

Growth Promotion of indigenous *Scenedesmus dimorphus* strain under different conditions using stirred tank photobioreactor

Okkou H¹, Naddaf M², Alinizam A³, Azmeh M F¹

¹Department of Biodiversity, National Commission for Biotechnology, P.O.Box 31902, Damascus, Syria.

²Department of Food Science, agriculture faculty, Tishreen University, Lattakia, Syria.

³Department of Plant Biology, Science Faculty, Damascus University, Damascus, Syria

Abstract: The effects of culture conditions at different temperature (15-25-35°C) and nitrate concentration (0.1-0.25-0.4g/L) on biomass production and growth rate of *scenedesmus dimorphus* were determined. The results shown that the dry biomass weight (0.153g/L) at 15°C was the lowest. No significant difference was observed between the dry biomasses at 25°C (0.289g/L) and 35°C (0.240g/L). While the lowest growth rate was at 15°C (0.157d⁻¹), the highestone was at 25°C (0.229d⁻¹) and 35°C (0.195d⁻¹) with insignificant deference, and The nitrate concentration had no effect on both of biomass production and growth rate.

Key words: *Scenedesmus dimorphus*, biomass, growth rate, Photobioreactor.

Introduction

Microalgae are a group of fast growing unicellular or simple multicellular microorganisms¹ which have several advantages to consider as energy crop, including higher photosynthetic efficiency, higher growth rate and higher biomass production². They are promising for economic industrial-scale production in the 21st century. Microalgae can be utilized in the production of nutritional supplements, antioxidants, cosmetics, natural dyes³. They can also be used in gum, beverages, candy and snack foods⁴.

Among microalgae, *Scenedesmus* genus has the most desirable features for efficient and economic combination of CO₂ fixation, wastewater treatment, and lipid synthesis toward biodiesel production^{5,6}. Recently, it has drawn attention as commercially a valuable source for a wide spectrum of compounds such as; high quality protein, B₁₂, C and E vitamins, pigments and other bioactive chemicals, long chain polyunsaturated fatty acids especially "omega-3" and "omega-6", glycolipids, sulfolipids and phenolic compounds^{7,8,9}. *Scenedesmus dimorphus* seems to be the ideal species, due to its good ability to grow and tolerate different environmental conditions¹⁰.

Recently, many studies reported various cultivation technologies to produce microalgae¹¹. Besides open ponds, the primary and effective methods for the large-scale production^{3, 12}, photobioreactors offered a better opportunity for controlling and optimizing all cell growth parameters to meet the specific demands^{13, 14}. The most important and common design of this reactor is Stirred-tank reactors (STR)¹⁵, for being simple in operation and structure, and ideal for cultivating different types of cells¹⁶.

The main environmental factors influencing microalga growth and its chemical composition are light, nutrients, temperature and pH¹⁷.

So, several strategies have been applied to improve the growth, biomass production and lipid content, by optimization of the medium composition (e.g., carbon source, nitrogen, phosphorus, vitamins and salts)¹⁸, and physical parameters (e.g., pH, temperature and light intensity)¹⁹.

Temperature strongly influences the growth rates for all algae species²⁰, and for *Scenedesmus sp.*, the optimum temperature to grow is between 20 and 40°C²¹, but the optimal one to produce biomass and lipids was 20°C²². Cassidy (2011) measured *Scenedesmus* and *Chlorella's* growth rate at different temperatures (25, 30 or 35°C), and found that 30°C had the best growth rates for both algae.

Nitrogen was quantitatively the most important nutrient affecting the biomass and lipid productivity²³. Therefore, it is important to utilize an appropriate nitrogen source at a certain concentration²⁴, so many studies investigated the effect of both nitrogen source and its concentration on biomass growth. According to Arumugam *et al.* (2013) nitrate was found to be a preferred form of nitrogen source, that potassium nitrate (0.32 g/L) and sodium nitrate (0.28 g/L) were the best for biomass growth of *Scenedesmus sp.* For *Scenedesmus dimorphus* species, the maximum growth rate in terms of biomass and lipid productivity was at 0.1 g/L urea, but the biomass and cell numbers decreased gradually by increasing urea concentration beyond 0.1 g/L².

The aim of this study was to evaluate the effect of temperature and nitrogen at various levels on *Scenedesmus dimorphus* growth using Photobioreactor to determine the best conditions for cell growth and biomass production.

