



Comparison Of Biological Characteristic And Osteogenic Differentiation Between Bone Marrow And Adipose Mesenchymal Stem Cell In Various Age Group

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Abstract : Introduction. Mesenchymal stem cell is the answer of complicated medicine problems, including orthopaedic. Bone marrow is still the main source. Because of limited source, invasive procedure, pain, and relative less cell, adipose will be a promising source with equal regenerating and differentiating ability. Along with increasing life expectancy, geriatric population is increasing as well as the potential need for stem cell application. Yet there is still controversy about stem cell quality in aging.

Methods. This study was conducted in Stem Cell Medical Technology Integrated Service Unit Cipto Mangunkusumo Hospital, Faculty of Medicine Universitas Indonesia, Jakarta, October 2015 - March 2016. 12 patients were divided into 3 age group; 15-30 year, 31-40 year, and 41-55 year. Bone marrow from posterior iliac crest and adipose tissue were collected, mesenchymal stem cell isolation and culture were done subsequently. Biological characterization, Population Doubling Time, osteogenic differentiation, and alizarin red assay were carried out. All data was analyzed using SPSS 20.

Results. No significant difference was observed in biological characteristic and osteogenic differentiation of bone marrow and adipose mesenchymal stem cell among age group ($p > 0.05$). There is significant difference in Population Doubling time in 31-40 year group ($p = 0.000$) and 41-55 year group ($p = 0.000$).

Conclusions. Adipose mesenchymal stem cell had steady biological characteristic, Population Doubling Time, and osteogenic differentiation. Bone marrow mesenchymal stem cell had increasing population doubling time in increasing age, apart from biological characteristic and osteogenic differentiation. Adipose could be the source of choice in harvesting mesenchymal stem cell at any age.

Keywords: Mesenchymal Stem Cell, Osteogenic Differentiation, Bone Marrow, Adipose, Age.

Introduction

Mesenchymal stem cell has self regenerating, rapid proliferating, and multipotent capability.^{1,2} Bone marrow was the main source. Limited source, relative less cell, and invasive procedure aggravated exploration of other source.^{3,4} With equal regenerating and differentiating capability, adipose tissue recently rised up as another source. Furthermore, it had abundant resources, more amount of cells, and relative less invasive harvesting procedure.^{4,6}

Commonly, research was stressed on applying stem cell in curing disease but currently there was no direct study comparing adipose and bone marrow derived mesenchymal stem cells from one individual source.⁷ Increasing human life expectancy creates elderly people who prefer their own autogenous stem cell, and automatically raise the question about aging in mesenchymal stem cell.^{3,8,9} Some studies had been carried out and gave various results. Indeed, there is need to do comparative study of characterization and osteogenic differentiation of bone marrow and adipose mesenchymal stem cells in various age groups.

Experimental

This observational analytical *in vitro* study was conducted in Stem Cell Medical Technology Integrated Service Unit Cipto Mangunkusumo Hospital, Faculty of Medicine Universitas Indonesia, Jakarta, October 2015-March 2016. 6 patients were divided into 3 age groups; group 1 (15-30 year), group 2 (31-40 year), and group 3 (41-55 year). This study was approved by the Ethics Committee of the Faculty of Medicine, Universitas Indonesia. Bone marrow aspiration from posterior iliac crest and adipose tissue harvest (hip, elbow, and back) were carried out in every subject. Mesenchymal stem cell isolation and culture were done afterwards.

Bone marrow was cultured in medium of 100U Penicilin/100 µg Streptomycin/ml, 0.25 µg Fungizone/ml, 10U heparin, 1% Glutamax, 10% PRP, and alpha minimum essential medium (α -MEM). The samples were then centrifugated and cultured on media with temperature 37°C, 5% CO₂ and 20% O₂ until there was confluence. Meanwhile, adipose tissues were cut into small pieces, type I collagenase was added. After centrifugation, the samples were cultured in medium of 1% Amphotericin B and 10% PBS with temperature 37°C, 5% CO₂ and 20% O₂ until there was confluence. After reaching confluence on the first passage, mesenchymal stem cells were cryopreserved on MVE 800 Series Cryo Tank with temperature -190°C.

After thawing, subculture were performed until passage 3. Population Doubling Time (PDT) and Flow cytometry analysis were taken part. PDT was calculated with formula $t/n; n = \log(N_h) - \log(N_i) / \log 2$, where N_h : total cell count at harvest, N_i : total cell at first, t : duration of culture, and n : population doubling time. In Flow cytometry analysis, samples were then incubated with fluorescein isothiocyanate (FITC) that conjugated with anti-monoclonal CD34, CD73, CD 105, and CD90. Cells were analyzed using BD FACSCalibur flow cytometer (BD Biosciences).

