakhsh S, Azadpour M, Tavakoli Kareshk A, Keyhani A. Mahmoudvand H. *Zataria multiflora* Bioss: lethal effects of methanolic extract against protoscoleces of *Echinococcus granulosus*. *J Parasit Dis* 2015, DOI 10.1007/s12639-015-0670-4.
21. Rezaeifar R, Ayatollahi Mousavi SA, Mehrabani M, Sepahvand A. Evaluation of the antifungal effects of various extracts of *Amygdalus eburnean* on some fungal pathogens. *Der Pharma Chemica* 2016, 8(6)l, 140-142.
22. Rezaeifar M, Behfarnezhad M, Moradi M, Mehrabani M, Mahmoudvand H. Antibacterial effects of various extracts of *Amygdalus eburnea* on some most common bacteria in burning. *Der Pharma Lettre.* 2016;8(6): 110-112.
23. Mahmoudvand H, Asadi A, Harandi MF, Sharififar F, Jahanbakhsh S, Dezaki ES. In vitro lethal effects of various extracts of *Nigella sativa* seed on hydatid cyst protoscoleces. *Iran J Basic Med Sci.* 2014, 17(12), 1001-6.
24. Rezaeifar M, Mahmoudvand H, Amiria M. Formulation and evaluation of diphenhydramine gel using different gelling agents. *Der Pharma Chemica*, 2016, 8 (5), 243-249.
25. Mahmoudvand H, Sharififar F, Rahmat MS, Tavakoli R, Dezaki ES, Jahanbakhsh S, Sharifi I. Evaluation of antileishmanial activity and cytotoxicity of the extracts of *Berberis vulgaris* and *Nigella sativa* against *Leishmania tropica*. *J Vector Borne Dis.* 2014, 51(4), 294-9.
26. Teimori H, Sabzi, F, Hassani V, Nadri S, Mahmoudvand H. Base deficit in immediate postoperative period of open heart surgery and outcome of patients. *Acta Medica Iranica* 2007; 47(3): 227-232.
27. Mahmoudvand H, Saedi Dezaki E, Kheirandish F, Ezatpour B, Jahanbakhsh S, Fasihi Harandi M. Scolicidal effects of black cumin seed (*Nigella sativa*) essential oil on hydatid cysts. *Korean J Parasitol.* 2014, 52, 653-659.
28. Mahmoudvand H, Shakibaie M, Tavakoli R, Jahanbakhsh S, Sharifi, I. In vitro study of leishmanicidal activity of biogenic selenium nanoparticles against Iranian isolate of sensitive and glucantime-resistant *Leishmania tropica*. *Iran J Parasitol* 2014, 9(4), 452-460.
29. Ahmadinejad M, Aliepour A, Anbari K, Kaviani M, Ganjizadeh H, Nadri S, et al. Fine-Needle Aspiration, Touch Imprint, and Crush Preparation Cytology for Diagnosing Thyroid Malignancies in Thyroid Nodules. *Indian J Surg* 2013, 1-4.
30. Mahmoudvand H, Ziaali N, Ghazvini H, Shojaee S, Keshavarz H, Esmaeelpour K, Sheibani V. *Toxoplasma gondii* infection promotes neuroinflammation through cytokine networks and induced hyperalgesia in BALB/c mice. *Inflammation.* 2016, 39(1), 405-412.
31. Mahmoudvand H, Ziaali N, Aghaei I, Sheibani V, Shojaee S, Keshavarz H, Shabani M. The possible association between *Toxoplasma gondii* infection and risk of anxiety and cognitive disorders in BALB/c mice. *Pathogen Glob Health.* 109(8):369-76.
32. Cristani M, D'Arrigo M, Mandalari G, Castelli F, Sarpietro MG, Micieli D, Venuti V, Bisignano G, Saija A, Trombetta D. Interaction of four monoterpenes contained in essential oils with model membranes: implications for their antibacterial activity. *J. Agric. Food. Chem.* 2007, 55(15), 6300-8.
33. Mahmoudvand H, Nadri S, Rezaeifar M, Rezaeifar M. Scolicidal effects of Myrtle methanolic extract on hydatid cyst protoscoleces. *Der Pharmacia Lettre*, 2016, 7 (8): 27-30.

Extra Page not to be Printed out.

For your Research work, for citations/References Log on to=

www.sphinxesai.com

International Journal of ChemTech Research

International Journal of PharmTech Research

Sai Scientific Communications