Materials and methods

Microalgae and culture conditions

The experimental organism *scenedesmus dimorphus* was isolated from fresh water ponds located in Quneitra Province (south of Syria). The algae broth medium from sigma co., solidified with agar, was used to isolate and purify the algae in Petri-Dishes, then species was identified at the Center of Advanced Studies in Botany, University of Madras, Chennai, Egypt.

After the inoculum was prepared, the initial cell concentration was set to be its OD₆₈₀ = 0.150 (optical density at 680nm). All experiments were carried out in the Stirred Tank (STR) with a maximum capacity of 14L and 10L as working volume figure (1).

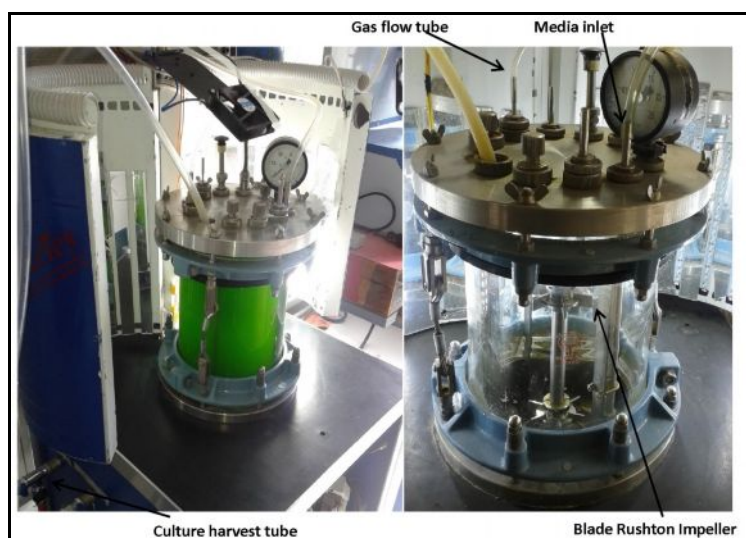


Fig. 1: The modified Stirred Tank Reactor

STR was equipped with PLC (Programmable Logic Controller), controlling and monitoring all parameters (Temperature, light, pH, gas flow; CO₂ and air, mixing speed).

All Growth experiments were done at three different temperatures (15, 25, 35°C), then three different nitrate concentrations (0.1, 0.25, 0.4g/l) in 10 L autoclaved Bissch off & Bold medium (BBM). Each batch cultivation was carried out three times for 15 days at fixed parameters (6000Lux continuous illumination, 250 rpm mixing speed, PH= 7, air mixed with CO₂ 500ppm with flow rate 2L/min, temperature 25°C with variable nitrate concentration, and nitrate concentration 0.25g/L at variable temperature) all parameters were established on the basis of the last studies^{26,27}. The central values of temperature, 25°C were chosen depending on Hernandez *et al.* (2009) and Brown *et al.* (1998). But the additional experiments were done either increasing or reducing the growth temperature by 5°C. The central concentration of nitrogen in medium (0.25 g/ L) was selected according to Guillard (1975), and the additional cultivations were run at 0.1 and 0.4g/L.

Estimation of Algal Growth

The growth rate (GR) was calculated by fitting OD₆₈₀ of the daily samples using spectrophotometer (HITACHI U-2900) in the following formula²⁹:

$$GR = (\ln OD_t - \ln OD_0) / t.$$

OD₀: the optical density at inoculation day. OD_t: the optical density measured on day t.

Determination of Biomass Dry Weight (DW)

Dry Biomass content was determined according to the modified method of Yadavalli *et al* (2012), that used a standard curve of OD₆₈₀ plotting versus serial dilutions of DW figure (2).