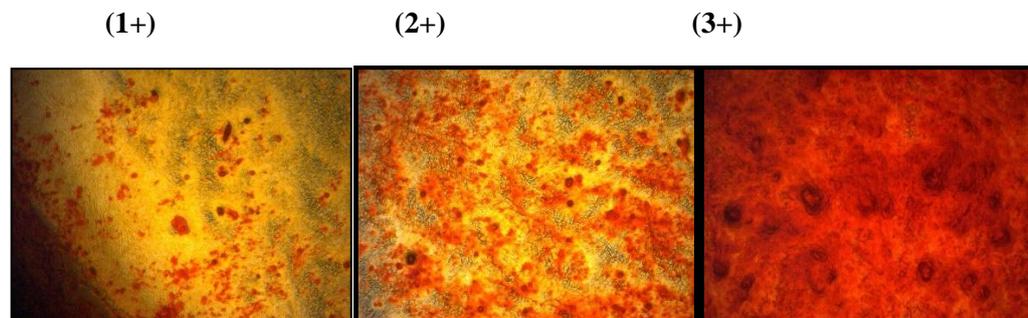


Figure 1. Alizarin Red assay assessment criteria.

Osteogenic differentiation was carried out in 8 mL DMEM, using StemPro® Osteogenesis Differentiation Kit (Gibco) in 37°C, 20% O₂, 5% CO₂ for 21 days. Alizarin Red Assay was performed. Cell count was done in 5 high power field (HPF) for 2 times by author, and was interpreted as such: (1+) if $< 1/3$ HPF was enhanced; (2+) if $1/3 - 2/3$ hpf was enhanced; and (3+) if $> 2/3$ hpf was enhanced (figure 1). Data analysis was done using Statistical Product and Service Solution (SPSS) version 20.0 program.

Results

The mean age of subjects was $34,92 \pm 8.15$ years old. Most of the subjects were male (83,33%) and harvested from the hip adipose. Characterization result of surface biomarker CD34, CD73, CD90, and CD105 showed no significant differences among various age groups. ($p > 0.05$, 95% CI) (figure 2).

Population Doubling Time as evaluation of growth pace was conducted. Mann-Whitney Test showed significant difference on group 2 (31-40 year old)($p < 0.05$, 95%CI) and group 3 (41-55 year old) ($p < 0.05$, 95%CI) as shown in Figure 3. Population Doubling Time was also analyzed based on the origin of source, bone marrow and adipose tissue. On bone marrow derived mesenchymal stem cells, post hoc analysis with Bonferroni multiple comparisons showed significant difference between group 1 (15-30 year old) and group 3 (41-55 year old) ($p = 0.048$, 95%CI). On adipose tissue, one-way Anova test showed no significant difference among the age group ($P > 0.05$, 95%CI) as shown in Figure 4.