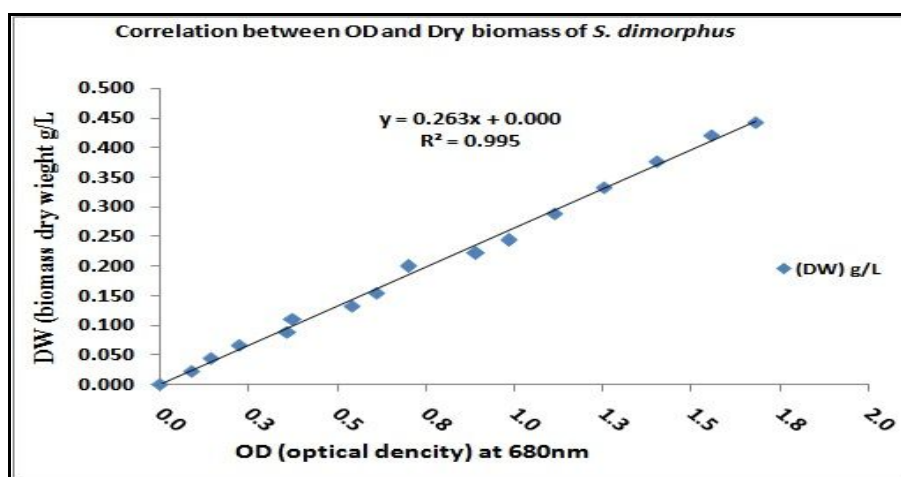


Fig. 2 Linear regression equation of relationship between OD and dry biomass

The algal cells were collected by centrifugation (5,000×g, 10 min), and washed with water, then dried for determining its DW gravimetrically.

All statistical analyses were done using SPSS (version 17), according to ANOVA followed by Tukey test.

Results and discussion

Effect of temperature:

Biomass production

The daily increase in dry biomass at the three temperatures showed that the actual increase in dry biomass began after the second day of inoculation at all temperatures, figure (3). This

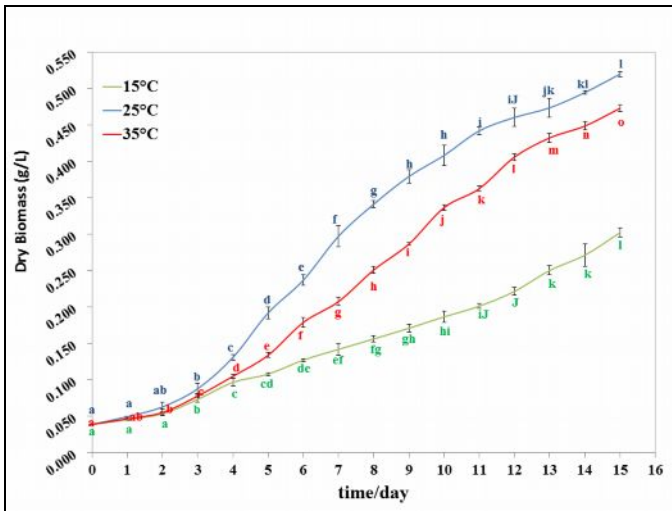


Fig. 3: dry biomass curve grown in different temperatures. Different letters above the lines indicate significant differences ($p < 0.001$) among means within each temperature. (ANOVA followed by Tukey Test).

Suggests that the studied temperature had no effect during the lag phase. As that the Exponential phase started in the third day of inoculation, and continued with a significant increase ($P > 0.001$), in most days, till day 15; the harvest day, but at 25°C, the difference between the fourteenth and fifteenth days wasn't significant, this may make the fourteenth day more economical to harvest. This could refer to the acceleration of growth at 25, that led to shortcut the exponential phase. This result was lower than that of Velichkova *et al* (2013), who clarified that the sixteen day was the harvest day. On the other hand, the effect of temperature in table (1) shown that the difference between the means of biomass at 25°C and

Growth rate

The daily growth rate for the three temperatures is shown in Figure (4), where the growth rate

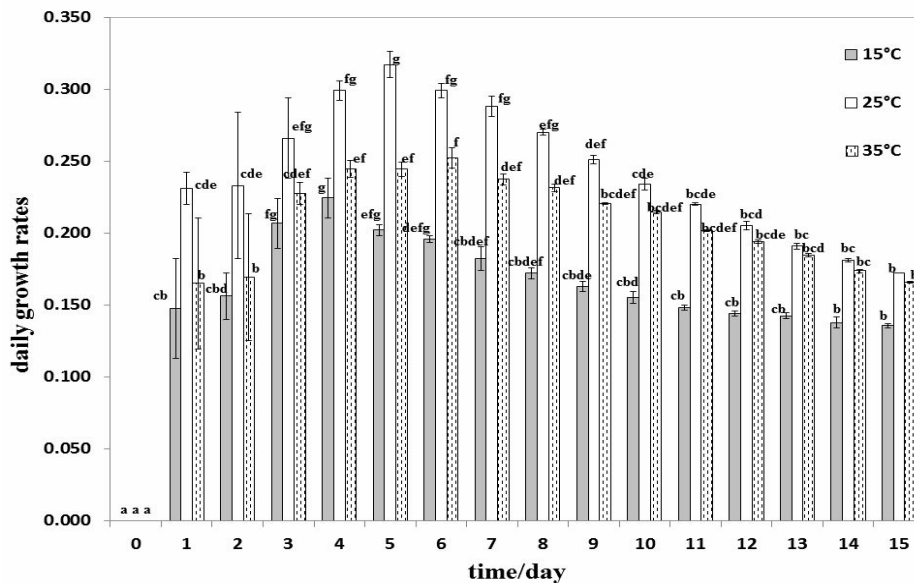


Fig. 4: Comparison of daily growth rate grown in different temperatures. Different letters above the bars indicate significant differences ($p < 0.001$) among means within each temperature. (ANOVA followed by Tukey Test).

Table 1: Means \pm SD of dry biomasses and growth rate during the entire growth period within three temperatures

Temperature	dry biomass \pm SD		Growth rate/day \pm SD	
15°C	0.227 \pm 0.150	^a 0.153 \pm 0.080	^a 0.157 \pm 0.050	0.079 \pm 0.041
25°C		^b 0.289 \pm 0.171	^b 0.229 \pm 0.075	
35°C		^b 0.240 \pm 0.152	^b 0.195 \pm 0.060	

Different letter in each Column for each species indicate significant differences ($p < 0.001$) between means (ANOVA followed by Tukey Test) 35°C was not significant. X *et al.* (2011) showed that the temperature 20°C gave the highest biomass in *Scenedesmus sp.*, so it is possible that the range 20°C to 35°C is the best to get the highest dry biomass.

Increased up to the highest values 0.224, 0.317 and 0.252 d⁻¹ in days fourth, fifth and sixth at 15°C, 25°C and 35°C respectively, that indicated to a positive relation between temperature (15 to 35°C) and time elapsed to achieve the highest growth rate. This seemed to be logical, as the lower temperature reduces the length of the exponential Phase.

According to the table (1), the lowest growth rate was at 15°C 0.157 \pm 0.050d⁻¹, while the difference between the growth rates at 25°C and 35°C, respectively, was not significant.

This agreed with other studies, as Westerhoff *et al.* (2010) found that growth rates did not vary by temperatures ranging between 27 and 39°C. Also, Xin *et al.* (2011) showed that the highest growth rate was obtained at 25°C. In addition, Sanchez *et al.* (2008) explained that the ideal growth of *Scenedesmus sp.* was within the range of temperature 20 to 40°C.

Effect of nitrate concentrations:

Biomass production

The actual increase of biomass, at concentrations 0.4 and 0.25g/L, began after two days of inoculation, while it delayed till the third day for the concentration 0.1g/L Figure (5). That

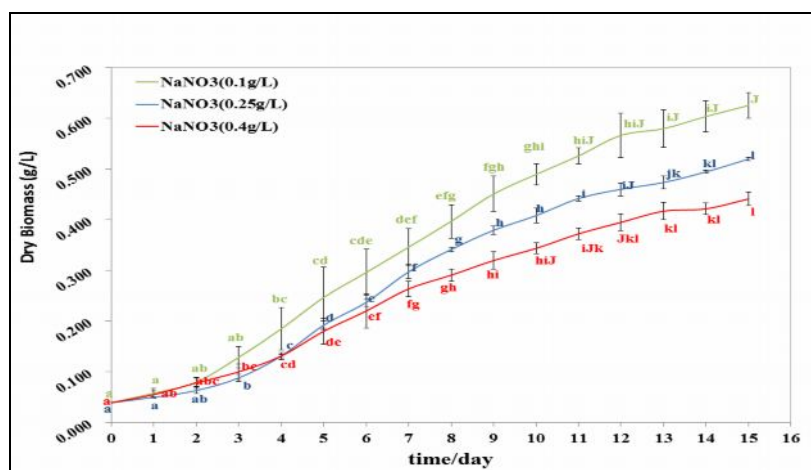


Fig. 5: The change of dry biomass by the time, in different concentrations of sodium nitrate. Different letters above the lines indicate to significant differences ($p < 0.001$) among means within each concentration. (ANOVA followed by Tukey Test). might be explained by elongation of the Lag phase, that algae need to adapt with the growth medium according to Spencer (1954), at the low concentration of nitrate.