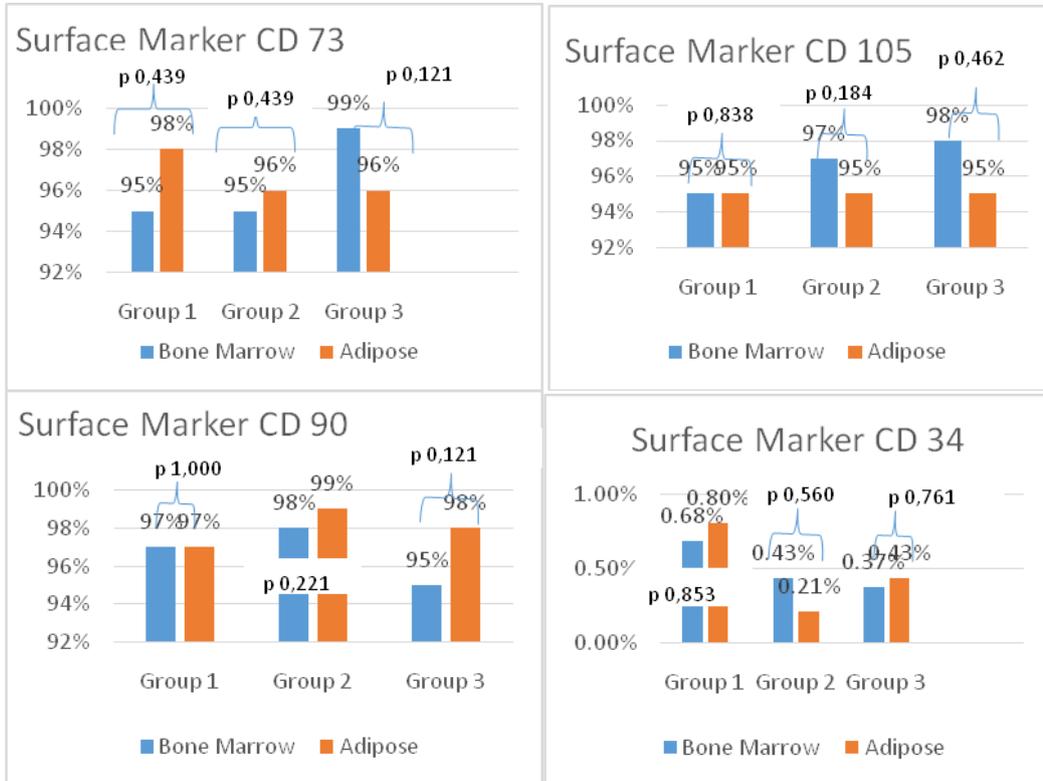


Figure 2. Surface Marker CD 73, CD 105, CD 90, CD 34 evaluation

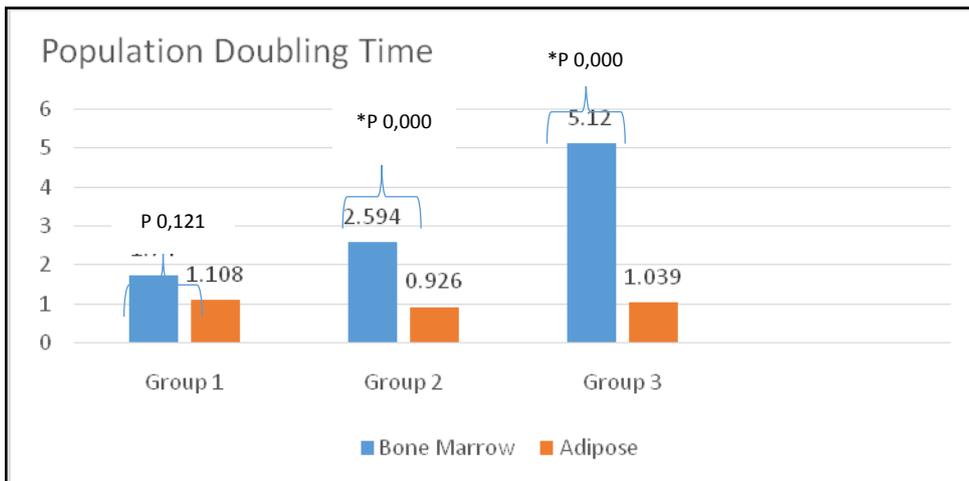


Figure 3. Population Doubling Time evaluation in various age group

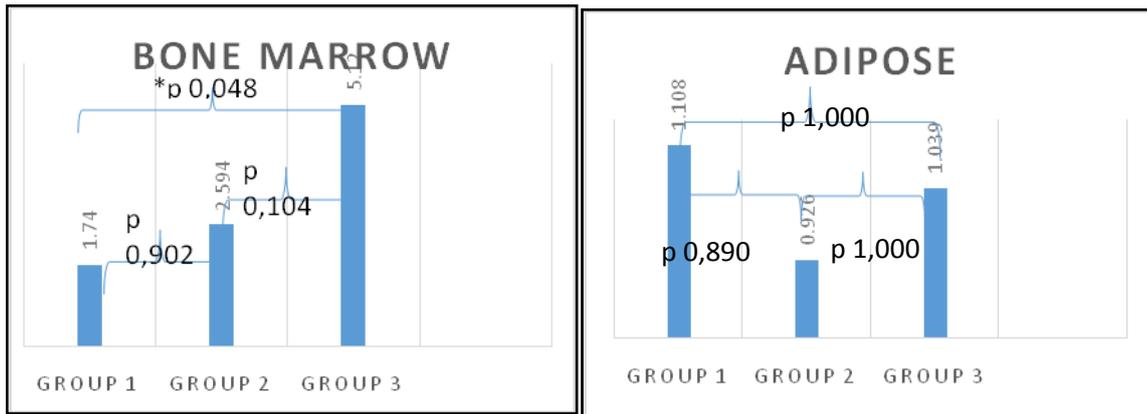
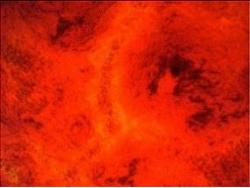
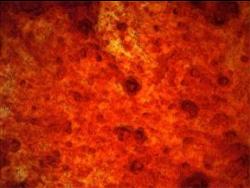


Figure 4. Population Doubling Time evaluation in various sources

Alizarin Red assay was performed to evaluate extra cellular calcification which signified osteogenic differentiation. Kruskal-Wallis test showed no significant difference was found in osteogenic differentiation among various age groups. ($p > 0.05$, 95%CI) (Table 1).