During the Exponential phase, the dry biomass increased significantly by the time in all concentrations, in most days. This is consistent with Becker (1994), who explained the systematic duplication of the cells in this

phase. In the table (2), the insignificant differences ($P > 0.001$) in dry biomass values might be referred to that the concentration of nitrates

Table 2: Means \pm SD of dry biomasses and growth rate during the entire growth period within three concentrations of sodium nitrate

NaNO ₃ (g/L)	dry weight \pm SD		Growth rate \pm SD	
0.1	^a 0.298 \pm 0.177	^a 0.351 \pm 0.205	^a 0.272 \pm 0.106	^a 0.113 \pm 0.047
0.25		^a 0.289 \pm 0.171	^a 0.229 \pm 0.075	
0.4		^a 0.254 \pm 0.139	^a 0.233 \pm 0.090	

Different letter in each Column for each species indicate significant differences ($p < 0.001$) between means (ANOVA followed by Tukey Test) within the range (0.1g/L to 0.4g/L) did not have any effect on the produced dry biomass. While in the study carried out by Arumugam *et al.* (2013) that tested four different concentrations of sodium nitrates 05, 10, 15 and 20 mM, the best one was (10Mm=0.28g/L). This could be attributed to the effects of other condition on growth.

Growth rate:

The daily growth rate at different concentrations of sodium nitrate shown in Figure (6), where the highest growth rates for concentrations 0.1, 0.25, 0.4g/L were at days third, fifth and second respectively.

Also, according to table (2) the increase of sodium nitrate concentration from 0.1g/L to 0.4g/L had no effect on growth rate of algae. This result correspond with Goswami and Kalita (2011) who used urea as a nitrate source, since the highest growth rate achieved at 0.1g/L, then the more increase in the urea concentration led to a gradual decline in growth.

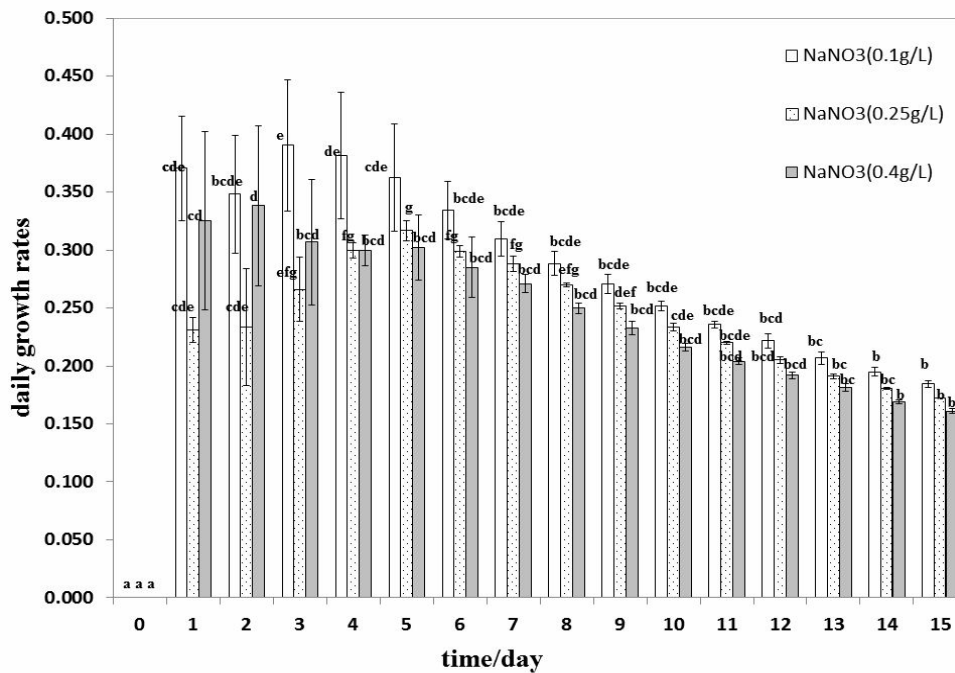


Fig. 6: Comparison of daily growth rate grown in different concentrations of sodium nitrate. Different letters above the bars indicate to significant differences ($p < 0.001$) among means within each concentration. (ANOVA followed by Tukey Test).

Conclusion

Because of *Scenedesmus dimorphus* is used in food and pharmaceutical industries, the objective of this research was to optimize the best culture conditions for the highest biomass production. It has been mentioned above that 25 and 35°C as a growth temperature, achieved the optimal biomass yield and the highest growth rate, too. In addition, biomass and growth rate of *S. dimorphus* was not significantly influenced by nitrate concentration, so any increase in nitrate concentration above 0.1g/L was useless and not economic. Therefore, we recommended culturing *S. dimorphus* at 25-35°C and 0.1g/L NaNO₃ to get the optimal production of its biomass.

Acknowledgements

We are grateful to Syrian National Commission for Biotechnology (NCBT Damascus, Syria and Science Faculty, Department of Plant Biology Damascus, Syria and Department of Food Science, agriculture faculty, Tishreen University, Lattakia Syria.

References

1. Wang, B., Li, Y., Wu, N. and Lan, C.Q., CO₂ bio-mitigation using microalgae. *Appl. Microbiol. Biotechnol.*, 2008, 79, 5, 707-718.
2. Goswami, R. and Kalita, M., *Scenedesmus dimorphus* and *Scenedesmus quadricauda*: two potent indigenous microalgae strains for biomass production and CO₂ mitigation - A study on their growth behavior and lipid productivity under different concentration of urea as nitrogen source. *Journal of Algal Biomass Utilization*, 2011, 2: 42- 49.
3. Rosenberg, J.N., Oyler, G.A., Wilkinson, L. and Betenbaugh, M.J., A green light for engineered algae: Redirecting metabolism to fuel a biotechnology revolution. *Biotechnol*, 2008, 19: 430-6.
4. Spolaore, P., Joannis-cassan, C., Duran, E. and Isambert, A., Commercial applications of microalgae, *Journal of Bioscience and Bioengineering*, 2006, 101, 2, 87-96.
5. Xin, L., Ying, H.H., Ke, G. and Xue, S.Y., Effects of Different Nitrogen and Phosphorus Concentrations on the Growth, Nutrient Uptake, and Lipid Accumulation of a Freshwater Microalga *Scenedesmus* sp. *Bioresource Technology*, 2010, 101, 5494-5500.
6. Bliersch D.M., Kangas P.C. and Mulbry W.W., Turbulence and Nutrient Interactions That Control Benthic Algal Production in an Engineered Cultivation Raceway, *Algal Research journal*, 2013, 2, 2, 107-112.
7. Herrero, M., Cifuentes, A. and Ibanez, E., Supercritical fluid extraction of functional ingredients from different natural sources: Plants, food-by-products, algae and microalgae A review. *Food Chemistry*, 2006, 98, 136-148.
8. Abd El-Baky H.H. and El-Baroty G.S., Characterization and bioactivity of phycocyanin isolated from *Spirulina maxima* grown under salt stress, *Food & function Journal*, 2012, 3, 4, 381-388.
9. Guedes, A.C., Catarina, R.B., Helena, M.A., Pereira, C.I. and Francisco, X. M., Microalgal and cyanobacterial cell extracts for use as natural antibacterial additives against food pathogens. *Inter. J. Food Sci. Technol.*, 2011, 46, 862-870.
10. Yaakob, Z., Kamarudin, K. F., Rajkumar, R., Takriff, M. S. and Badar, S. N., the Current Methods for the Biomass Production of the Microalgae from Wastewaters: An Overview. *World Applied Sciences Journal*, 2014, 31, 1744-1758.
11. Olaizola, M., Commercial development of microalgal biotechnology: from the test tube to the marketplace. *Biomol Eng.*, 2003, 20, 459-66.
12. Chisti, Y., Biodiesel from microalgae, *Journal of Biotechnology Advances*, 2007, 25, 294-306.
13. Lebeau, T. and Robert, J.M., Diatom cultivation and biotechnologically relevant products. Part I: Cultivation at various scales. *Microbiol Biotechnol.*, 2003, 60, 612-623.
14. Sato, T., Usui, S., Tsuchiya, Y. and Kondo, Y., Invention of outdoor closed type photobioreactor for microalgae. *Energ Convers Manage.*, 2006, 47, 791-799.
15. Sacasa-Castellanos, C., Batch and continuous studies of *Chlorella vulgaris* in photo-bioreactors. Thesis format: The University of Western Ontario, 2013, 67p.