Table 1. Alizarin Red assay evaluation in various age group

	Group 1	Group 2	Group 3
Bone Marrow	 3+	 2+	 1+
Adipose	 3+	 3+	 3+
P value	1,000	0,083	0,083

Discussion

This study was an observational analytical *in vitro* methods with 12 samples of mesenchymal stem cells derived from bone marrow and adipose tissue in various age groups. Subjects was divided into three age groups: 15-30 years old, 31-40 years old, and 41-55 years old. Stolzing et al stated that in >40 years old people, bone marrow derived mesenchymal stem cell had decreased proliferating and differentiating potency.¹⁰ Another studies stated that PDT on adipose derived mesenchymal stem cell was significantly different at >40 years old¹¹ and the amount of apoptoses increased by age.¹²

Bone marrow was selected due to the widely accepted and gold standard source. Indeed, it had several weaknesses, including less number of cell and declining proliferating and differentiating ability by age.^{1,7} Because of abundant sources, less invasive procedure, and relative more cell, adipose will be a promising source with equal regenerating and differentiating ability.¹³

All of the subjects showed homogenous distribution in the range of 19-44 years old. Bone marrow was harvested from posterior illiac crest which was thought to have the largest amount of bone marrow and reproducibile result. While the location of the adipose tissue derived largely from the hip. Subjects were mainly man to minimize decreasing bone marrow differentiation capability in perimenopause age.

All MSC samples was cryopreservated at passage 1. This was conducted because bone marrow and adipose tissue samples were obtained in different time over a period of 3 months. Furthermore, cryopreservation was aimed to standardized the process for each sample. Minonzio et al showed that cryopreservation until 193 days didn't changed the amount, characteristic, viability, and proliferation potency of MSC.¹⁴

Flowcytometry was carried out to compare the surface marker CD 73, CD 105, CD 90 and CD 34 on the samples at passage 3. All samples had fulfilled minimal criteria of MSC^{7,15} and did not have significant difference among age group. This was equal with Choudhery et al that stated the age did not affect the difference of morphology and phenotype of MSC from adipose tissue and Labora et al that stated there was no significant difference in MSC surface marker by age.^{8,16}

PDT of bone marrow and adipose MSCs was compared among age group. In 31-40 year old group and 41-55 year old group, bone marrow and adipose tissue derived MSC was significantly different. This was equal with Choudhery et al, Schipper et al, and Labora et al that showed the age did not affect proliferation rate of adipose MSC, in contrast to bone marrow.^{8,12,16} This study showed a significant decrease in proliferation rate of MSC derived bone marrow along with age, similar to Nakahara et al.¹⁷ Another study showed Vincolin gene that was expressed on low quantities at a young age and increasing with age also affect the rate of proliferation.¹⁶ Bujalska et al showed that the enzyme activity of hydroxysteroid 11 β -dehydrogenase type 1 (11 β -HSD1) was responsible for the decline in the proliferation on elderly that the enzyme had a role in producing glucocorticoid in the cell. Transforming growth factor β 1 (TGF- β 1) induced aging on MSC derived bone marrow by increasing production of reactive oxygen species (ROS) from mitochondria, resulting in a decrease in mitochondrial membrane potential, the destruction of DNA, and cell aging.^{10,18} This was explained Labora et al where TGF- β 1 and ROS increased significantly in elderly.¹⁶ This study showed that in elderly, MSCs are better harvested from adipose tissue that has a PDT constantly in every age.

All samples showed enhancement in Alizarin Red assay, so osteogenic differentiation was confirmed. Author created a semi quantitative assessment of osteogenic differentiation in Alizarin Red assay. This assessment showed no significant difference of extracellular calcification among age group. There was roughly a difference in enhancement but it was not statistically different. Zhu et al stated there age did not significantly affect osteogenesis in adipose tissue derived MSC but Choudhery et al found a contradiction. It was still a debatable thing, and will further need another osteogenic marker. Estrogen was thought to influence the osteogenic differentiation in elderly. In this study, there was no woman subject in perimenopausal age so it was neglectable.