16. Yang, J. and Wang, N.S., Cell inactivation in the presence of sparging and mechanical agitation. *Biotechnol. Bioeng.* 1992, 40, 806-816.
17. Rousch, J.M., Bingham, S.E. and Sommaerfeld, M.R., Change in fatty acid profiles of thermo-intolerent and thermo tolerant marine diatoms during temperature stress, *J. Exp. Mar. Biol. Ecol.*, 2003, 295, 145-156.
18. Mata, T.M., Martins, A.A. and Caetano, N.S., Microalgae for Biodiesel Production and Other Applications: A Review, *Journal of Renewable and Sustainable Energy Reviews*, 2010, 14, 217-232.
19. Rawat, I., Ranjith Kumar, R., Mutanda, T. and Bux, F., Biodiesel from Microalgae: A Critical Evaluation from Laboratory to Large Scale Production. *Applied Energy*, 2013, 103, 444-467.
20. Cassidy, K. O., Evaluating Algal Growth at Different Temperatures, Kentucky, United States: University of Kentucky, MSc thesis, 2011.
21. Sanchez, J. F., Fernandez-Sevilla, J. M., Acien, F. G., Ceron, M. C., Perez-Parra, J., and Molina-Grima, E., Biomass and lutein productivity of *Scenedesmus almeriensis*: influence of irradiance, dilution rate and temperature, *Applied Microbiology and Biotechnology*, 2008, 79, 719-729.
22. Xin, L., Hong-ying, H. and Yu-ping, Z., Growth and lipid accumulation properties of a freshwater microalga-*Scenedesmus* sp. under different cultivation temperature, *Bioresource Technology*, 2011, 102, 3098–3102.
23. Griffiths, M.J. and Harrison, S.T.L., Lipid productivity as a key characteristic for choosing algal species for biodiesel production, *J. Appl. Phycol.*, 2009, 21, 493–507.
24. Yeh, K.L. and Chang, J.S., Effects of Cultivation Conditions and Media Composition on Cell Growth and Lipid Productivity of Indigenous Microalga *Chlorella vulgaris* ESP-31, *Bioresource Technology*, 2012, 105, 120-127.
25. Arumugam M., Agarwal A., Arya M.C. and Ahmed, Z., Influence of nitrogen sources on biomass productivity of microalgae *Scenedesmus bijugatus*, *Bioresource Technoloy*, 2013, 131, 246–249.
26. Guillard R.R.L., Culture of phytoplankton for feeding marine invertebrates, in: W.L. Smith, M.H. Chanley (Eds.), *Culture of Marine Invertebrate Animals*, Plenum Press, New York. 1975, 26–60.
27. Brown M.R., McCausland M.A. and Kowalski K., The nutritional value of four Australian microalga strains fed to Pacific oyster *Crassostrea gigas*, *Journal of Aquaculture*, 1998, 165, 281–293.
28. Hernandez J.P., de-Bashan L.E., Rodriguez D.J., Rodriguez Y. and Bashan Y., Growth promotion of the freshwater microalga *Chlorella vulgaris* by the nitrogen-fixing, plant growth-promoting bacterium *Bacillus pumilus* from arid zone soils, *Eur. J. Soil Biol.*, 2009, 45, 88–93.
29. Hasan R., Zhang B., Wang L. and Shahbazi A., Bioremediation of Swine Wastewater and Biofuel Potential by using *Chlorella vulgaris*, *Chlamydomonas reinhardtii*, and *Chlamydomonas debaryana*, *Journal of Petroleum and Environmental Biotechnology*, 2014, 3, 175-180.
30. Yadavalli R., Rao S.R. and Rao C.S., Lipid accumulation studies in *Chlorella pyrenoidosa* using customized photobioreactor- effect of nitrogen source, light intensity and mode of operation. *International Journal of Engineering Research and Applications*, 2012, 2, 2446-2453.
31. Velichkova K., Sirakov I. and Georgiev G., Cultivation of *Scenedesmus dimorphus* strain for biofuel production, *Agricultural science and technology*, 2013, 5, 181-185.
32. Westerhoff P., Hu Q., Esparza-Soto M. and Vermaas W., Growth parameters of microalgae tolerant to high levels of carbon dioxide in batch and continuous-flow photobioreactors. *Environmental Technology*, 2010, 31, 5, 523-U580.
33. Spencer C. P., Studies on the culture of a marine diatom, *J. mar. biol. Ass. U.K.*, 1954, 33, 265-290.
34. Becker, E.W., *Microalgae: Biotechnology and Microbiology*, Cambridge, Cambridge University Press, 1994.