Conclusion

There was no difference on biological characteristic of bone marrow and adipose tissue MSC in every age group. There was significant difference in PDT between bone marrow and adipose MSC in 31-40 year old and 41-55 year old group. No difference in osteogenic differentiation and Alizarin Red assay was found between bone marrow and adipose MSC in each age group. Adipose MSC had steady biological characteristic, osteogenic differentiation, and Population Doubling Time in each age group. Meanwhile, bone marrow MSC had increasing population doubling time in increasing age, apart from biological characteristic and osteogenic differentiation that constant at any age. Adipose could be the source of choice in harvesting mesenchymal stem cell at any age.

References

1. Hu L, Hu J, Zhao J, Liu J, Ouyang W, Yang C, et al. Side-by-Side Comparison of the Biological Characteristics of Human Umbilical Cord and Adipose Tissue-Derived Mesenchymal Stem Cells. 2013;2013.
2. Pawitan JA. Prospect of Adipose Tissue Derived Mesenchymal Stem Cells in Regenerative Medicine. Cell Tissue Transplant Ther. 2009;(2):7-9.
3. Lubis AM, Lubis VK. Adult bone marrow stem cells in cartilage therapy. Acta Med Indones. 2012;44(1):62-8.
4. Sachs PC, Francis MP, Zhao M, Brumelle J, Rao RR, Elmore LW, et al. Defining essential stem cell characteristics in adipose-derived stromal cells extracted from distinct anatomical sites. Cell Tissue

- Res. 2012;349(2):505–15.
5. Jin H, Bae Y, Kim M, Kwon S-J, Jeon H, Choi S, et al. Comparative Analysis of Human Mesenchymal Stem Cells from Bone Marrow, Adipose Tissue, and Umbilical Cord Blood as Sources of Cell Therapy. *Int J Mol Sci.* 2013;14(9):17986–8001.
 6. Kornicka K, Marycz K, Tomaszewski KA, Mar M, Agnieszka U. The Effect of Age on Osteogenic and Adipogenic Differentiation Potential of Human Adipose Derived Stromal Stem Cells (hASCs) and the Impact of Stress Factors in the Course of the Differentiation Process. 2015;2015.
 7. Lubis AMT, Sandhow L, Lubis VK, Noor A, Gumay F, Merlina M, et al. Isolation and cultivation of mesenchymal stem cells from iliac crest bone marrow for further cartilage defect management. *Acta Med Indones.* 2011;43:178–84.
 8. Choudhery MS, Badowski M, Muise A, Pierce J, Harris DT. Donor age negatively impacts adipose tissue-derived mesenchymal stem cell expansion and differentiation. *J Transl Med.* 2014;12:8.
 9. Ulrich M, Peter wiesmann hans. *Bone and Cartilage Engineering.* 2006. 73-78 p.
 10. Yu KR, Kang KS. Aging-related genes in mesenchymal stem cells: A mini-review. *Gerontology.* 2013;59(6):557–63.
 11. Zhu M, Kohan E, Bradley J, Hedrick M, Benhaim P, Zuk P. The effect of age on osteogenic , adipogenic and proliferative potential of female adipose-derived stem cells. 2009;(3):290–301.
 12. Cells HAS, Rubin JP. Regional Anatomic and Age Effects on Cell Function of. 2008;60(5):538–44.
 13. Guofang Zeng^{1,†}, Kuan Lai^{2,†}, Jin Li¹, Yaqin Zou¹, Haili Huang³, Jie Liang¹, Xudong Tang⁴, Jing Wei⁵, and Peihua Zhang¹. A rapid and efficient method for primary culture of human adipose-derived stem cells. *Organogenesis.* 2013;9(4):286–95.
 14. Minonzio G, Corazza M, Mariotta L, Gola M, Zanzi M, Gandolfi E, et al. Frozen adipose-derived mesenchymal stem cells maintain high capability to grow and differentiate. *Cryobiology.* Elsevier Inc.; 2014;69(2):211–6.
 15. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, et al. Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy.* 2006;8(4):315–7.
 16. Fafián-Labora J, Fernández-Pernas P, Fuentes I, De Toro J, Oreiro N, Sangiao-Alvarellos S, et al. Influence of age on rat bone-marrow mesenchymal stem cells potential. *Sci Rep.* 2015;5:16765.
 17. Nakahara M, Takagi M, Hattori T, Wakitani S, Yoshida T. Effect of subcultivation of human bone marrow mesenchymal stem on their capacities for chondrogenesis, supporting hematopoiesis, and telomea length. *Cytotechnology.* 2005;47(1-3):19–27.
 18. Kasper G, Mao L, Geissler S, Draycheva A, Trippens J, Kühnisch J, et al. Insights into mesenchymal stem cell aging: Involvement of antioxidant defense and actin cytoskeleton. *Stem Cells.* 2009;27(6):1288–97.
